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Biomethanation of Carbon Monoxide by Hyperthermophilic Artificial Archaeal Co-Cultures

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Abstract: Climate neutral and sustainable energy sources will play a key role in future energy production. Biomethanation by gas to gas conversion of flue gases is one option with regard to renewable energy production. Here, we performed the conversion of synthetic carbon monoxide (CO)-containing flue gases to methane (CH₄) by artificial hyperthermophilic archaeal co-cultures, consisting of *Thermococcus onnurineus* and *Methanocaldococcus jannaschii*, *Methanocaldococcus vulcanius*, or *Methanocaldococcus villosus*. Experiments using both chemically defined and complex media were performed in closed batch setups. Up to 10 mol% CH₄ was produced by converting pure CO or synthetic CO-containing industrial waste gases at a high rate using a co-culture of *T. onnurineus* and *M. villosus*. These findings are a proof of principle and advance the fields of Archaea Biotechnology, artificial microbial ecosystem design and engineering, industrial waste-gas recycling, and biomethanation.

Keywords: Archaea Biotechnology; anaerobic microbiology; methanogenesis; biohydrogen; biological gas conversion



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1. Introduction

In the European Union, around 70% of primary energy is generated by the combustion of fossil fuels, contributing about 78% (3367 Tg-CO₂ equ.) of the total emitted greenhouse gases [1,2]. A transition to a carbon dioxide (CO₂)-neutral and sustainable energy production system is urgently needed. One of the possibilities is to utilize the power-to-gas process [3–5]. Within this process, biomethanation of CO₂ to methane (CH₄) offers a sustainable opportunity to enable the transition from fossil fuels, as it is an autocatalytic process. Therefore, it is envisioned that biomethanation will become an essential part of future energy production systems, as CH₄ could be produced at a stable pace and stored in vast amounts in the natural gas grid network [6]. Pure cultures of methanogenic archaea (methanogens) [7–12] and enrichment cultures containing methanogens [13–17] can be utilized for in situ or ex situ biomethanation [18–20].

Carbon monoxide (CO)-containing rich waste gases are a by-product of industrial processes such as steelmaking [21]. CO-containing gases can also be obtained by gasification of carbon-rich materials, such as domestic organic waste or lignocellulose conversion to syngas [22]. The fact that biofuel production directly from lignocellulose is still costly and biotechnologically challenging makes microbial gas to gas conversion from CO a promising alternative bioprocess [7,23]. Developments in the transformation of gaseous waste products to energetically valuable compounds emphasize the potential for syngas as a substrate [24–27]. CO has a high potential for donating electrons, making it a favourable

substrate for lithotrophic microorganisms [28,29]. Besides CO, waste and syngas mainly consist of molecular hydrogen (H₂), CO₂, and CH₄. As early as 1990, it was demonstrated that methanogens can metabolize some components from syngas [30].

The direct conversion of pure CO to CH₄ was performed by hydrogenotrophic methanogens and subsequently analysed [31–34]. Furthermore, growth adaption to CO did not change the CH₄ production rates significantly in the case of *Methanothermobacter marburgensis*, and the specific growth rate (μ) of *Methanothermobacter thermautotrophicus* on pure CO was only a hundredth of the growth rate achieved using H₂:CO₂ as a substrate [31]. This led to the assumption that an artificial co-culture, where CO is converted to CH₄ in a successive bioprocess, would lead to higher efficiency, as microorganisms specifically adapted to the task of converting CO and producing CH₄ can be selected. Studies showed that the independent performance of the water gas shift reaction (WGSR) and biomethanation by two different organisms in the same vessel resulted in a more than 20-fold faster conversion than direct conversion by a single organism [34]. Therefore, we hypothesized that an artificial co-culture of a carboxydrotrophic, hydrogenogenic microbe with a hydrogenotrophic, autotrophic organism would drive favourable thermodynamic conditions for the WGSR. These conditions are created by direct removal of the gaseous metabolic end products of the WGSR, that is H₂:CO₂, by the methanogen. The conversions that are successively performed by the two organisms can be summarized with the following equation:



Overall, studies using a co-culture approach showed promising results for biomethane production rates [34,35]. Based on a previous study where a bacterial/archaeal co-culture was utilized [36], we wanted to investigate the potential of artificial archaeal co-cultures consisting of a carboxydrotrophic, hydrogenogenic archaeon, and different hydrogenotrophic, autotrophic, and methanogenic archaea for biomethanation. We selected *Thermococcus onnurineus* for performing the catalysis of the WGSR, as it was shown that μ and H₂ productivity on CO was substantially higher compared to other carboxydrotrophic and hydrogenogenic microbes [37–40]. To catalyse the second part of the reaction, *Methanocaldococcus jannaschii*, *Methanocaldococcus vulcanius*, and *Methanocaldococcus villosus* were selected, because of their similar cultivation requirements to *T. onnurineus* with respect to temperature, salt concentration, and pH optimum. All four organisms were isolated from deep-sea hydrothermal vents and belong to the Euryarchaeota. They are able to grow in a temperature range of 63 to 86 °C, a salt concentration of 1 to 5%, and a pH of 5.5 to 7.0. *T. onnurineus* is a heterotroph, while the methanogens are chemolithoautotrophs [41–44]. Moreover, hyperthermophilic organisms are more advantageous over mesophiles, since at higher temperatures, a faster conversion of CO by the carboxydrotrophic microorganism occurs, and a three times faster removal of H₂ is obtained by the methanogen [34,45]. Therefore, the properties of a hyperthermophilic environment positively affects growth and conversion rates and is, thus, advantageous over mesophilic conditions. Here, we analysed whether *T. onnurineus* together with one of the three methanogens can be grown as a powerful artificial archaeal co-culture to efficiently generate biological CH₄ from synthetic waste gases.

2. Materials and Methods

2.1. Chemicals

CO (99.999 Vol.-%), H₂:CO (60 Vol.-% in CO), H₂:CO₂ (80 Vol.-% in CO₂), and an artificial CO-containing syngas (CO₂ 16.7 Vol.-%, H₂ Vol.-% 16.8%, CH₄ Vol.-%14.7, N₂ Vol.-% 14.5%, and CO 37.3 Vol.-%) were used for closed batch experiments. For gas chromatography (GC), H₂ (99.999 Vol.-%), CO₂ (99.999 Vol.-%), CO (99.999 Vol.-%), H₂/CO₂ (80 Vol.-% in CO₂), H₂/N₂ (4.5 Vol.-% H₂ in N₂), CH₄ (99.995 Vol.-%), and the standard test gas (Messer GmbH, Wien, Austria) (containing 0.01 Vol.-% CH₄, 0.08 Vol.-% CO₂ in N₂) were used in addition to the gases mentioned above. All gases, except the standard

test gas, were purchased from Air Liquide (Air Liquide GmbH, Schwechat, Austria). All other chemicals were of the highest grade available.

2.2. Media

Medium A is a modified version of the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ) medium 282. The exact composition of medium A and medium B can be found in Supplementary Materials, Tables S1 and S2. Balch's vitamin solution was used [46]. For *M. marburgensis* and *M. thermotrophicus*, a phosphate-buffered medium was used [47].

2.3. Strains and Cultivation Conditions

The strains *M. jannaschii* JAL-1, *M. villosus* KIN24-T80, *M. vulcanius* M7, *M. marburgensis* DSM 2133 (Marburg), and *M. thermotrophicus* DSM 1053 (delta H) were purchased from the DSMZ. *T. onnurineus* NA1 was provided by Prof. Dr. Sung Gyun Kang (Korea Institute of Ocean Science and Technology (KIOST), Ansan, Korea).

Every cultivation was performed in closed batch mode [48]. Experiments were conducted in 120 mL serum bottles (Ochs Glasgeraetebau, Langerwehe, Germany), sealed with a 20 mm butyl rubber stopper and an aluminium crimp cap (Chemglass Life Science LLC, Vineland, NJ, USA). Serum bottles were filled with the corresponding medium and sealed. Anaerobic conditions were created by evacuating and re-pressurizing with H₂:CO₂ (4:1) to 0.5 barg five times. The bottles were autoclaved and stored at 4 °C until further use. Unless otherwise stated, the medium was augmented with the following sterile filtered stock solutions before inoculation: NaHCO₃, L-Cysteine-HCL, and Balch's Vitamin solution [46] (see Supplementary Materials, Tables S1 and S2). Afterwards, the bottles were flushed with H₂:CO₂ (4:1) to 0.5 barg, before the addition of autoclaved Na₂S·9H₂O.

The inoculum, 1 mL (2% v/v) of an actively grown culture, was added anaerobically. The final liquid phase in each serum bottle added up to 50 mL. Depending on the experiment, the headspace gas phase was exchanged with the corresponding gas. Bottles were incubated at 80 °C in either a double-layer shaking incubator at 100 rpm (LABWIT ZWYR-2102C, Labwit Scientific Pty Ltd., Burwood East, Australia) or in a water bath at ~100 lateral shakes per minute (GFL 1083, LAUDA-GFL, Burgwedel, Germany).

2.4. Pure-Culture Closed Batch Experiments

The methanogens *M. villosus*, *M. vulcanius*, and *M. jannaschii* were grown under 2 barg H₂:CO₂ in either medium A or B. A reduced version of them without vitamins, yeast extract, trace elements, or a combination of the three was also tested. The incubation rhythm consisted of 13 and 7 h incubation periods, with 2 h of sampling in between every period. The same rhythm was applied for cultivation of *T. onnurineus*. It was grown in the same medium, but under 1 barg CO. These incubation periods did not apply for *M. marburgensis* and *M. thermotrophicus*, because of their slow growth. Furthermore, they were grown in a phosphate-buffered *M. marburgensis* medium with either 1.7 barg CO, 1.7 barg H₂:CO (4:1), or 1.7 H₂:CO₂ (4:1).

2.5. Co-Culture Closed Batch Experiments

The co-cultures consisted of *T. onnurineus* and either *M. villosus*, *M. vulcanius*, or *M. jannaschii*. The experiments were conducted according to two general schemes. In the first setup, the experiment started by growing the respective *Methanocaldococcus* strain under 2 barg H₂:CO₂ first. After 13 h in the incubator, *T. onnurineus* was added. The bottles were then pressurized with CO, H₂:CO, or artificial syngas to 1 barg. In the second setup, *T. onnurineus* grew 13 h under pure CO, H₂:CO, or artificial syngas before one of the *Methanocaldococcus* strains was added. From this point onward, both schemes followed the same incubation rhythm, as described in the pure-culture experiments, and ended after a maximum of 80 h of cumulative incubation time. Every experiment was performed in hexuplicates (n = 6) on media A and B, as well as on reduced versions of them.

2.6. Sampling

The routinely performed sampling consisted of removing the cultures from the incubator and measuring the pressure as soon as the bottles cooled down to room temperature (~60 min). To analyse growth, 0.7 mL of the cultures were withdrawn for optical density (OD) measurements ($\lambda = 578$ nm). Lastly, the bottles were flushed and re-pressurized with the corresponding gas.

2.7. Analytical Procedures

OD of the cultures was measured via a spectrophotometer at $\lambda = 578$ nm (Specord 200 Plus, Analytik Jena, Jena, Germany). The headspace gas composition before and after inoculation in pure cultures was determined via the gas headspace pressure difference [18,47,49]. Gas evolution and uptake rates were calculated according to methods described in refs. [47,50,51]. Samples that were withdrawn for gas chromatography (GC) analysis did not undergo the sampling procedure. The headspace gas composition in co-culture experiments was analysed via GC, and the evolution and uptake rates were calculated [52]. Some of the negative controls revealed “air contamination” and were removed from the calculations of the results. To maintain the correct atmosphere, OD was measured after completion of the GC run and as such is only an estimate of the true OD.

3. Results

3.1. Growth Kinetics of Methanogens in Defined Medium

Growth and gas conversion by the methanogens *M. villosus*, *M. vulcanius*, and *M. jannaschii* were analysed in defined versions of media A and B on $\text{H}_2:\text{CO}_2$ (4:1), by removing complex components. Removal of the trace elements from the media hindered growth. *M. villosus* showed a higher methane evolution rate (MER) than the other two methanogens in medium A and B (Figure 1).

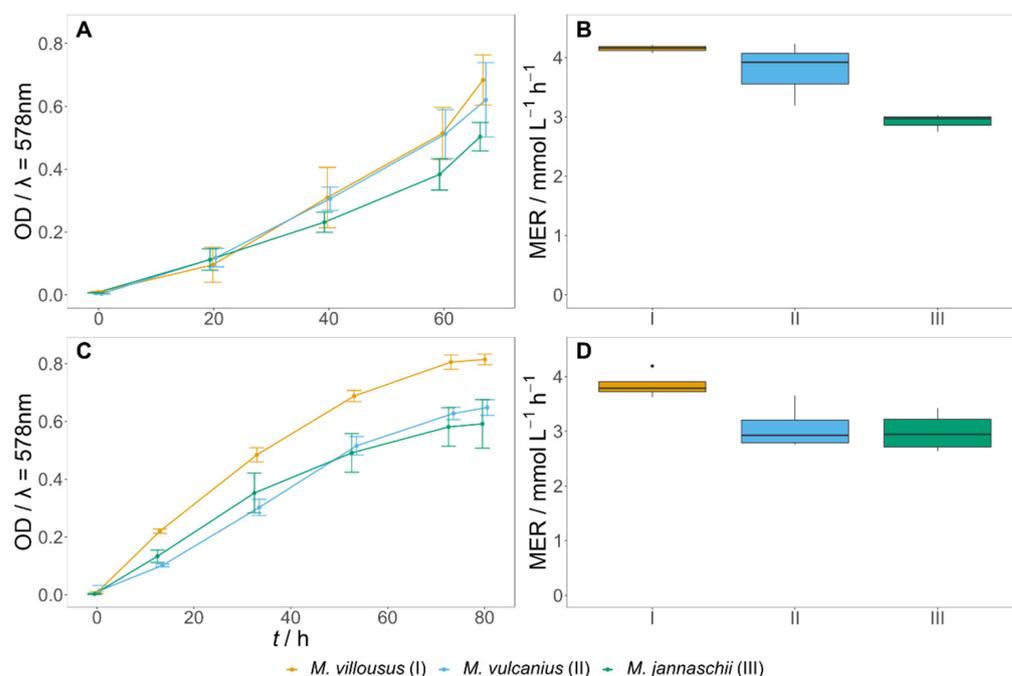


Figure 1. Growth kinetics and MER of *M. villosus*, *M. vulcanius*, and *M. jannaschii*. (A,B) grown in medium A; (C,D) grown in medium B. Both media were without yeast extract and vitamins. Error bars in the line graphs show the standard deviation. All experiments are N = 1, n = 3. However, N = 2, n = 3 for *M. villosus* using medium B.

M. marburgensis and *M. thermotrophicus* were grown on CO, H₂:CO (3:1) and H₂:CO₂ (4:1) [53]. The analysis showed that *M. marburgensis* and *M. thermotrophicus*, when grown on H₂:CO₂ (4:1), had a high turnover, and furthermore, experiments on H₂:CO resulted in poor CH₄ conversion. No growth was observed on pure CO (see Supplementary Materials, Figure S1, Table S5). Due to these findings, *M. marburgensis* and *M. thermotrophicus* were excluded from the co-culture experiments.

3.2. *T. onnurineus* Grown on CO

T. onnurineus was grown individually in a pure culture in media A and B. Results indicated that the organism reached higher OD₅₇₈ when grown on medium A as well as achieved a higher gas conversion than in media B. Omission of vitamins from medium B showed no difference regarding the H₂ evolution rate (HER) (see Supplementary Materials, Table S6, Figure S2).

3.3. Artificial Archaeal Co-Culture Engineering

Conversion of CO to CH₄ and CO₂ was performed with a co-culture consisting of *T. onnurineus* together with either *M. villosus*, *M. jannaschii*, or *M. vulcanius*. After pre-growth of the methanogens for 13 h, the cultures were inoculated with *T. onnurineus*. Growth and gas rates were analysed for all three co-cultures in media A and B (Tables 1 and 2, Figure 2). Co-cultures in medium A had relatively similar MERs ranging from 1.4 to 1.6 mmol L⁻¹ h⁻¹, whereas in medium B, the co-culture consisting of *M. villosus* and *T. onnurineus* achieved MERs between 1.6 and 2.0 mmol L⁻¹ h⁻¹. In general, the measured mean gas evolution and uptake rates were either equal or slightly higher in medium B. The higher CO uptake rate (COUR) and CO₂ evolution rate (CER) in medium B indicated that *T. onnurineus* performed better under this condition. It is likely that the H₂ uptake rate (HUR) of the methanogens positively influenced the COUR of *T. onnurineus* by creating favourable thermodynamic conditions [34].

Table 1. Mean gas evolution and uptake rates of the co-cultures cultivated in medium A on 100% CO¹.

Co-Culture	MER/mmol L ⁻¹ h ⁻¹	HUR/mmol L ⁻¹ h ⁻¹	CUR/mmol L ⁻¹ h ⁻¹	COUR/mmol L ⁻¹ h ⁻¹	CER/mmol L ⁻¹ h ⁻¹
<i>M. villosus</i> + <i>T. onnurineus</i>	1.4 ± 0.4	6.4 ± 1.5	1.4 ± 0.4	6.4 ± 1.5	5.0 ± 1.1
<i>M. vulcanius</i> + <i>T. onnurineus</i>	1.6 ± 0.3	7.0 ± 1.3	1.6 ± 0.3	7.0 ± 1.3	5.4 ± 0.9
<i>M. jannaschii</i> + <i>T. onnurineus</i>	1.5 ± 0.4	6.5 ± 1.5	1.5 ± 0.3	6.5 ± 1.5	5.0 ± 1.2

¹ Measurements were taken after a 7 h incubation period. Data collected at indicated timepoints in Figure 2 A. (N = 1, n = 6). Values are shown with standard deviation.

Table 2. Mean gas evolution and uptake rates of the co-cultures cultivated in medium B on 100% CO¹.

Co-Culture	MER/mmol L ⁻¹ h ⁻¹	HUR/mmol L ⁻¹ h ⁻¹	CUR/mmol L ⁻¹ h ⁻¹	COUR/mmol L ⁻¹ h ⁻¹	CER/mmol L ⁻¹ h ⁻¹
<i>M. villosus</i> + <i>T. onnurineus</i>	2.0 ± 0.3	8.3 ± 1.3	2.1 ± 0.3	8.4 ± 1.3	6.3 ± 0.9
<i>M. vulcanius</i> + <i>T. onnurineus</i>	1.6 ± 0.5	10.3 ± 0.2	2.6 ± 0.3	11.3 ± 0.7	8.7 ± 0.5
<i>M. jannaschii</i> + <i>T. onnurineus</i>	1.8 ± 0.4	9.0 ± 1.5	1.9 ± 0.5	9.1 ± 1.6	7.2 ± 1.2

¹ Measurements were taken after a 7 h incubation period. Data collected at indicated timepoints in Figure 2 B. (N = 1, n = 6). Values are shown with standard deviation.

The results suggested that the co-culture with *M. vulcanius* was less efficient in medium B, as it had a higher CO₂ uptake rate (CUR) and HUR, compared to medium A, but not a higher MER. This agrees with the results obtained from pure-culture experiments, where medium B led to a slightly lower MER of *M. vulcanius* compared to the other methanogens.

Comparison of the MERs between the pure and the co-cultures suggested that, on average, a lower MER was measured in the co-cultures (Tables 1 and 2, Figure 1). A likely reason for this was that they were dependent on the conversion of CO to CO₂ and H₂ by *T. onnurineus*. Figure 3 shows that the CH₄ production and growth occurred at the highest capacity, as the H₂ content in most cultures was below 1 mol%, except for one case, suggesting it was fully converted. As such, the rate-limiting step in CH₄ production by the methanogens in the co-culture was the availability of H₂.

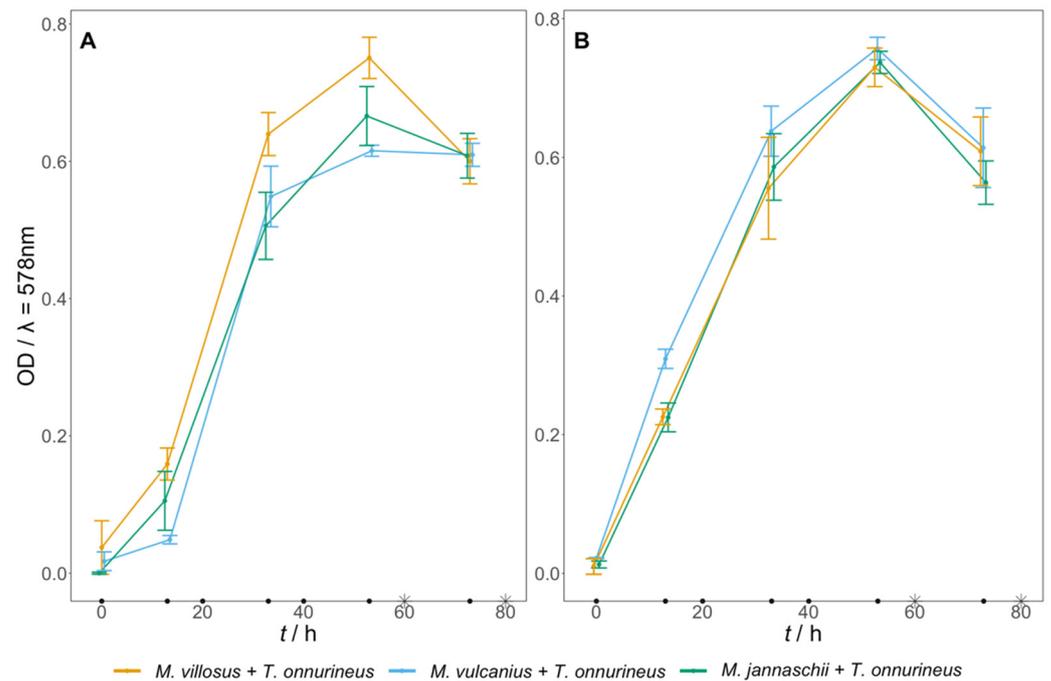


Figure 2. Growth kinetics of either *M. villosus*, *M. jannaschii*, or *M. vulcanius* with *T. onnurineus*. After the first 13 h, the medium was inoculated with *T. onnurineus* and the gas phase was exchanged from H₂:CO₂ to CO. The error bars show the standard deviation. Closed black circles on the x-axis mark the re-pressurising of the headspace with CO. The asterisks on the x-axis indicate the sampling points for GC analysis. (A) Growth in medium A; (B) growth in medium B. N = 1, n = 6.

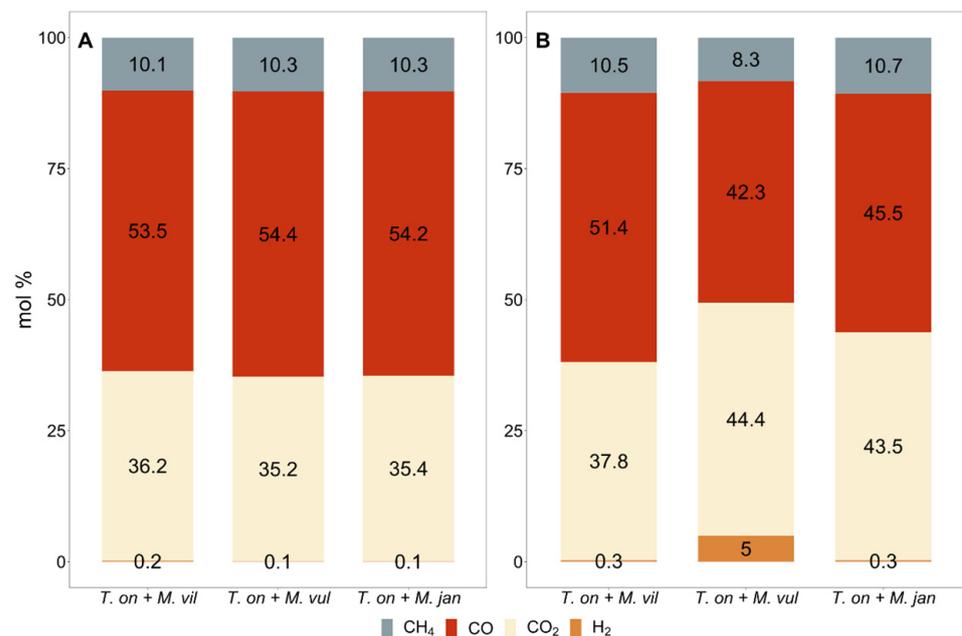


Figure 3. Relative mean molar gas composition in the co-cultures headspace after a 7 h incubation period on CO. Measurements were taken at the timepoints indicated in Figure 2. (A) Co-culture cultivated in medium A; (B) co-culture cultivated in medium B. N = 1, n = 6.

To circumvent H₂ limitation in the co-culture experiments, two strategies were employed. First, to create a functional culture with a high H₂ production rate in advance of the methanogen being added, an inoculation scheme was tested, where *T. onnurineus* was initially grown 13 h on CO, prior to addition of said methanogen. Second, a different gas

composition, H₂:CO (3:1) instead of pure CO, was used to provide additional H₂ available in the atmosphere. In addition, an experiment with a gas composition resembling that of industrial waste or syngases was performed [21]. The co-cultures were exposed to an artificial syngas (see Section 2.1) that could arise by industrial processes (Table 3). As *T. onnurineus* grown with *M. villosus* in medium B was considered the best co-culture, this pair was used. However, it should be noted that these were closed batch experiments and that the productivities are dependent on the conversion kinetics and on the tested time interval.

Table 3. Mean gas evolution rates of co-cultures in medium B under different gas compositions and inoculation orders ¹.

Co-Culture	Gas	MER/mmol L ⁻¹ h ⁻¹	CER/mmol L ⁻¹ h ⁻¹
<i>M. villosus</i> + <i>T. onnurineus</i>	CO	2.0 ± 0.3	6.3 ± 0.9
<i>T. onnurineus</i> + <i>M. villosus</i>	CO	1.1 ± 0.1	3.7 ± 1.0
<i>M. villosus</i> + <i>T. onnurineus</i>	H ₂ :CO	1.0 ± 0.4	0.0 ± 0.0
<i>T. onnurineus</i> + <i>M. villosus</i>	H ₂ :CO	1.0 ± 0.3	0.0 ± 0.0
<i>M. villosus</i> + <i>T. onnurineus</i>	Art. syngas	1.5 ± 0.5	0.9 ± 0.6
<i>T. onnurineus</i> + <i>M. villosus</i>	Art. syngas	0.9 ± 0.3	NA

¹ Measurements were taken after a 7 h incubation period (N = 1, n = 6). Values are shown with standard deviation.

In experiments with pure CO, the MER was twice as high when *M. villosus* was added first. The average H₂ content turned out to remain below 0.3 mol% (see Supplementary Materials, Table S7). When gassing with H₂:CO, interestingly, both inoculation schemes showed a decreasing performance over time, as revealed by a MER, which was only half of what was achieved when pure CO was used (Table 3). However, the measured CO₂ levels were below 0.1 mol% in both cases (see Supplementary Materials, Table S7). This could have been caused by the presence of H₂ in the H₂:CO, as the additional application of H₂ within the gas negatively affected growth and performance of *T. onnurineus*, which in turn most likely hindered the MER. The reduced CER by *T. onnurineus* may have been caused by the lower partial pressure of CO in the serum bottle headspace.

In the experiment where the artificial syngas mix was used and where *M. villosus* was pre-grown, the MER was 1.5 ± 0.5 mmol L⁻¹ h⁻¹. The average CH₄ mol% increased by about 10% to 24.8 mol%. This increase was similar to what was obtained when using pure CO, although the MER decreased as the experiment progressed. We did not observe a negative impact on the MER during co-culture experiments when elemental sulphur was added.

4. Discussion

This study is a new brick in the emerging research field of Archaea Biotechnology [54] and artificial microbial ecosystem design and engineering [55]. We investigated the conversion of one-carbon substrates (CO, H₂:CO, and CO-rich waste gases) by the artificially designed co-cultures of either *M. villosus*, *M. jannaschii*, or *M. vulcanius* together with *T. onnurineus*. Our results showed fast and reliable gas conversion, with a reduction in pure CO to about 50 mol% and simultaneous production of ~10 mol% CH₄ within 7 h. The most efficient conversion of the artificial syngas was performed by the co-culture *M. villosus* with *T. onnurineus*, inoculated in this order. This culture showed a CO reduction of 7%, starting from 37.3 mol% and an increase in CH₄ by ~10 mol% within 7 h. This proved the ability of the co-cultures to convert a variety of different CO-containing gas compositions with a lower proportion of CO.

A study with a similar experimental setup reported an ~6% CH₄ increase after 22 h [35]. We obtained 10% increase in 7 h. This suggests that the established co-culture (*T. onnurineus*/*M. villosus*) is of higher catalytic power. Nonetheless, most up-to-date published co-cultures were tested in different setups than the one reported in this study, making a direct comparison of evolution and uptake rates rather difficult [26,30,34]. Establishing different co-cultures in a bioreactor setup will, thus, be of great importance to fully understand their growth and production kinetics.

Pure-culture closed batch experiments of the methanogens in a defined medium on H₂:CO₂ showed a higher MER ($4.2 \pm 0.1 \text{ mmol L}^{-1} \text{ h}^{-1}$) than the co-cultures ($2.0 \pm 0.3 \text{ mmol L}^{-1} \text{ h}^{-1}$) (Figure 1, Table 3). Therefore, the potential for achieving a higher MER in co-cultures is possible if the necessary gas supply can be performed, and the inhibitory concentration of CO would not be surpassed [56]. Although the main limitation of the co-culture grown on pure CO was the availability of H₂ for methanogenesis and, hence, the conversions of CO to H₂ and CO₂ by *T. onnurineus* (Figure 3 and Supplementary Materials, Table S7). The addition of H₂ to the gas phase did not provide an increase in the MER. Rather, it led to a limitation of CO₂ availability (Supplementary Materials, Table S7), most likely due to the reduced performance of *T. onnurineus* under the lower CO partial pressure. Consequently, application of the artificial syngas led to an increase in MER, as biomethanation seems to be neither limited by CO₂ nor H₂ availability. Nonetheless, it did not reach the same values as in pure CO (Table 3). This can be explained by the concentration reduction in CO, CO₂, and H₂ in syngas by the other initially present gases, reducing substrate availability.

A change in the cultivation method to a bioreactor setup with higher pressure, agitation, and a constant gassing, resulting in having a higher gas solubility and higher gas transfer rate to the liquid phase, might increase the conversion of CO by *T. onnurineus* and the potential of the co-culture [37,39,48]. However, the ratio of the gas in the liquid phase has to be considered carefully, as high agitation might also lead to an excessive CO availability for the methanogens [34]. As recently more and more genetic tools for archaea become available, an overexpression of the carbon monoxide dehydrogenase could also be a solution to debottleneck CO conversion by *T. onnurineus* [39,57].

Unfortunately, *T. onnurineus* is still dependent on a complex medium containing yeast extract, which limits the potential industrial applicability. However, co-cultivation could be used to gain advantages or improve growth of both microorganisms through improving their syntrophic relationships [34]. Finding a suitable defined medium for *T. onnurineus* or replacing it by a different organism that can catalyse the WGSR from CO in minimal conditions is one suggested avenue of research. *Carboxydocella thermautotrophica* or *Carboxydocella sporoproducens* would fall into the same pH and temperature conditions as the herein employed methanogens [35].

A direct conversion of CO to CH₄ by a single organism such as *M. marburgensis* or *M. thermautotrophicus* is likely not the most suitable biotechnological approach, as the inhibitory concentration of CO for these methanogens is very low [31,34]. The artificially created archaeal co-cultures consisting of one of the hyperthermophilic methanogenic archaea *M. villosus*, *M. jannaschii*, and *M. vulcanius* together with *T. onnurineus*, as performed in this study, are highly efficient and reliable for biomethanation. By adaptation to a minimal medium and by performing targeted bioprocess development, artificial archaeal co-cultures could be of environmental, economic, and industrial value in renewable energy production and storage.

5. Conclusions

This contribution is another step in the rapidly developing research field of Archaea Biotechnology. Furthermore, it is a proof of principle for the design and engineering of artificial microbial co-cultures. Additionally, it marks a cornerstone in the use of artificial archaeal co-cultures for the biomethanation of syngas. This design of artificial archaeal co-cultures represents a novelty in the field of biomethanation, due the unique selection of

organisms and their hyperthermophilic growth conditions. The created co-cultures convert a variety of different CO-containing gases, including syngas, to CH₄. As the WGSR might still be the limiting factor, finding the best organism for conversion of H₂O:CO to H₂:CO₂ will be of future relevance to enhance the kinetics of the artificial co-culture. Apart from the choice of the archaea, the inoculation sequence of the strains and medium development are crucial factors for a successful design. These factors will be important in engineering the next steps for scaling up the bioprocess of artificial archaeal co-cultures. Inferring from our results, hyperthermophilic and anaerobic archaea are of great biotechnological relevance, as they act as highly efficient H₂ and CH₄ cell factories in artificial co-culture.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/fermentation7040276/s1>, Table S1: Chemical composition of medium A, Table S2: Chemical composition of medium B, Table S3: 141 trace element solution modified from DSMZ, Table S4: Holden's trace element solution 2, Table S5: The physiological maximal and mean values of CH₄ production and growth kinetics of *M. marburgensis* and *M. thermautotrophicus*, Table S6: HER of *T. onnurineus* after 7 h of incubation, Table S7: Relative mean molar gas composition of the co-culture's headspace after a 7 h incubation period, Figure S1: Growth of *M. marburgensis* and *M. thermautotrophicus* in defined medium, Figure S2: Growth of *T. onnurineus* at 1 barg CO.

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Abbreviations

CO ₂	Carbon dioxide
CH ₄	Methane
CO	Carbon monoxide
H ₂	Molecular hydrogen
WGSR	Water gas shift reaction
DSMZ	Deutsche Sammlung von Mikroorganismen und Zellkulturen
OD	Optical density
GC	Gas chromatography
MER	Methane evolution rate
HER	Molecular hydrogen evolution rate
COUR	Carbon monoxide uptake rate
CER	Carbon dioxide evolution rate
HUR	Molecular hydrogen uptake rate
CUR	Carbon dioxide uptake rate

References

1. EEA (European Environment Agency). *Primary Energy Consumption by Fuel in the EU-27*; EEA: København, Denmark, 2020; Volume 28.
2. EEA (European Environment Agency). *National Emissions Reported to the UNFCCC and to the EU Greenhouse Gas Monitoring Mechanism*; EEA: København, Denmark, 2019; Available online: <https://www.eea.europa.eu/data-and-maps/data/national-emissions-reported-to-the-unfccc-and-to-the-eu-greenhouse-gas-monitoring-mechanism-14> (accessed on 10 November 2021).
3. Sterner, M.; Specht, M. Power-to-Gas and Power-to-X—The History and Results of Developing a New Storage Concept. *Energies* **2021**, *14*, 6594. [[CrossRef](#)]
4. Götz, M.; Lefebvre, J.; Mörs, F.; McDaniel Koch, A.; Graf, F.; Bajohr, S.; Reimert, R.; Kolb, T. Renewable Power-to-Gas: A Technological and Economic Review. *Renew. Energy* **2016**, *85*, 1371–1390. [[CrossRef](#)]
5. Rönsch, S.; Schneider, J.; Matthischke, S.; Schlüter, M.; Götz, M.; Lefebvre, J.; Prabhakaran, P.; Bajohr, S. Review on Methanation – From Fundamentals to Current Projects. *Fuel* **2016**, *166*, 276–296. [[CrossRef](#)]
6. Eyl-Mazzega, M.-A.; Mathieu, C.; Boesgaard, K.; Daniel-Gromke, J.; Denysenko, V.; Liebetau, J.; Cornot-Gandolphe, S. *Biogas and Bio-Methane in Europe: Lessons from Denmark, Germany and Italy*; Études de l’Ifri; French Institute of International Relations (IFRI): Paris, France, 2019; ISBN 979-10-373-0025-6.
7. Rittmann, S.K.-M.; Seifert, A.H.; Bernacchi, S. Kinetics, Multivariate Statistical Modelling, and Physiology of CO₂-Based Biological Methane Production. *Appl. Energy* **2018**, *216*, 751–760. [[CrossRef](#)]
8. Mauerhofer, L.-M.; Zwirtmayr, S.; Pappenreiter, P.; Bernacchi, S.; Seifert, A.H.; Reischl, B.; Schmider, T.; Taubner, R.-S.; Paulik, C.; Rittmann, S.K.-M.R. Hyperthermophilic Methanogenic Archaea Act as High-Pressure CH₄ Cell Factories. *Commun. Biol.* **2021**, *4*, 289. [[CrossRef](#)]
9. Mauerhofer, L.-M.; Reischl, B.; Schmider, T.; Schupp, B.; Nagy, K.; Pappenreiter, P.; Zwirtmayr, S.; Schuster, B.; Bernacchi, S.; Seifert, A.H.; et al. Physiology and Methane Productivity of Methanobacterium Thermaggregans. *Appl. Microbiol. Biotechnol.* **2018**, *102*, 7643–7656. [[CrossRef](#)] [[PubMed](#)]
10. Abdel Azim, A.; Pruckner, C.; Kolar, P.; Taubner, R.-S.; Fino, D.; Saracco, G.; Sousa, F.L.; Rittmann, S.K.-M.R. The Physiology of Trace Elements in Biological Methane Production. *Bioresour. Technol.* **2017**, *241*, 775–786. [[CrossRef](#)]
11. Seifert, A.H.; Rittmann, S.; Herwig, C. Analysis of Process Related Factors to Increase Volumetric Productivity and Quality of Biomethane with Methanothermobacter Marburgensis. *Appl. Energy* **2014**, *132*, 155–162. [[CrossRef](#)]
12. Seifert, A.H.; Rittmann, S.; Bernacchi, S.; Herwig, C. Method for Assessing the Impact of Emission Gasses on Physiology and Productivity in Biological Methanogenesis. *Bioresour. Technol.* **2013**, *136*, 747–751. [[CrossRef](#)]
13. Burkhardt, M.; Jordan, I.; Heinrich, S.; Behrens, J.; Ziesche, A.; Busch, G. Long Term and Demand-Oriented Biocatalytic Synthesis of Highly Concentrated Methane in a Trickle Bed Reactor. *Appl. Energy* **2019**, *240*, 818–826. [[CrossRef](#)]
14. Burkhardt, M.; Koschack, T.; Busch, G. Biocatalytic Methanation of Hydrogen and Carbon Dioxide in an Anaerobic Three-Phase System. *Bioresour. Technol.* **2015**, *178*, 330–333. [[CrossRef](#)] [[PubMed](#)]
15. Jensen, M.B.; Strübing, D.; de Jonge, N.; Nielsen, J.L.; Ottosen, L.D.M.; Koch, K.; Kofoed, M.V.W. Stick or Leave—Pushing Methanogens to Biofilm Formation for Ex Situ Biomethanation. *Bioresour. Technol.* **2019**, *291*, 121784. [[CrossRef](#)]
16. Nock, W.J.; Serna-Maza, A.; Heaven, S.; Banks, C.J. Evaluation of Microporous Hollow Fibre Membranes for Mass Transfer of H₂ into Anaerobic Digesters for Biomethanization. *J. Chem. Technol. Biotechnol.* **2019**, *94*, 2693–2701. [[CrossRef](#)]
17. Tao, B.; Alessi, A.M.; Zhang, Y.; Chong, J.P.J.; Heaven, S.; Banks, C.J. Simultaneous Biomethanisation of Endogenous and Imported CO₂ in Organically Loaded Anaerobic Digesters. *Appl. Energy* **2019**, *247*, 670–681. [[CrossRef](#)]
18. Rittmann, S.K.-M.; Seifert, A.; Herwig, C. Essential Prerequisites for Successful Bioprocess Development of Biological CH₄ Production from CO₂ and H₂. *Crit. Rev. Biotechnol.* **2015**, *35*, 141–151. [[CrossRef](#)] [[PubMed](#)]
19. Rittmann, S.K.-M.R.; Seifert, A.H.; Krajete, A. Biomethanisierung—ein Prozess zur Ermöglichung der Energiewende? *BIOspektrum* **2014**, *20*, 816–817. [[CrossRef](#)]
20. Rittmann, S.K.-M.R. A Critical Assessment of Microbiological Biogas to Biomethane Upgrading Systems. In *Biogas Science and Technology*; Guebitz, G.M., Bauer, A., Bochmann, G., Gronauer, A., Weiss, S., Eds.; Advances in Biochemical Engineering/Biotechnology; Springer International Publishing: Cham, Switzerland, 2015; pp. 117–135. ISBN 978-3-319-21993-6.
21. Van der Stricht, W.; De Maré, C.; Plattner, T.; Fleischanderl, A.; Haselgribler, M.; Nair, P.; Wolf, C. *Sustainable Production of Low Carbon, Renewable Fuels by Fermenting Industrial Process Gasses from the Iron and Steel Industry*; ArcelorMittal, Primetals Technologies, LanzaTech: Gent, Belgium, 2017.
22. Safari, F.; Tavasoli, A.; Ataei, A.; Choi, J.-K. Hydrogen and Syngas Production from Gasification of Lignocellulosic Biomass in Supercritical Water Media. *Int. J. Recycl. Org. Waste Agric.* **2015**, *4*, 121–125. [[CrossRef](#)]
23. Dürre, P.; Eikmanns, B.J. C1-Carbon Sources for Chemical and Fuel Production by Microbial Gas Fermentation. *Curr. Opin. Biotechnol.* **2015**, *35*, 63–72. [[CrossRef](#)]
24. de Medeiros, E.M.; Noorman, H.; Maciel Filho, R.; Posada, J.A. Multi-Objective Sustainability Optimization of Biomass Residues to Ethanol via Gasification and Syngas Fermentation: Trade-Offs between Profitability, Energy Efficiency, and Carbon Emissions. *Fermentation* **2021**, *7*, 201. [[CrossRef](#)]
25. Phillips, J.R.; Huhnke, R.L.; Atiyeh, H.K. Syngas Fermentation: A Microbial Conversion Process of Gaseous Substrates to Various Products. *Fermentation* **2017**, *3*, 28. [[CrossRef](#)]

26. Westman, S.Y.; Chandolias, K.; Taherzadeh, M.J. Syngas Biomethanation in a Semi-Continuous Reverse Membrane Bioreactor (RMBR). *Fermentation* **2016**, *2*, 8. [[CrossRef](#)]
27. Devarapalli, M.; Lewis, R.S.; Atiyeh, H.K. Continuous Ethanol Production from Synthesis Gas by *Clostridium Ragsdalei* in a Trickle-Bed Reactor. *Fermentation* **2017**, *3*, 23. [[CrossRef](#)]
28. Rittmann, S.K.-M.; Lee, H.S.; Lim, J.K.; Kim, T.W.; Lee, J.-H.; Kang, S.G. One-Carbon Substrate-Based Biohydrogen Production: Microbes, Mechanism, and Productivity. *Biotechnol. Adv.* **2015**, *33*, 165–177. [[CrossRef](#)] [[PubMed](#)]
29. Thauer, R.K. Citric-Acid Cycle, 50 Years on: Modifications and an Alternative Pathway in Anaerobic Bacteria. *Eur. J. Biochem.* **1988**, *176*, 497–508. [[CrossRef](#)]
30. Klasson, K.; Cowger, J.; Ko, C.; Vega, J.; Clausen, E.; Gaddy, J. Methane Production from Synthesis Gas Using a Mixed Culture OfR. Rubrum M. Barkeri, and M. Formicicum. *Appl. Biochem. Biotechnol.* **1990**, *24*, 317–328. [[CrossRef](#)]
31. Daniels, L.; Fuchs, G.; Thauer, R.; Zeikus, J. Carbon Monoxide Oxidation by Methanogenic Bacteria. *J. Bacteriol.* **1977**, *132*, 118–126. [[CrossRef](#)]
32. O'Brien, J.M.; Wolkin, R.; Moench, T.; Morgan, J.; Zeikus, J. Association of Hydrogen Metabolism with Unitrophic or Mixotrophic Growth of *Methanosarcina Barkeri* on Carbon Monoxide. *J. Bacteriol.* **1984**, *158*, 373–375. [[CrossRef](#)]
33. Diender, M.; Pereira, R.; Wessels, H.J.; Stams, A.J.; Sousa, D.Z. Proteomic Analysis of the Hydrogen and Carbon Monoxide Metabolism of *Methanothermobacter Marburgensis*. *Front. Microbiol.* **2016**, *7*, 1049. [[CrossRef](#)]
34. Diender, M.; Uhl, P.S.; Bitter, J.H.; Stams, A.J.; Sousa, D.Z. High Rate Biomethanation of Carbon Monoxide-Rich Gases via a Thermophilic Synthetic Culture. *ACS Sustain. Chem. Eng.* **2018**, *6*, 2169–2176. [[CrossRef](#)]
35. Kohlmayer, M.; Robert, H.; Raimund, B.; Wolfgang, M. Simultaneous CO₂ and CO methanation using microbes. *Microbiology* **2018**. [[CrossRef](#)]
36. Schmider, T. Ecophysiology and Methane Productivity of Carboxydrotrophy-Based Archaeal Co-Cultures. Master's Thesis, University of Vienna, Vienna, Austria, 2018.
37. Kim, M.-S.; Fitriana, H.N.; Kim, T.W.; Kang, S.G.; Jeon, S.G.; Chung, S.H.; Park, G.W.; Na, J.-G. Enhancement of the Hydrogen Productivity in Microbial Water Gas Shift Reaction by *Thermococcus Onnurineus* NA1 Using a Pressurized Bioreactor. *Int. J. Hydrog. Energy* **2017**, *42*, 27593–27599. [[CrossRef](#)]
38. Lee, S.H.; Kim, M.-S.; Lee, J.-H.; Kim, T.W.; Bae, S.S.; Lee, S.-M.; Jung, H.C.; Yang, T.-J.; Choi, A.R.; Cho, Y.-J.; et al. Adaptive Engineering of a Hyperthermophilic Archaeon on CO and Discovering the Underlying Mechanism by Multi-Omics Analysis. *Sci. Rep.* **2016**, *6*, 22896. [[CrossRef](#)]
39. Kim, M.-S.; Bae, S.S.; Kim, Y.J.; Kim, T.W.; Lim, J.K.; Lee, S.H.; Choi, A.R.; Jeon, J.H.; Lee, J.-H.; Lee, H.S.; et al. CO-Dependent H₂ Production by Genetically Engineered *Thermococcus Onnurineus* NA1. *Appl. Environ. Microbiol.* **2013**, *79*, 2048–2053. [[CrossRef](#)] [[PubMed](#)]
40. Bae, S.S.; Kim, T.W.; Lee, H.S.; Kwon, K.K.; Kim, Y.J.; Kim, M.-S.; Lee, J.-H.; Kang, S.G. H₂ Production from CO, Formate or Starch Using the Hyperthermophilic Archaeon, *Thermococcus onnurineus*. *Biotechnol. Lett.* **2012**, *34*, 75–79. [[CrossRef](#)] [[PubMed](#)]
41. Bae, S.-S.; Kim, Y.-J.; Yang, S.-H.; Lim, J.-K.; Jeon, J.-H.; Lee, H.-S.; Kang, S.-G.; Kim, S.; Lee, J. *Thermococcus onnurineus* Sp. Nov., a Hyperthermophilic Archaeon Isolated from a Deep-Sea Hydrothermal Vent Area at the PACMANUS Field. *J. Microbiol. Biotechnol.* **2006**, *16*, 1826.
42. Bellack, A.; Huber, H.; Rachel, R.; Wanner, G.; Wirth, R. *Methanocaldococcus villosus* Sp. Nov., a Heavily Flagellated Archaeon That Adheres to Surfaces and Forms Cell–Cell Contacts. *Int. J. Syst. Evol. Microbiol.* **2011**, *61*, 1239–1245. [[CrossRef](#)] [[PubMed](#)]
43. Jeanthon, C.; l'Haridon, S.; Reysenbach, A.-L.; Corre, E.; Vernet, M.; Messner, P.; Sleytr, U.; Prieur, D. *Methanococcus vulcanius* Sp. Nov., a Novel Hyperthermophilic Methanogen Isolated from East Pacific Rise, and Identification of *Methanococcus* Sp. DSM 4213Tas *Methanococcus feruens* Sp. Nov. *Int. J. Syst. Evol. Microbiol.* **1999**, *49*, 583–589. [[CrossRef](#)]
44. Jones, W.; Leigh, J.; Mayer, F.; Woese, C.; Wolfe, R. *Methanococcus jannaschii* Sp. Nov., an Extremely Thermophilic Methanogen from a Submarine Hydrothermal Vent. *Arch. Microbiol.* **1983**, *136*, 254–261. [[CrossRef](#)]
45. Diender, M.; Stams, A.J.; Sousa, D.Z. Pathways and Bioenergetics of Anaerobic Carbon Monoxide Fermentation. *Front. Microbiol.* **2015**, *6*, 1275. [[CrossRef](#)]
46. Balch, W.E.; Wolfe, R. New Approach to the Cultivation of Methanogenic Bacteria: 2-Mercaptoethanesulfonic Acid (HS-CoM)-Dependent Growth of *Methanobacterium Ruminantium* in a Pressurized Atmosphere. *Appl. Environ. Microbiol.* **1976**, *32*, 781–791. [[CrossRef](#)]
47. Taubner, R.-S.; Rittmann, S.K.-M. Method for Indirect Quantification of CH₄ Production via H₂O Production Using Hydrogenotrophic Methanogens. *Front. Microbiol.* **2016**, *7*, 532. [[CrossRef](#)]
48. Rittmann, S.K.-M.; Herwig, C. A Comprehensive and Quantitative Review of Dark Fermentative Biohydrogen Production. *Microb. Cell Factories* **2012**, *11*, 115. [[CrossRef](#)] [[PubMed](#)]
49. Pappenreiter, P.A.; Zwiirtmayr, S.; Mauerhofer, L.-M.; Rittmann, S.K.-M.R.; Paulik, C. Development of a Simultaneous Bioreactor System for Characterization of Gas Production Kinetics of Methanogenic Archaea at High Pressure. *Eng. Life Sci.* **2019**, *19*, 537–544. [[CrossRef](#)] [[PubMed](#)]
50. Rittmann, S.K.-M.; Seifert, A.; Herwig, C. Quantitative Analysis of Media Dilution Rate Effects on *Methanothermobacter Marburgensis* Grown in Continuous Culture on H₂ and CO₂. *Biomass Bioenergy* **2012**, *36*, 293–301. [[CrossRef](#)]
51. Bernacchi, S.; Rittmann, S.K.-M.; Seifert, A.H.; Krajete, A.; Herwig, C. Experimental Methods for Screening Parameters Influencing the Growth to Product Yield (Y (x/CH₄)) of a Biological Methane Production (BMP) Process Performed with *Methanothermobacter Marburgensis*. *AIMS Bioeng.* **2014**, *1*, 72–86. [[CrossRef](#)]
52. Reischl, B.; Ergal, I.; Rittmann, S.K.-M. Biohydrogen Production Characteristics of *Desulfurococcus Amylyolyticus* DSM 16532. *Int. J. Hydrog. Energy* **2018**, *43*, 8747–8753. [[CrossRef](#)]

-
53. Pruckner, C. Evolutionary and Physiological Aspects of Potentially Carboxydophilic Archaea. Master's Thesis, University of Vienna, Vienna, Austria, 2019.
 54. Pfeifer, K.; Ergal, I.; Koller, M.; Basen, M.; Schuster, B.; Rittmann, S.K.-M.R. Archaea Biotechnology. *Biotechnol. Adv.* **2021**, *47*, 107668. [[CrossRef](#)]
 55. Ergal, I.; Bochmann, G.; Fuchs, W.; Rittmann, S.K.-M.R. Design and Engineering of Artificial Microbial Consortia for Biohydrogen Production. *Curr. Opin. Biotechnol.* **2022**, *73*, 74–80. [[CrossRef](#)]
 56. Guiot, S.R.; Cimpoaia, R.; Carayon, G. Potential of Wastewater-Treating Anaerobic Granules for Biomethanation of Synthesis Gas. *Environ. Sci. Technol.* **2011**, *45*, 2006–2012. [[CrossRef](#)]
 57. Straub, C.T.; Counts, J.A.; Nguyen, D.M.N.; Wu, C.-H.; Zeldes, B.M.; Crosby, J.R.; Conway, J.M.; Otten, J.K.; Lipscomb, G.L.; Schut, G.J.; et al. Biotechnology of Extremely Thermophilic Archaea. *FEMS Microbiol. Rev.* **2018**, *42*, 543–578. [[CrossRef](#)]