



Article Effect of Feeding Discarded Durian Peel Ensiled with Lactobacillus casei TH14 and Additives in Total Mixed Rations on Digestibility, Ruminal Fermentation, Methane Mitigation, and Nitrogen Balance of Thai Native–Anglo-Nubian Goats

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Abstract: The objective of this study was to evaluate the effect of fermented discarded durian peel with Lactobacillus casei TH14, cellulase, and molasses separately or in combination in total mixed rations on feed utilization, digestibility, ruminal fermentation, and nitrogen utilization in growing crossbreed Thai Native-Anglo-Nubian goats. Five crossbreed Thai Native-Anglo-Nubian goats (50%) at 9 to 12 months of age and 20 \pm 1 of body weight (BW) were assigned to a 5 \times 5 Latin square design. Evaluated treatments were fermented discarded durian peel without additives (FDP), fermented discarded durian peel with 5% of molasses (FDPM), fermented discarded durian peel with 2% of cellulase (FDPC), fermented discarded durian peel with 1.0×10^5 cfu/g fresh matter of L. *casei* TH14 (FDPL), and fermented discarded durian peel with 5% of molasses and 1.0×10^5 cfu/g fresh matter of L. casei TH14 (FDPML). This study showed that acid detergent fiber intake was different (p < 0.05) between goats fed FDP and those fed FDPLM, 0.24 g/d and 0.20 g/d, respectively. The FDPML ration had significantly (p < 0.05) greater apparent nutrient digestibility and a better propionate concentration compared with other treatments. FDPML treatment significantly (p < 0.05) decreased the acetate-to-propionate ratio, methane production, and urinary nitrogen. Therefore, treated discarded durian peel with molasses and L. casei TH14 in combination could add 25% of dry matter into the diet for growing goats without a negative impact.

Keywords: goat feeding; durian peel; silage additives; propionate; methane mitigation; nitrogen balance

1. Introduction

Durian, a seasonal fruit, is grown widely in tropical countries, where Malaysia and Thailand are the main producers [1]. Approximately 20 to 30% of durian is appropriate for human consumption, and 80 to 70% accounts for the durian peel, which is discarded as waste [2]. Discarded durian peel (DP) contains 10.30% crude protein (CP), 3.24% fat, 22.33% crude fiber (CF), 50.51% nitrogen-free extract (NFE), 9.50% cellulose, and 10.32% acid detergent lignin (ADL) [3]. Due to a high NFE content, DP spoils shortly after discarding. Ensiling is a well-known technique and is used to preserve high-fermentable-containing feed resources using lactic acid bacteria (LAB), converting sugar into lactic acid, resulting in low pH [4]. Ensiling additives including *Lactobacillus* strains, cellulase, and molasses are usually added to improve fermentation quality [5–8]. *Lactobacillus casei* TH14 (*L. casei* TH14), LAB strain, is a local strain isolated from sweet corn silage, which has high lactic acid production with a low pH range [9]. Cellulase is a popular fibrolytic enzyme added to break down cellulose, releasing soluble carbohydrate for LAB growth [10,11], while



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). molasses is added as a carbon source for LAB to ensure adequate lactic acid production if ensiling materials contain low water-soluble carbohydrate numbers [5]. Using *L. casei* TH14, cellulase and molasses have been reported to improve quality of sorghum [4], rice straw [12], and sugarcane bagasse [5]. In addition to fermentation quality improvement, *L. casei* TH14, cellulase, and molasses addition also improves feed utilization, propionate production, and methane mitigation [7,8,13]. However, the effect of *L. casei* TH14, cellulase, and molasses on DP quality and using fermented DP as roughage source in goat rations have never been evaluated. This study hypothesized that *L. casei* TH14 combined with molasses could improve DP quality, nutrient digestibility, propionate production, and methane mitigation. The objective of this study was to evaluate the effect of fermented discarded durian peel with *Lactobacillus casei* TH14, cellulase, and molasses separately or in combination in total mixed rations on feed utilization, digestibility, ruminal fermentation, and nitrogen utilization in growing crossbreed Thai Native–Anglo-Nubian goats.

2. Materials and Methods

2.1. Animal Ethics

The use of goats in this study was approved (MHESI 68014/674) by Institutional Animal Care and Use Committee, Prince of Songkla University.

2.2. Animals and Experimental Design

Five crossbreed Thai Native–Anglo-Nubian goats (50%) at 9 to 12 months of age and 20 ± 1 of body weight (BW) were used in this study. All goats were injected with ivermectin (IDECTIN[®] The British Dispensary (L.P.) CO., Ltd., Bangkok, Thailand) with 1 mL dose per 50 kg of BW to kill parasites before starting the experiment. Goats were assigned to a 5×5 Latin square design. Treatments were fermented discarded durian peel without additives (FDP), fermented discarded durian peel with 5% of molasses (FDPM), fermented discarded durian peel with 1.0×10^5 cfu/g fresh matter of *L. casei* TH14 (FDPL), and fermented discarded durian peel with 5% of molasses, and 1.0×10^5 cfu/g fresh matter of *L. casei* TH14 (FDPML).

2.3. Fermented Discarded Durian Peel Preparation

Discarded durian peel (Monthong-Durio zibthinus Murray) was obtained from Seahorse Intertrade Company Limited in Chana District, Songkhla Province, Thailand, and cut into 1 to 2 cm pieces. Then, discarded durian peel was fermented with the respective additives including molasses at 5% [5], cellulase at 2% [14], and L. casei TH14 at 1.0×10^5 cfu/g fresh matter [4]. Cellulase (powder form, 5×10^5 U/g activity, CAS number: 9004-34-6, Sinobios Imp. & Exp., Thanghai, China) and L. casei TH14 as a silage starter (composed of 80% trehalose, 15% lactose, and 1.0×10^{11} cfu/g L. casei TH14; Bio Ag Khon Kaen, Khon Kaen, Thailand) were used. Molasses was purchased from a local supplier located in Hat Yai District, Songkla Province, Thailand. Additives were dissolved in clean water, sprayed on discarded durian peel, mixed well, and fermented in 50 L plastic buckets (Changzhou Treering Plastics Co., Ltd., Changzhou, China) for 30 days. After fermenting for 30 days, fermented discarded durian peel samples were collected, dried at 60 °C for 72 h, and ground into 1 mm pieces to analyze the dry matter (DM), CP, and ash according to AOAC [15], and neutral detergent fiber (NDF), acid detergent fiber (ADF), and acid detergent lignin according to Van Soest et al. [16]. The chemical composition of fermented discarded durian peel is provided under Table 1. Fermentation characteristics of fermented DP were assessed. pH was measured according to Chen et al. [14] using pH meter (HANNA instruments HI 98153 microcomputer pH meter, Kallang Avenue, Singapore); briefly, 20 g of fermented DP samples were taken and mixed with 80 mL of distilled water and kept at 10 °C for 24 h. Samples were prepared with ammonia nitrogen (NH₃–N) using spectrophotometer (UV/VIS Spectrometer, PG Instruments Ltd., London, UK) and volatile fatty acids (lactic acid, acetate, and butyrate) using gas chromatography and analyzed according to So

Nutrient Composition			Treatments				
(% of DM)	FDP	FDPM	FDPC	FDPL	FDPML	Rice Straw	
DM	42.50	42.72	37.20	37.77	37.53	92.12	
OM	93.27	93.26	93.09	93.28	93.96	91.80	
СР	15.51	15.39	15.80	15.39	15.69	2.81	
NFC [†]	10.82	12.30	19.55	17.21	21.49	12.72	
NDF	65.45	63.12	57.26	57.30	57.01	74.71	
ADF	29.75	28.51	28.28	27.41	27.22	56.55	
ADL	8.52	7.98	8.26	6.74	6.59	4.59	
GE kcal/kg DM	4361.00	4306.26	4315.18	4363.60	4315.40	3501.53	
Ũ		Fermented of	discarded durian p	peel quality			
DM, %	17.0	17.3	16.5	16.5	17.4		
OM, % of DM	94.0	93.4	92.9	93.7	93.4		
CP, % of DM	7.3	8.3	7.2	7.2	7.5		
NDF, % of DM	62.5	61.0	73.4	61.9	65.5		
ADF, % of DM	41.7	38.6	45.8	43.9	42.3		
GE, kcal/kg DM	4413.8	4314.7	4737.4	4449.1	4205.4		
pH	3.74	3.79	3.66	3.73	3.74		
$NH_3 - N_7$, % of CP	1.29	0.70	1.18	0.74	0.89		
Lactic acid, % of DM	10.12	10.79	10.89	10.54	10.98		
Acetic acid, % of DM	1.02	1.05	1.07	1.04	1.09		
Butyric acid, % of DM	1.32	1.30	1.37	1.29	1.28		

et al. [8] The pH, NH₃–N, lactate, acetate, and butyrate of fermented discarded durian peel are provided in Table 1.

Table 1. Nutrient composition of TMR diets, rice straw, and fermented discarded durian peel quality treated with or without additives.

FDP = untreated discarded durian peel in TMR; FDPM = treated discarded durian peel with molasses in TMR; FDPC = treated discarded durian peel with cellulase in TMR; FDPL = treated discarded durian peel with *L. casei* TH14 in TMR; FDPML = treated discarded durian peel with molasses and *L. casei* TH14 in TMR. TMR compositions contain 25% fermented discarded durian peel with or without additives, 15% rice straw, 35.8% ground corn, 7.9% soybean meal, 0.4% fish meal, 5.4% leucaena meal, 7.2% palm kernel cake, 2.2% molasses, 0.3% dicalcium phosphate, 0.2% salt, and 0.6% premix. Premix per kg contains vitamin A: 10,000,000 IU; vitamin E: 70,000 IU; vitamin D: 1,600,000 IU; Fe: 50 g; Zn: 40 g; Mn: 40 g; Co: 0.1 g; Cu: 10 g; Se: 0.1 g; and I: 0.5 g. DM = dry matter; OM = organic matter; CP = crude protein; NFC = non-fiber carbohydrate; NDF = neutral detergent fiber; ADF = acid detergent fiber; ADL = acid detergent lignin; GE = gross energy. ¹ NFC = 100 – (% NDF + % CP + % EE + % ash) [17].

2.4. Feeding, Sample Collection, and Analysis

Feeding trial consisted of five 21-day periods, in which 14 days were used for dietary treatment adaptation and 7 days were used for sample analysis. Goats were separately stored in pens (0.11×0.95 m) with free access to clean water and mineral lick and fed daily *ad libitum* total mixed rations at a 40:60 ratio (25% fermented discarded durian peel, 15% rice straw, and 60% concentrate) at 8:00 a.m. and 16:00 p.m. The diets were formulated to meet the nutrient requirements of goats according to NRC [18], and chemical composition of dietary treatments is provided in Table 1. Diets offered and refusal were recorded daily to calculate DM intake. Goats were weighed at the beginning of the trial and at the end of each period throughout the trial to adjust DM intake and calculate the BW change of goats at the end of the trial.

During the last 7 days of each period, goats were kept in metabolism crate for sample collection and digestibility study. Diet and refusal data were collected throughout 7 days and divided into two portions. The first portion was used to analyze for DM content using oven drying at 100 °C, and second portion was deposited according to goat and period and stored at -20 °C for chemical composition analysis. Fecal and urine samples were gathered using total collection procedure. A total of 200 g of fecal sample was collected and oven-dried at 100 °C for DM analysis, and 5% of total feces was collected, deposited according to goat and period, and stored at -20 °C until analysis. Urine yield was collected using 5 L plastic tank consisting of 1 M H₂SO₄ to prevent nitrogen loss, and 10% of total

urine was taken, deposited according to goat and period, and stored at -20 °C. Before analysis, diet, refusal, and fecal samples were thawed, oven-dried at 60 °C for 72 h, and ground through a 1 mm screen to analyze for DM, CP, and OM using AOAC [15]. The NDF, ADF and ADL content were analyzed using Ankom fiber analyzer according to Van Soest et al. [16]. Gross energy content in diet, refusal, and fecal samples was analyzed using Bomb calorimetry (LECO, Berrien, MI, USA). Urine samples were thawed and analyzed for nitrogen content using AOAC [15] method to study nitrogen balance.

On day 21 of each period, at 0 h before feeding and 4 h after feeding, approximately 100 mL of ruminal fluid was collected using vacuum pump attached with stomach tube. Ruminal pH measurement was conducted immediately using pH meter (HANNA instruments HI 98153 microcomputer pH meter, Kallang Avenue, Singapore). Then, 60 mL of ruminal fluid samples was kept in plastic bottle containing $1 \text{ M H}_2\text{SO}_4$ at a ratio 1:10 (1 mL of H₂SO₄: 10 mL of ruminal fluid) and centrifuged at $3000 \times g$ for 15 min. Approximately 35 mL of supernatant was taken and stored at $-20 \,^{\circ}\text{C}$ to analyze NH₃–N using Kjeltec Auto 2200 analyzer (Foss, Tecator, UK) according to Bremner and Keeney [19] and volatile fatty acid including acetate, propionate, and butyrate were analyzed using gas chromatography (model HP6890, Hewlett-Packard, Palo Alto, CA, USA; column: Restek 1207384, Stabilwax -60 °C-250 °C, 30 m \times 250 µm \times 0.25 µm) according to Osaki et al. [20] as described by So et al. [8] Methane (CH_4) production was estimated using Moss et al. [21] equation, CH_4 $(g/d) = 0.45 \times acetate - 0.275 \times propionate + 0.4 \times butyrate$. Another 1 mL of ruminal fluid sample was kept in plastic bottle consisting of 9 mL of 10% formalin and stored in 4 °C in a refrigerator to count bacteria, protozoa, and fungi population using total direct count technique according to Galyean [22].

On day 21 of each period, samples of approximately 3 mL of blood were collected from the jugular vein at 0 h before feeding and 4 h after feeding and placed in heparinized tubes. Then, blood samples were centrifuged at $3000 \times g$ for 10 min, and plasma samples were taken and stored at -20 °C until analysis. Plasma samples were sent to Stanbio Laboratory (An EKF Diagnostics Company, Boerne, TX, USA) and used to analyze blood urea nitrogen (procedure no. 2020), glucose (procedure no. 1070), total protein (procedure no. 0250), and albumin (procedure no. 0285). Pack cell volume was measured using micro-hematocrit method, and mean corpuscular hemoglobin concentration, globulin, and albumin to globulin ratio were obtained by calculation. Hemoglobin was measured using commercial kits (Diamond Diagnostics, Egypt). Red blood cell count, mean corpuscular volume, and RBC distribution width were measured using hematological analyzer (ABX Micros 60, HORBA ABX, France).

2.5. Statistical Analysis

All data were analyzed using *Proc* Mixed model of SAS as follows:

$$Y_{ijkl} = \mu + \rho_i + \gamma_j + t_l + \tau_k + \varepsilon_{ijkl}$$
(1)

where Y_{ijkl} are the observation parameters, μ is the overall mean, ρ_i is the random effect of animal, γ_j is the random effect of periods, t_l is the random effect of time, τ_k is the fixed effect of treatments, and ε_{ijkl} is the error term. Differences among treatments were compared using Duncan's multiple range test, statistically accepted at p < 0.05.

3. Results

3.1. Nutrient Content of Diets

Dietary treatments were formulated to have approximately 15% CP content (Table 1). NFC, NDF, ADF, and ADL content varied among dietary treatments due to the quality of untreated and treated discarded durian peel used in the formulation. The diet containing untreated discarded durian peel (FDP) showed the highest NDF, ADF, and ADLs content and the lowest NFC content compared with FDPM, FDPC, FDPL, and FDPML diets. The chemical composition of untreated and treated discarded durian peel is provided (footnote of Table 1).

3.2. Nutrient Intake and Body Weight Change

Table 2 presents the intake of DM, OM, CP, NDF, and ADF, weight gain, and BW change in growing goats fed TMR containing untreated and treated discarded durian peel with additives. DM, OM, CP, and NDF intake were not different among treatments except ADF. ADF intake was different (p < 0.05) between FDP and FDPLM, which contained 0.24 g/d and 0.20 g/d, respectively. Weight gain and BW change in goats were not affected (p > 0.05) by dietary treatments.

Table 2. Effects of untreated and treated discarded durian peel in TMR on intake and body weight change in growing goats.

T .	Dietary Treatments						
Items –	FDP	FDPM	FDPC	FDPL	FDPML	SEM	<i>p</i> -Value
DM intake							
kg/d	0.796	0.757	0.785	0.797	0.806	0.02	0.56
%BW	2.96	2.84	2.89	2.94	3.04	0.06	0.28
g/kg BW ^{0.75}	67.39	64.44	66.08	67.66	68.98	1.56	0.35
0 0			Nutrient intake	, g/d			
OM	0.752	0.696	0.711	0.744	0.712	0.03	0.54
СР	0.125	0.115	0.120	0.123	0.119	0.01	0.42
NDF	0.528	0.471	0.437	0.457	0.432	0.02	0.05
ADF	0.236 ^a	0.215 ^{ab}	0.216 ^{ab}	0.216 ^{ab}	0.199 ^b	0.01	0.04
Weight gain, kg	2.40	1.90	1.90	2.80	2.30	0.38	0.45
BW change, kg/d	0.114	0.090	0.090	0.132	0.110	0.01	0.46
BW change,%	9.17	7.61	7.12	10.70	9.00	1.36	0.41

SEM = standard error of the mean; $BW^{0.75}$ = metabolic body weight; DM = dry matter, OM = organic matter; CP = crude protein; NDF = neutral detergent fiber; ADF = acid detergent fiber; BW = body weight. ^{a,b} Means in the same row with different letters differ (p < 0.05).

3.3. Apparent Nutrient Digestibility, Digestible Nutrient Intake, and Energy Intake

Apparent total tract digestibility was affected significantly (p < 0.05) by dietary treatments (Table 3). DM, OM, CP, NDF, and ADF digestibility were significantly lower in FDP compared with FDPML. FDPM, FDPC, FDPL, and FDPML were comparable in terms of DM, OM, CP, NDF, and ADF digestibility. Digestible nutrient intake including OM, CP, NDF, and ADF was not different (p > 0.05) among treatments. Estimated ME intake, expressed as Mcal/d, was not significant among treatments; however, estimated ME intake, expressed as per kg DM intake, was significant (p < 0.05) among treatments. ME intake, expressed as per kg DM intake, was significantly observed between FDP, FDPM and FDPML, in amounts of 2.55 Mcal/kg DM, 2.62 Mcal/kg DM, and 2.69 Mcal/kg DM, respectively.

T .		Dietary Treatments						
Items -	FDP	FDPM	FDPC	FDPL	FDPML	SEM	<i>p</i> -Value	
Apparent total								
tract digestibility,								
%								
DM	70.42 ^b	72.79 ^a	73.91 ^a	74.07 ^a	73.81 ^a	0.45	< 0.01	
OM	71.82 ^b	73.95 ^a	75.02 ^a	75.48 ^a	75.16 ^a	0.49	< 0.01	
СР	68.42 ^c	71.11 ^b	72.66 ^{ab}	73.07 ^{ab}	73.63 ^a	0.61	< 0.01	
NDF	63.83 ^b	70.48 ^a	70.23 ^a	70.96 ^a	71.03 ^a	0.47	< 0.01	
ADF	40.96 ^b	47.13 ^a	47.09 ^a	47.10 ^a	46.08 ^a	0.96	< 0.01	
Digestible nutrient								
intake, kg/d								
OM	0.543	0.515	0.534	0.562	0.536	0.02	0.47	
СР	0.087	0.084	0.086	0.090	0.087	0.01	0.68	
NDF	0.338	0.332	0.307	0.325	0.307	0.01	0.21	
ADF	0.098	0.102	0.102	0.102	0.092	< 0.01	0.38	
Estimated energy								
intake								
ME Mcal/d	2.06	1.96	2.03	2.14	2.04	0.07	0.48	
ME Mcal/kg DM	2.55 ^c	2.62 ^b	2.65 ^{ab}	2.68 ^{ab}	2.69 ^a	0.02	< 0.01	

Table 3. Effects of untreated and treated discarded durian peel in TMR on nutrient digestibility and digestible nutrient intake of growing goats.

FDP = untreated discarded durian peel; FDPM = treated discarded durian peel with molasses; FDPC = treated discarded durian peel with cellulase; FDPL = treated discarded durian peel with *L. casei* TH14; FDPML = treated discarded durian peel with molasses and *L. casei* TH14; SEM = standard error of the mean; DM = dry matter; OM = organic matter; CP = crude protein; NDF = neutral detergent fiber; ADF = acid detergent fiber; ME = metabolizable energy. ^{a-c} Means in the same row with different letters differ (p < 0.05). [†] 1 kg DOM = 3.8 Mcal ME/kg [23].

3.4. Rumen Fermentation Characteristics

Dietary treatments significantly affected ruminal pH and NH₃–N but not ruminal temperature (Table 4). Mean ruminal pH and NH₃–N were significantly (p < 0.05) observed between FDP and FDPML but FDPM, FDPC, FDPL, and FDPML were comparable. Ruminal pH and NH₃–N were 6.71 and 22.58 mg/dL in FDP and 6.44 and 17.29 mg/dL in FDPML, respectively. Dietary treatments significantly affected propionate concentration, the acetate-to-propionate ratio and acetate—butyrate-to-propionate ratio except total VFA, acetate, and butyrate concentration. The propionate concentration was significantly higher, while the acetate-to-propionate ratio and acetate—butyrate-to-propionate ratio were significantly lower in FDPML than FDP, FDPM, FDPC, and FDPL. FDPML showed the highest mean propionate concentration (20.31%) and lowest mean acetate-to-propionate ratio (3.58) and acetate + butyrate-to-propionate ratio (3.94). CH₄ production was affected significantly (p < 0.05) by dietary treatments. Mean CH₄ production was significant between FDP and FDPML, at 32.93 g/d and 29.41 g/d, respectively, but FDP, FDPM, FDPC, and FDPL were comparable for CH₄ production.

τ.		1	Dietary Treatm	ents		CEN	
Items	FDP	FDPM	FDPC	FDPL	FDPML	SEM	<i>p</i> -Value
Ruminal pH							
0 h	6.61	6.56	6.55	6.56	6.50	0.09	0.94
4 h	6.81	6.64	6.39	6.57	6.39	0.11	0.06
Mean	6.71 ^a	6.60 ^{ab}	6.47 ^b	6.56 ^{ab}	6.44 ^b	0.05	0.04
Temperature, °C							
0 h	39.0	39.2	39.0	39.2	39.2	0.15	0.50
4 h	39.6	39.4	39.2	39.2	39.6	0.25	1.00
Mean	39.3	39.3	39.1	39.2	39.4	0.19	0.78
			Ammonia–niti	rogen, mg/dL			
0 h	22.29 ^a	18.57 ^b	20.29 ^{ab}	19.14 ^b	18.00 ^b	0.78	0.02
4 h	22.86 ^a	18.57 ^b	18.29 ^b	18.57 ^b	16.57 ^b	0.63	< 0.01
Mean	22.58 ^a	18.57 ^b	19.29 ^b	18.86 ^b	17.29 ^b	0.43	< 0.01
				tty acid, mM/L			
0 h	86.50	93.13	97.63	96.79	99.54	6.26	0.62
4 h	95.12	97.09	100.26	102.30	104.58	8.42	0.93
Mean	90.81	95.11	98.95	99.55	102.06	4.98	0.56
Acetate, %							
0 h	74.17	77.22	75.94	74.31	71.73	1.49	0.17
4 h	73.99	73.77	72.31	72.89	70.75	1.14	0.32
Mean	74.08	75.49	74.12	73.61	71.24	1.21	0.23
Propionate, %							
0 h	13.88 ^b	13.23 ^b	15.03 ^b	15.82 ^b	19.77 ^a	0.82	< 0.01
4 h	16.28 ^b	16.83 ^b	18.17 ^b	17.92 ^b	20.85 ^a	0.73	< 0.01
Mean	15.08 ^b	15.03 ^b	16.59 ^b	16.87 ^b	20.31 ^a	0.66	< 0.01
Butyrate, %							
0 h	9.77	7.81	7.45	8.06	7.18	0.87	0.30
4 h	8.91	7.96	8.32	8.24	7.50	0.54	0.49
Mean	9.34	7.89	7.89	8.15	7.34	0.64	0.31
			Acetate:Prop				
0 h	6.02 ^a	5.98 ^a	5.32 ^a	4.92 ^a	3.73 ^b	0.38	< 0.01
4 h	4.62 ^a	4.49 ^a	4.16 ^{ab}	4.14 ^{ab}	3.42 ^b	0.27	0.05
Mean	5.32 ^a	5.23 ^a	4.74 ^a	4.53 ^a	3.58 ^b	0.27	< 0.01
1,10ull	0.02			e:Propionate rat		0.2	10101
0 h	6.74 ^a	6.58 ^a	5.85 ^a	5.45 a	4.11 ^b	0.40	< 0.01
4 h	5.18 ^a	4.96 ^a	4.62 ^{ab}	4.60 ^{ab}	3.78 ^b	0.28	0.04
Mean	5.96 ^a	5.77 ^{ab}	5.24 ^{ab}	5.03 ^b	3.94 °	0.27	< 0.01
Methane, g/d	0.70	0.77	0.21	0.00	0.71	0.27	N0.01
0 h	33.47 ^a	34.23 ^a	33.02 ^a	32.32 ^a	29.72 ^b	0.65	< 0.01
4 h	32.38 ^a	31.75 ^a	30.88 ^a	31.17 ^a	29.72 29.10 ^b	0.56	0.01
Mean	32.38 32.93 ^a	32.99 ^a	31.95 ^a	31.74 ^a	29.10 ^b	0.50	< 0.01
Ivieal1	32.93	32.99	31.93	31.74	27.41	0.32	<0.01

Table 4. Effects of untreated and treated discarded durian peel on rumen characteristics in growing goats.

FDP = untreated discarded durian peel; FDPM = treated discarded durian peel with molasses; FDPC = treated discarded durian peel with cellulase; FDPL = treated discarded durian peel with *L. casei* TH14; FDPML = treated discarded durian peel with molasses and *L. casei* TH14; SEM = standard error of the mean. ^{a-c} Means in the same row with different letters differ (p < 0.05).

3.5. Microbial Population

Dietary treatments did not affect (p > 0.05) the bacteria, fungal zoospores, total protozoa, *Holotrich* sp., or *Entodiniomorphs* sp. populations (Table 5). The mean total protozoal population was lowest in FDPML, at 2.46×10^6 cell/mL.

τ.		D	ietary Treatmen	ts			a Value
Items FDP	FDPM	FDPC	FDPL	FDPML	SEM	<i>p</i> -Value	
			Bacteria, ×1	0 ¹⁰ cell/mL			
0 h	1.60	1.56	1.45	1.35	1.45	1.38	0.50
4 h	1.90	2.20	1.67	1.63	1.56	2.01	0.67
Mean	1.75	1.88	1.56	1.49	1.51	1.67	0.43
			Fungal zoospore	es, $\times 10^6$ cell/ml	Ĺ		
0 h	2.18	1.92	1.67	1.67	1.55	0.29	0.13
4 h	2.25	2.67	2.16	1.65	1.62	0.40	0.19
Mean	2.21	2.29	1.92	1.66	1.59	0.30	0.15
			Total Protozoa	, $\times 10^6$ cell/mL			
0 h	2.88	2.51	2.47	2.21	2.29	0.26	0.11
4 h	3.16	3.47	3.15	2.63	2.61	0.32	0.13
Mean	3.02	2.99	2.81	2.41	2.46	0.26	0.09
			Holotrich sp.,	$\times 10^5$ cell/mL			
0 h	0.63	0.57	0.40	0.72	0.27	0.28	0.74
4 h	0.50	0.75	0.57	1.07	1.15	0.45	0.34
Mean	0.56	0.66	0.49	0.90	0.72	0.21	0.44
		l	Entodiniomorphs s	sp., $\times 10^6$ cell/m	۱L		
0 h	2.82	2.45	2.43	2.14	2.26	1.47	0.11
4 h	3.11	3.40	3.09	2.52	2.50	1.44	0.10
Mean	2.96	2.92	2.76	2.32	2.39	1.45	0.13

Table 5. Effect of untreated and treated discarded durian peel in TMR on ruminal microbe population in growing goats.

FDP = untreated discarded durian peel; FDPM = treated discarded durian peel with molasses; FDPC = treated discarded durian peel with cellulase; FDPL = treated discarded durian peel with *L. casei* TH14; FDPML = treated discarded durian peel with molasses and *L. casei* TH14; SEM = standard error of the mean.

3.6. Blood Metabolites

Dietary treatments did not affect (p > 0.05) blood metabolites such as glucose, pack cell volume, BUN, total protein, albumin, globulin, the albumin-to-globulin ratio, red blood cells, hemoglobin, mean corpuscular volume, mean corpuscular hemoglobin concentration, or the RBC distribution width (Table 6). FDPML had a lower BUN concentration and higher blood glucose concentration compared with FDP, FDPM, FDPC, and FDPL.

Table 6. Effects of untreated and treated discarded durian peel in TMR on blood metabolites in growing goats.

Attribute		D	(E) (
	FDP	FDPM	FDPC	FDPL	FDPML	SEM	<i>p</i> -Value
Glu, mg/dL	68.70	70.00	69.90	70.00	70.20	1.68	0.99
PCV, %	28.40	28.70	29.30	29.40	28.20	0.57	0.51
BUN, mg/dL	20.97	20.09	20.54	20.45	17.62	1.24	0.10
TP, g/L	6.21	6.19	6.33	6.24	6.20	0.08	0.82
ALB, g/L	3.67	3.68	3.71	3.69	3.70	0.03	0.92
GLB, g/L	2.53	2.51	2.62	2.54	2.50	0.06	0.73
A/G ratio	1.46	1.50	1.44	1.49	1.50	0.03	0.61
RBC, 10 ⁶ /μL	4.39	4.52	4.68	4.36	4.17	0.12	0.05
Hb, g/dL	10.60	10.65	10.92	10.84	10.87	0.23	0.67
MCV, fL	64.92	63.62	62.97	67.69	67.93	1.42	0.05
MCHC, g/dL	37.28	37.21	37.27	36.99	37.11	0.32	0.96
RDW, %	29.10	29.20	29.50	29.12	29.25	0.29	0.87

FDP = untreated discarded durian peel; FDPM = treated discarded durian peel with molasses; FDPC = treated discarded durian peel with cellulase; FDPL = treated discarded durian peel with *L. casei* TH14; FDPML = treated discarded durian peel with molasses and *L. casei* TH14; SEM = standard error of the mean; Glu = glucose; PCV = pack cell volume; BUN = blood urea nitrogen; TP = total protein; ALB = albumin; GLB = globulin; A/G ratio = albumin-to-globulin ratio; RBC = red blood cell; Hb = hemoglobin; MCV = mean corpuscular volume; MCHC = mean corpuscular hemoglobin concentration; RDW = RBC distribution width.

3.7. Nitrogen Utilization

Dietary treatments did not affect (p > 0.05) nitrogen intake, total nitrogen loss, fecal nitrogen, nitrogen absorption, or nitrogen retention but significantly affected urinary nitrogen (Table 7). Urinary nitrogen was significantly observed between FDP and FDPML, 2.58 g/d and 2.32 g/d, respectively. Apparent nitrogen absorption expressed as % of nitrogen intake was significant among treatments, in which FDPML had the highest apparent nitrogen absorption, 73.63% of nitrogen intake, respectively.

Table 7. Effects of untreated and treated discarded durian peel in TMR on nitrogen utilization in growing goats.

Items ——		D					
	FDP	FDPM	FDPC	FDPL	FDPML	SEM	<i>p</i> -Value
Balance, g/d							
N intake	20.03	18.39	19.31	19.65	19.03	0.60	0.41
Total N loss	8.34	7.55	7.55	7.77	7.23	0.37	0.40
Fecal N	5.77	5.27	5.24	5.25	4.97	0.15	0.05
Urine N	2.58 ^a	2.28 ^b	2.32 ^b	2.52 ^a	2.32 ^b	0.30	0.03
Absorbed N	14.27	13.12	14.07	14.40	14.06	0.48	0.93
Retained N	11.69	10.84	11.75	11.88	11.74	0.45	0.39
% of N intake							
Fecal N	29.19	28.90	27.34	26.94	26.37	0.41	0.05
Urine N	13.18	12.32	11.78	12.96	11.85	1.44	0.94
Absorbed N	70.81 ^b	71.10 ^b	72.66 ^a	73.06 ^a	73.63 ^a	0.40	< 0.01
Retained N	57.63	58.78	60.88	60.10	61.79	1.48	0.35

FDP = untreated discarded durian peel; FDPM = treated discarded durian peel with molasses; FDPC = treated discarded durian peel with cellulase; FDPL = treated discarded durian peel with *L. casei* TH14; FDPML = treated discarded durian peel with molasses and *L. casei* TH14; SEM = Standard error of the mean. ^{a,b} Means in the same row with different letters differ (p < 0.05).

4. Discussion

Using untreated and treated discarded durian peel at the same amount in the diet mainly caused a change in fiber content such as NDF and ADF content. Using untreated discarded durian peel in the diet (FDP) had a higher NDF and ADF content compared with diets containing discarded durian peel treated with molasses (FDPM), discarded durian peel treated with cellulase (FDPC), discarded durian peel treated with *L. casei* TH14 (FDPL) and discarded durian peel treated with molasses and *L. casei* TH14 (FDPML). This was caused by the higher NDF and ADF content presented in untreated discarded durian peel than treated discarded durian peel. A lower NDF and ADF content in treated discarded durian peel is due to acid hydrolysis action during fermentation and cellulase activity. So et al. [7] used sugarcane bagasse treated with molasses in combination with cellulase or *L. casei* TH14 in TMR for dairy cows had a lower NDF content compared with the TMR diet containing untreated sugarcane bagasse. Oba and Allen [24] stated that fiber intake, ruminal fermentation, and production efficiency could be influenced by dietary NDF content and digestibility.

In this study, the intake of DM, expressed as either kg/d or %BW or g/kg BW^{0.75}, was not significant among treatments. Thus, using untreated or treated discarded durian peel with additives at 25% DM in the diet did not affect the daily DM intake of growing goats. Similarly, So et al. [7] found that 50% DM of untreated and treated sugarcane bagasse combined with additives in TMR diets did not affect the daily DM intake of dairy cows. This suggests that any roughage feeds treated with or without silage additives does not affect the DM intake in ruminants. Intake of ADF (g/d) was significantly different between the FDP and FDPML treatments. The reason for this effect is not clear, although DM, OM, and NDF intake were not different among these treatments. So et al. [7] found nutrient intake was unchanged in dairy cows fed TMR containing untreated and treated sugarcane bagasse with additives. Cherdthong et al. [13] fed untreated and treated rice straw with

molasses in combination with cellulase or *L. casei* TH14 to Thai Native beef cattle and found no effect on nutrient intake. Dietary treatments did not affect weight gain or BW change in growing goats. The effect of additives in combination on performance is small and unclear [25]. Addah et al. [26] compared untreated and treated whole-crop barley with inoculant combination (*L. buchneri*, *L. plantarum*, and *L. casei*) and found no change in

weight gain in growing feedlot steers. Using pretreatment roughage feeds with additives such as LAB, molasses, and fibrolytic enzymes in the diet has been reported to improve nutrient digestibility in the rumen [6,7,13,27]. Additives contribute two mechanisms during fermentation: (1) acid hydrolysis reaction and (2) direct effect of fibrolytic enzymes on polysaccharide structure, which increases feed digestion efficiency of ruminal microbes [10,28]. This study showed that FDPM, FDPC, FDPL, and FDPML were significantly better in terms of DM, OM, CP, NDF and ADF digestibility compared with FDP. This could be due to the pretreatment effect of molasses, cellulase, and L. casei TH14 during fermentation of carbohydrate structures, subsequently resulting in nutrient digestibility improvement. So et al. [7] reported that L. casei TH14, cellulase, and molasses in combination with treated sugarcane bagasse significantly increased OM, CP, NDF, and ADF digestibility in dairy cows fed TMR diets compared with untreated sugarcane bagasse. Zhao et al. [29] evaluated the in vitro degradability of untreated rice straw and rice straw treated with L. plantarum and molasses and found a significant improvement in DM and NDF degradability by L. plantarum and molasses in combination. FDPML significantly increased estimated ME intake expressed per kg DM intake compared with FDP. This could be due to a significantly higher OM digestibility found in FDPML than FDP. So et al. [7] similarly found that a combination of sugarcane bagasse treated with molasses and L. casei TH14 in TMR-fed dairy cows increased their estimated ME intake compared untreated sugarcane bagasse.

Ruminal pH significantly determines the normal function of microbes in the rumen [30–33]. The normal ruminal pH ranges from 5.5 to 7.0 [34]. This study showed that FDPML significantly decreased the mean ruminal pH by 0.27 compared with FDP; however, the pH was in a normal range (6.4 to 6.7). This could be due to the higher lactic acid and LAB population found in FDPML than FDP. In addition, a significantly higher propionate concentration in FDPML compared with FDP could contribute to a lower pH in FDPML compared with FDP. pHs ranging from 6.4 to 6.7 showed improved fiber digestibility (Table 3) as activity of cellulolytic bacteria slows down at a pH less than 6 [35]. Similarly, So et al. [7] showed that a combination of sugarcane bagasse treated with molasses and L. casei TH14 in TMR fed to dairy cows significantly decreased their ruminal pH by 0.07 after 4 h of feeding. Zhang et al. [36] revealed that whole-plant corn ensiled with complex inoculants (L. plantarum L28, Enterococcus faecium EF08, and Lactobacillus buchneri LBC136) significantly decreased ruminal pH by 0.21 compared with ensiled whole-plant corn without inoculants in growing-finishing cattle. Lower ruminal pH leads to lactic-acid-dependent acid production in the rumen and is achieved approximately 2 to 6 h after feeding [30]. Time after offering feed and lactic acid supply rate mainly determine the change in ruminal pH [2,37]. NH₃–N is a main nitrogen source for microbial synthesis in the rumen (5 mg/dL minimum and 30 mg/dL maximum requirement) [38]. Additive-treated discarded durian peel significantly decreased mean NH₃–N concentration. This could be due to the activity of LAB present in fermented discarded durian peel that affected deamination, resulting in less ruminal protein degradation and enhancing nitrogen utilization in the lower digestive tract. So et al. [7] similarly revealed that additives combined with treated sugarcane bagasse significantly decreased NH3-N concentration from 22 to 20 mg/dL after 4 h of feeding compared with untreated sugarcane bagasse in dairy cows. FDPML significantly increased the propionate concentration, resulting in lowering the acetate-to-propionate ratio and acetate-butyrate-to-propionate ratio compared with other treatments. Increasing the propionate concentration could explain the high nutrient digestibility (Table 3) found in FDPML. In addition, a high lactic acid concentration in FDPML may contribute to an increase in the propionate concentration as lactic acid is

biologically converted into propionate by ruminal microbes in the rumen. Similarly, So et al. [7] showed that L. casei TH14 combined with cellulase- and molasses-treated sugarcane bagasse in TMR significantly increased the propionate concentration compared with untreated sugarcane bagasse in dairy cows. Zhang et al. [36] revealed that whole-corn plant treated with complex inoculants significantly increased the propionate concentration from 6.40 mmol/L to 8.98 mmol/L in growing-finishing cattle. Cherdthong et al. [13] similarly found that rice straw treated with L. casei TH14 and molasses fed to Thai Native beef cattle significantly increased their propionate concentration from 20.3 mol/100 mol to 23.2 mol/100 mol compared with untreated rice straw. Estimated CH₄ production was significantly lower in FDPML. The reason could be explained by the increase in propionate concentration found in FDPML as hydrogen was used for propionate synthesis, resulting in less hydrogen available in the methanogenesis pathway of methanogen bacteria to produce CH_4 as the main end-product [39]. Similarly, So et al. [7] showed that sugarcane bagasse treated with L. casei TH14 combined with cellulase and molasses in TMR fed to dairy cows significantly decreased CH₄ production by 4%. Monteiro et al. [40] tested L. plantarum as direct-fed microbial in high-producing cows and similarly found decreased in CH₄ production compared with no additive treatment.

Dietary treatments did not affect bacteria, fungal zoospore, total protozoa, *Holotrich* sp., or *Entodiniomorphs* sp. populations. Ruminal bacteria favor a pH around 7 for optimum growth [35]; this study found the pH ranged from 6.44 to 6.71, which may have contributed to the unchanged bacteria population. Similarly, Bureenok et al. [41] showed that ruzi grass ensiled with molasses or fermented juice of epiphytic lactic acid bacteria fed to cows separately or as a combination did not change ruminal bacteria. However, not all previous studies found unchanged ruminal bacteria when inoculants wer eused. Cherdthong et al. [13] found that rice straw treated with molasses and *L. casei* TH14 combined fed to Thai Native beef cattle significantly increased ruminal bacteria population; the changed in bacteria population may have been due to the optimum ruminal pH ranging from 7.0 to 7.1 for bacteria growth.

Blood metabolites including glucose, pack cell volume, blood urea nitrogen, total protein, albumin, globulin, the albumin-to-globulin ratio, red blood cell, hemoglobin, mean corpuscular volume, mean corpuscular hemoglobin concentration, RBC distribution width, white blood cells, and lymphocytes were similar among dietary treatments. This suggests that the goats were in good health and had a normal metabolism status. Blood metabolites are usually used to evaluate the nutritious plane and health status in ruminants [42,43]. As well as blood metabolites, glucose, blood urea nitrogen, total protein, and albumin concentration were commonly used to evaluate protein and carbohydrate metabolism, where the higher mean value suggests a better nutrient metabolism when these metabolites changed within a normal range [44]. Liver is the main hub where glucose, albumin, and blood urea nitrogen are synthesized [45,46], and glucose and albumin concentrations were greater in goats fed FDPML. Mean glucose concentration ranged from 68.70 to 70.20 mg/dL, which varied in a normal range of 50 to 75 mg/dL [47]. Blood urea nitrogen is the product of NH₃–N recycling and is produced from protein degradation by ruminal microbes [48,49]. The lower blood urea nitrogen paralleled the lower NH₃–N concentration found in goats fed FDPML.

Dietary treatments did not influence nitrogen intake, fecal nitrogen, or apparent nitrogen retention, expressed as g/d or% of nitrogen intake in goats. Urine nitrogen was significantly lower, at 11%, in FDPC and FDPML when compared with FDP. The effect of cellulase or a combination of molasses and *L. casei* TH14 on urinary nitrogen reduction was unknown; it may possibly be inconsistent with retained nitrogen, as it was found to be the highest in FDPC and FDPML. Cherdthong et al. [13] showed that rice straw treated with cellulase or *L. casei* TH14 separately or as a combination fed to Thai Native beef cattle reduced nitrogen loss both in the urine and feces but failed to reach statistical significance.

5. Conclusions

This study showed that discarded durian peel fermented with a combination molasses and *L. casei* TH14 (FDPML) had significantly greater nutrient digestibility and propionate concentration, while estimated methane production, the acetate-to-propionate ratio and urinary nitrogen decreased when compared with untreated discarded durian peel (FDP). Therefore, a combination treated discarded durian peel with molasses and *L. casei* TH14 could add 25% of dry matter to the diet for growing goats without a negative impact. Further studies should evaluate the effect of fermented discarded durian peel with an additive content of higher than 25% in the diet on feed utilization, digestibility, rumen characteristics, blood metabolites, and nitrogen balance in ruminants.

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References

- Nordin, N.; Shamsudin, R.; Azlan, A.; Effendy, M. Dry matter, moisture, ash and crude fibre content in distinct segments of 'Durian Kampung' husk. Int. J. Chem. Eng. 2017, 11, 788–792.
- 2. Waramit, W.; Phuangborisut, S.; Wetchagool, W.; Wetchagool, N.; Phattapanit, V. Effect of dietary substitution of durian seed starch for broken rice on productive performance in broiler. *Prawarun Agric. J.* **2016**, *13*, 145–152.
- 3. Nuraini, A.D.; Mahata, M.E. Improving the nutrient quality of durian (*Durio zibethinus*) fruit waste through fermentation by using *Phanerochaete chrysosporium* and *Neurospora crassa* for poultry diet. *Int. J. Poult. Sci.* **2015**, *14*, 354–358.
- Khota, W.; Pholsen, S.; Higgs, D.; Cai, Y. Fermentation quality and in vitro methane production of sorghum silage prepared with cellulase and lactic acid bacteria. *Asian-Australas. J. Anim. Sci.* 2017, 30, 1568–1574. [CrossRef] [PubMed]
- So, S.; Cherdthong, A.; Wanapat, M. Improving sugarcane bagasse quality as ruminant feed with *Lactobacillus*, cellulase, and molasses. J. Anim. Sci. Technol. 2020, 62, 648–658. [CrossRef]
- So, S.; Cherdthong, A.; Wanapat, M.; Uriyapongson, S. Fermented sugarcane bagasse with *Lactobacillus* combined with cellulase and molasses promotes in vitro gas kinetics, degradability, and ruminal fermentation patterns compared to rice straw. *Anim. Biotechnol.* 2020, 18, 1–12. [CrossRef]
- So, S.; Wanapat, M.; Cherdthong, A. Effect of sugarcane bagasse as industrial by-products treated with *Lactobacillus casei* TH14, cellulase and molasses on feed utilization, ruminal ecology and milk production of mid-lactating Holstein Friesian cows. *J. Sci. Food Agric.* 2021, 101, 481–4489. [CrossRef]
- So, S.; Cherdthong, A.; Wanapat, M. Growth performances, nutrient digestibility, ruminal fermentation and energy partition of Thai native steers fed exclusive rice straw and fermented sugarcane bagasse with *Lactobacillus*, cellulase and molasses. *J. Anim. Physiol. Anim. Nutr.* 2021, 106, 45–54. [CrossRef]
- Pholsen, S.; Khota, W.; Pang, H.; Higgs, D.; Cai, Y. Characterization and application of lactic acid bacteria for tropical silage preparation. *Anim. Sci. J.* 2016, 87, 1202–1211. [CrossRef]

- Wang, S.; Guo, G.; Li, J.; Chen, L.; Dong, Z.; Shao, T. Improvement of fermentation profile and structural carbohydrate compositions in mixed silages ensiled with fibrolytic enzymes, molasses and *Lactobacillus plantarum* MTD-1. *Ital. J. Anim. Sci.* 2018, *18*, 328–335. [CrossRef]
- 11. Wang, S.; Li, J.; Dong, Z.; Chen, L.; Shao, T. Effect of microbial inoculants on the fermentation characteristics, nutritive value, and *in vitro* digestibility of various forages. *Anim. Sci. J.* **2019**, *90*, 178–188. [CrossRef] [PubMed]
- 12. Cherdthong, A.; Suntra, C.; Khota, W. Improving nutritive value of ensiled rice straw as influenced by *Lactobacillus casei*. *Khon Kaen Agric. J.* **2019**, *47*, 105–110.
- Cherdthong, A.; Suntara, C.; Khota, W.; Wanapat, M. Feed utilization and rumen fermentation characteristics of Thai-indigenous beef cattle fed ensiled rice straw with *Lactobacillus casei* TH14, molasses, and cellulase enzymes. *Livest. Sci.* 2021, 245, 104405. [CrossRef]
- Chen, X.Z.; Li, W.Y.; Gao, C.F.; Zhang, X.P.; Weng, B.Q.; Cai, Y.M. Silage preparation and fermentation quality of kudzu, sugarcane top and their mixture treated with lactic acid bacteria, molasses and cellulase. *Anim. Sci. J.* 2017, *88*, 1715–1721. [CrossRef] [PubMed]
- 15. AOAC. Official Methods of Analyses, 15th ed.; Association of Official Analytical Chemists: Arlington, VA, USA, 1990.
- 16. Van Soest, P.J.; Robertson, J.B.; Lewis, B.A. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *J. Dairy Sci.* **1991**, *74*, 3583–3597. [CrossRef]
- 17. Mertens, D.R. Creating a system for meeting the fiber requirements of dairy cows. J. Dairy Sci. 1997, 80, 1463–1481. [CrossRef]
- 18. NRC. Nutrient Requirements of Goats: Angora, Dairy and Meat Goats in Temperate and Tropical Countries; National Academy Press: Washington, DC, USA, 1981.
- 19. Bremner, J.M.; Keeney, D.R. Steam distillation methods for determination of ammonium, nitrate and nitrite. *Anal. Chim. Act.* **1965**, *32*, 485–495. [CrossRef]
- Osaki, T.Y.; Kamiya, S.; Sawamura, S.; Kai, M.; Ozawa, A. Growth inhibition of *Clostridium difficile* by intestinal flora of infant faeces in continuous flow culture. J. Med. Microbiol. 1994, 40, 179–187. [CrossRef]
- Moss, A.R.; Jouany, J.P.; Newbold, J. Methane production by ruminants: Its contribution to global warming. *Annales Zootech.* 2000, 49, 231–2353. [CrossRef]
- 22. Galyean, M. Laboratory Procedures in Animal Nutrition Research; New Mexico State University: Las Cruces, NM, USA, 1989.
- 23. Kearl, L.C. *Nutrient Requirements of Ruminants in Developing Countries;* International Feedstuffs Institute, Utah State University: Logan, UT, USA, 1982.
- Oba, M.; Allen, M.S. Effects of brown midrib 3 mutation in corn silage on productivity of dairy cows fed two concentrations of dietary neutral detergent fiber: 1. Feeding behavior and nutrient utilization. J. Dairy Sci. 2000, 83, 1333–1341. [CrossRef]
- 25. Kung, L., Jr.; Shaver, R.D.; Grant, R.J.; Schmidt, R.J. Silage review: Interpretation of chemical, microbial, and organoleptic components of silages. *J. Dairy Sci.* 2018, 101, 4020–4033. [CrossRef]
- Addah, W.; Baah, J.; Okine, E.K.; McAllister, T.A. A third-generation esterase inoculant alters fermentation pattern and improves aerobic stability of barley silage and the efficiency of body weight gain of growing feedlot cattle. J. Anim. Sci. 2012, 90, 1541–1552. [CrossRef] [PubMed]
- Islam, M.; Enishi, O.; Purnomoadi, A.; Higuchi, K.; Takusari, N.; Terada, F. Energy and protein utilization by goats fed Italian ryegrass silage treated with molasses, urea, cellulase or cellulase + lactic acid bacteria. *Small Rumin. Res.* 2001, 42, 49–60. [CrossRef]
- Yuan, X.; Guo, G.; Wen, A.; Desta, S.T.; Wang, J.; Wang, Y.; Shao, T. The effect of different additives on the fermentation quality, *in vitro* digestibility and aerobic stability of a total mixed ration silage. *Anim. Feed Sci. Technol.* 2015, 207, 41–50. [CrossRef]
- Zhao, J.; Dong, Z.; Li, J.; Chen, L.; Bai, Y.; Jia, Y.; Shao, T. Effects of lactic acid bacteria and molasses on fermentation dynamics, structural and nonstructural carbohydrate composition and in vitro ruminal fermentation of rice straw silage. *Asian-Australas. J. Anim. Sci.* 2019, 32, 783–791. [CrossRef] [PubMed]
- 30. Sari, N.F.; Ridwan, R.; Fidriyanto, R.; Astuti, W.D.; Widyastuti, Y. The Effect of probiotics on high fiber diet in rumen fermentation characteristics. In *IOP Conference Series: Earth and Environmental Science*; IOP Publishing: Bristol, UK, 2019; Volume 251, p. 012057.
- Dagaew, G.; Cherdthong, A.; Wanapat, M.; So, S.; Polyorach, S. Ruminal fermentation, milk production efficiency, and nutrient digestibility of lactating dairy cows receiving fresh cassava root and solid feed-block containing high sulfur. *Fermentation* 2021, 7, 114. [CrossRef]
- 32. Seankamsorn, A.; Cherdthong, A.; So, S.; Wanapat, M. Influence of chitosan sources on intake, digestibility, rumen fermentation, and milk production in tropical lactating dairy cows. *Trop. Anim. Health Prod.* **2021**, *53*, 1–9. [CrossRef]
- 33. Seankamsorn, A.; Cherdthong, A.; So, S.; Wanapat, M. Using glycerin with chitosan extracted from shrimp residue to enhance rumen fermentation and feed use in native Thai bulls. *Vet. World* **2021**, *14*, 1158–1164. [CrossRef]
- 34. Dehority, B.A.; Tirabasso, P.A. Effect of feeding frequency on bacterial and fungal concentrations, pH, and other parameters in the rumen. *J. Anim. Sci.* 2001, *79*, 2908–2912. [CrossRef]
- 35. Weimer, P.J. Why don't ruminal bacteria digest cellulose faster? J. Dairy Sci. 1996, 79, 1496–1502. [CrossRef]
- Zhang, Y.; Zhao, X.; Chen, W.; Zhou, Z.; Meng, Q.; Wu, H. Effects of adding various silage additives to whole corn crops at ensiling on performance, rumen fermentation, and serum physiological characteristics of growing-finishing cattle. *Animals* 2019, 9, 695. [CrossRef] [PubMed]

- 37. Seo, J.K.; Kim, S.W.; Kim, M.H.; Upadhaya, S.D.; Kam, D.K.; Ha, J.K. Direct-fed microbials for ruminant animals. *Asian-Australas. J. Anim. Sci.* **2010**, *23*, 1657–1667. [CrossRef]
- McDonald, P.; Edwards, R.A.; Greenhalgh, J.F.D.; Morgan, C.A.; Sinclair, L.A.; Wilkinson, R.G. Animal Nutrition, 7th ed.; Prentice Hall: Harlow, UK, 2012.
- Doyle, N.; Mbandlwa, P.; Kelly, W.J.; Attwood, G.; Li, Y.; Ross, R.P.; Stanton, C.; Leahy, S. Use of lactic acid bacteria to reduce methane production in ruminants: A critical review. *Front. Microbiol.* 2019, *10*, 2207. [CrossRef]
- Monteiro, H.F.; Lelis, A.L.; Brandao, V.L.; Faccenda, A.; Avila, A.S.; Arce-Cordero, J.; Silva, L.G.; Dai, X.; Restelatto, R.; Carvalho, P.; et al. *In vitro* evaluation of *Lactobacillus plantarum* as direct-fed microbials in high-producing dairy cows diets. *Transl. Anim. Sci.* 2020, 4, 214–228. [CrossRef]
- Bureenok, S.; Suksombat, W.; Kawamoto, Y. Effects of the fermented juice of epiphytic lactic acid bacteria (FJLB) and molasses on digestibility and rumen fermentation characteristics of ruzigrass (*Brachiaria ruziziensis*) silages. *Livest. Sci.* 2011, 138, 266–271. [CrossRef]
- 42. Nozad, S.; Ramin, A.G.; Moghadam, G.; Asri-Rezaei, S.; Babapour, A.; Ramin, S. Relationship between blood urea, protein, creatinine, triglycerides and macro-mineral concentrations with the quality and quantity of milk in dairy Holstein cows. In *Veterinary Research Forum*; Faculty of Veterinary Medicine, Urmia University: Urmia, Iran, 2012; Volume 3, p. 55.
- Mohammadi, V.; Anassori, E.; Jafari, S. Measure of energy related biochemical metabolites changes during peri-partum period in Makouei breed sheep. In *Veterinary Research Forum*; Faculty of Veterinary Medicine, Urmia University: Urmia, Iran, 2016; Volume 7, p. 35.
- 44. Russell, K.E.; Roussel, A.J. Evaluation of the ruminant serum chemistry profile. *Vet. Clin. N. Am. Food Anim. Pract.* 2007, 23, 403–426. [CrossRef]
- Cozzi, G.; Ravarotto, L.; Gottardo, F.; Stefani, A.L.; Contiero, B.; Moro, L.; Brscic, M.; Dalvit, P. Reference values for blood parameters in Holstein dairy cows: Effects of parity, stage of lactation, and season of production. *J. Dairy Sci.* 2011, 94, 3895–3901. [CrossRef]
- Chanjula, P.; Ngampongsai, W.; Wanapat, M. Effects of replacing ground corn with cassava chip in concentrate on feed intake, nutrient utilization, rumen fermentation characteristics and microbial populations in goats. *Asian-Australas. J. Anim. Sci.* 2007, 20, 1557–1566. [CrossRef]
- 47. Kaneko, J.J. Appendixes. In *Clinical Biochemistry of Domestic Animals*, 3rd ed.; Kaneko, J.J., Ed.; Academic Press: New York, NY, USA, 1980.
- 48. Chanjula, P.; Pakdeechanuan, P.; Wattanasit, S. Effects of dietary crude glycerin supplementation on nutrient digestibility, ruminal fermentation, blood metabolites, and nitrogen balance of goats. *Asian-Australas. J. Anim. Sci.* **2014**, *27*, 365–374. [CrossRef]
- Chanjula, P.; Raungprim, T.; Yimmongkol, S.; Poonko, S.; Majarune, S.; Maitreejet, W. Effects of elevated crude glycerin concentrations on feedlot performance and carcass characteristics in finishing steers. *Asian-Australas. J. Anim. Sci.* 2016, 29, 80–88. [CrossRef]