



# Fast, Sensitive and Robust Determination of Nicotinic Acid (Vitamin B<sub>3</sub>) Contents in Coffee Beverages Depending on the Degree of Roasting and Brewing Technique <sup>†</sup>

Leon Buckel, Jonathan Isaak Kremer, Simone Stegmüller and Elke Richling \*

Department of Food Chemistry and Toxicology Technische Universität Kaiserslautern, Erwin-Schroedinger-Str. 52, 67663 Kaiserslautern, Germany; buckel@rhrk.uni-kl.de (L.B.); kremer@chemie.uni-kl.de (J.I.K.); stegmueller@chemie.uni-kl.de (S.S.)

\* Correspondence: richling@chemie.uni-kl.de

<sup>†</sup> Presented at Natural Products and the Hallmarks of Chronic Diseases—COST Action 16112, Luxembourg, 25–27 March 2019.

Published: 16 April 2019

**Abstract:** The vitamin B<sub>3</sub> (niacin) is present in various foods. During roasting of green coffee beans, niacin is formed from the alkaloid trigonelline. Therefore, we established a novel fast and sensitive HPLC-MS/MS method to determine niacin in coffee brews from five commercially available coffee samples. Additionally, we investigated the influence of the brewing method, brewing temperature, and degree of roasting on niacin contents. In the respective coffee brews, we were able to show that the content of niacin in coffee beverages is not only affected by the degree of roasting, but also by the extraction performance of different brewing methods to a lesser extent.

**Keywords:** niacin; coffee; HPLC-ESI-MS/MS; vitamin B<sub>3</sub>; stable isotope dilution analysis (SIDA)

## 1. Introduction

Vitamin B<sub>3</sub> (nicotinic acid (NA) and nicotinamide (NAM), known as niacin) is present in various foods. NA and NAM play an important role in physiology. Niacin is a part of the cofactors nicotinamide adenine dinucleotide and nicotinamide adenine dinucleotide phosphate [1,2]. The recommended daily intake of niacin is about 15 mg/day [3], but it is a semi-essential vitamin. It can be formed from the amino acid tryptophan in the human body, with approximately 60 mg of tryptophan being equivalent to 1 mg NA [4]. During roasting of green coffee beans, NA is formed from the alkaloid trigonelline. Powdered coffee contains up to 300 mg/kg of NA depending on the degree of roasting. Nevertheless, in literature, information on the NA contents of commercially available coffee samples as well as the influence of brewing techniques on the NA content in prepared beverages is rather scarce. Therefore, we developed a fast and sensitive HPLC-MS/MS method to determine NA in coffee beverages. Additionally, the influence of brewing methods, degree of roasting, and brewing temperature was investigated.

## 2. Materials and Methods

### 2.1. Materials

Nicotinic acid (NA) and d<sub>4</sub>-nicotinic acid (d<sub>4</sub>-NA) were obtained from Toronto Research Chemicals (Toronto, Canada). Acetonitrile HPLC grade was acquired from VWR (Darmstadt, Germany). Formic acid p.a. was purchased from Sigma-Aldrich (Taufkirchen, Germany).

## 2.2. Preparation of the Coffee Beverages

Five commercially available coffee pad samples of ground coffee from the same manufacturer, containing approximately 7.4 g, (mild (taste intensity = 2), classic (taste intensity = 3), extra (taste intensity = 4), Espresso (taste intensity = 5), and decaffeinated (taste intensity = 4)) were purchased in a local supermarket in Germany. The “taste intensity”-value on a scale of 1 to 5 were provided on the packing box from the manufacturer to characterize the consumer expectation and is mainly influenced by the degree of roasting. All coffees were 100% arabica. After opening coffee powders were completely emptied in all cases, except in tests with the pad machine, and weighted. Coffee beverages were prepared in agreement with the described methods [5] using the following machines: a standard “drip” coffee maker (Tchibo, Hamburg, Germany) fitted with filter paper, a Senseo coffee pad machine (Philips, Amsterdam, Netherlands) and an automatic coffee maker (Bosch, Gerlingen, Germany). Other samples were also prepared using: a manual plastic coffee filter with filter paper and boiling water; a pot for the “Turkish” coffee (powder and water heated together); a “French press” (Bodum AG, Triengen, Switzerland) and additionally, a so-called cold brew was prepared in the refrigerator for 24 h at +4 °C. For each method, triplicates were prepared and stored at −20 °C until the analysis. To investigate the influence of brewing temperatures, the ground coffee from the “classic” pads was emptied and brewed by hand filter at temperatures of 40, 60, 80, 90, and 100 °C.

## 2.3. Sample Preparation

Aliquots ( $n = 3$ ) of the coffee beverages were diluted 1:100, in the case of the coffee brew prepared from “mild” or “decaffeinated” coffee, as well as prepared at a temperature of 40 °C, the samples were diluted 1:50. The diluted coffee samples were spiked with an isotopically labelled standard  $d_4$ -NA to achieve a final concentration of 800 nM in each sample. After equilibration, the samples were passed through a syringe filter (Chromafil AO-45/25, Polyamid, 0.45 µm, Macherey-Nagel, Düren, Germany) and a 2 µL aliquotes were analyzed.

## 2.4. HPLC-ESI-MS/MS Analysis

HPLC-ESI-MS/MS system (1200 series, Agilent, Waldbronn, Germany and API 3200 triple quadrupole mass spectrometer from Applied Biosystems, Darmstadt, Germany) was used. NA was analyzed using a VDSpher PUR HILIC-Z column (100 × 2.0 mm, 100 Å, 5.0 µm, VDS optilab, Berlin, Germany) at 20 °C using 0.01% aqueous formic acid (eluent A) and acetonitrile (eluent B) at a flow rate of 1000 µL/min. The injection volume was 2 µL. Gradient started at 90% eluent B, isocratic for 0.5 min, then eluent B was reduced to 10% within 0.1 min. From minute 0.6 to 4.0, the system was kept isocratic at 10% eluent B. Afterwards the column was washed and equilibrated. The ESI-source was operated in positive ion mode (5.500 V), and nitrogen was used as nebulizer gas (55 psi), heater gas (65 psi, 350 °C), curtain gas (25 psi), and collision gas (2.0 psi). Nicotinic acid and  $d_4$ -NA were detected in multiple reaction monitoring mode. MS transitions were:  $m/z$  124→80 as qualifier and  $m/z$  124→78 as quantifier and the isotopically labelled analogue  $m/z$  128→84, respectively.

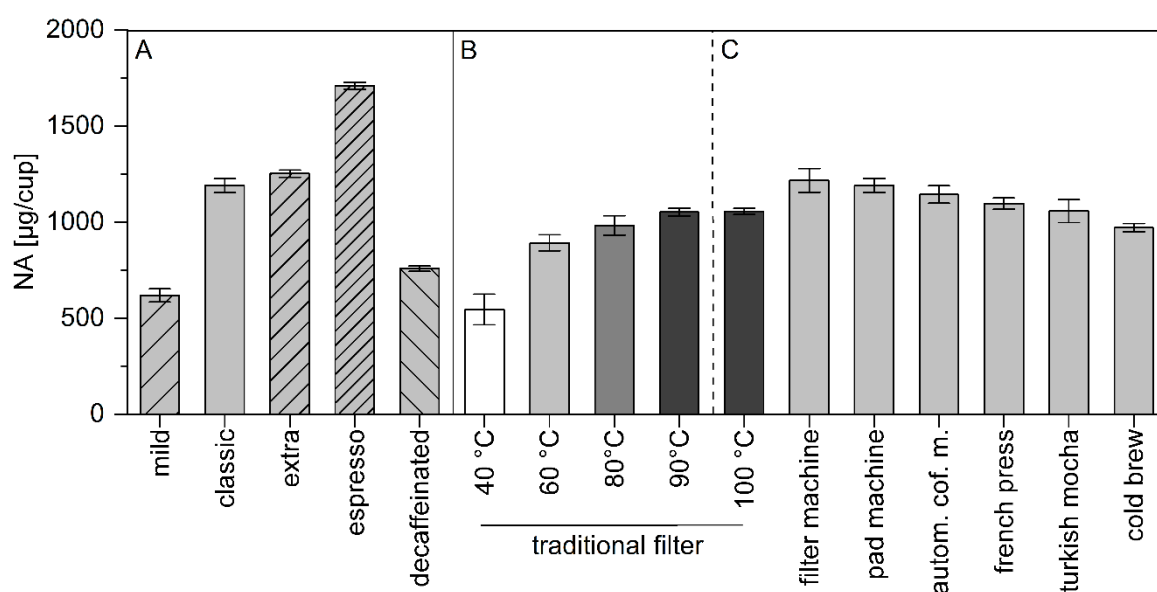
## 2.5. Quantitative Analysis and Method Validation

To quantitate NA mixtures at eight analyte/internal standard mole ratios in the range of concentrations from 80 to 10000 nM for each analyte and at 800 nM for the associated internal standard  $d_4$ -NA were analyzed by HPLC-MS/MS. The ratios of the peak areas of the ions selected for quantitation were plotted versus the weight ratio of analyte/internal standard. The linear regression coefficient ( $R^2$ ) of the calibration curve was >0.99. Limits of detection and quantification, defined as signal to-noise ratios of 3:1 and 10:1 [6], were 14.7 nM and 77.5 nM, respectively. Recovery rates were determined using 640, 800, and 960 nM of NA as well as the respective IS and were between 91 and 96%. The established HPLC-ESI-MS/MS method was fast and sensitive enough to determine nicotinic acid in coffee brews within 4 min.

Data were processed by Analyst 1.6 (AB Sciex, Darmstadt, Germany). Experimental results are reported as means of at least three independent preparations with ±SD as error bars.

### 3. Results

From the five commercially available coffee samples under study the respective brews were prepared by a pad machine and NA was quantitated. To observe the influence of the brewing method, a Senseo coffee pad machine using pads named “classic”, and the coffee powder from those pads was used to brew coffee by traditional filter, standard “drip” coffee maker, automatic coffee maker, manual plastic coffee filter with filter paper, “Turkish” coffee pot, “French press”, and so called cold brew method. Figure 1 represents the calculated quantities of NA extracted from the powdered coffee [ $\mu\text{g}/\text{cup}$  of coffee].



**Figure 1.** Nicotinic acid (NA) contents [ $\text{mg}/\text{cup}$ ] of (A) five different coffee samples (all 100% arabica) prepared by pad machine ( $\emptyset$  cup volume 112 mL); (B) coffee sample brewed at different temperatures with traditional filter method (all  $n = 3$ ); (C) the sort “classic” prepared by different brewing techniques: filter machine ( $\emptyset$  cup volume 112 mL), automatic coffee maker ( $\emptyset$  cup volume 110 mL), French press ( $\emptyset$  cup volume 98 mL), Turkish mocha ( $\emptyset$  cup volume 71 mL), cold brew ( $\emptyset$  cup volume 96 mL) and traditional filter ( $\emptyset$  cup volume 116 mL).

It is demonstrated that the degree of roasting seems to influence the amount of NA: with higher roasting degree, the amount of NA is increased. Therefore, “espresso” showed the highest NA content with  $1711 \pm 18 \mu\text{g}/\text{cup}$  of coffee. The influence of the preparation seems to be minor. Here the automatic coffee maker and filter machine revealed highest amounts of NA ( $1144 \pm 46$  and  $1217 \pm 62 \mu\text{g}/\text{cup}$ ), respectively. The lowest amount was determined in the cold brew with  $972 \pm 21 \mu\text{g}/\text{cup}$  NA.

One sample, namely the “classic” pad was used to observe the influence of brewing temperature on the NA transfer into the coffee brews. Water temperatures of 40, 60, 80, 90 and 100 °C were used to prepare traditional filtered coffee. Whereas at 40 °C the amount was only  $545 \pm 80 \mu\text{g}/\text{cup}$ , it increased with higher brewing temperatures:  $892 \pm 41$  at 60 °C,  $983 \pm 41$  at 80 °C,  $1052 \pm 20$  at 90 °C and  $1056 \pm 17 \mu\text{g}/\text{cup}$  at 100 °C. At temperatures higher than 60 °C, the NA content of coffee beverages was almost independent of the brewing temperature reaching a plateau at 90 to 100 °C.

### 4. Discussion

We were able to show, that the NA amount in coffee is dependent on the degree of roasting. These results are in agreement with the study by Lang et al., who investigated the influence of different roasting conditions on NA formation. The authors found that hotter or longer roasted coffee beans contain more NA [7]. Additionally, the brewing method seems to influence the amount of NA transferred into the respective brews. This can depend on the water temperature used to prepare the coffees. Here it was shown that the maximum amount of NA is liberated at water temperatures above

80 °C. In contrast, it was shown that the duration of extraction has an influence on the amount of extractable NA. With the cold brew method (4 °C for 24 h) it is possible to obtain about  $972 \pm 21$  µg/cup NA in contrast to  $545 \pm 80$  µg/cup NA with traditional filter at 40 °C. However, even the choice of brewing method itself can have an influence on the extractable amount of NA. Thus, mechanical methods that allow continuous extraction (e.g., pad, filter and automatic coffee maker) show a higher yield than discontinuous methods (e.g., French press and Turkish mocha).

The results are presented in µg NA extracted per cup of coffee, accounting for up to 11 mg NA per liter, present in the coffee brew prepared by the coffee pad machine from the “classic” coffee sample. This corresponds to 1.2 mg per 125 mL cup covering about 9% of the recommended daily intake (RDA) of 15 mg NA. [3] It could be shown that coffee consumption contributes to the daily niacin (vitamin B<sub>3</sub>) intake.

**Author Contributions:** Conceptualization, E.R.; investigation, L.B.; validation, L.B.; formal analyzes, L.B.; methodology, L.B., J.I.K. and S.S.; project administration, E.R.; supervision, E.R.; writing—original draft preparation, E.R.; writing—review and editing, J.I.K., S.S. and E.R.

**Funding:** This article is based upon work from COST Action NutRedOx-CA16112 supported by COST (European Cooperation in Science and Technology).

**Acknowledgments:** We thank Tchibo GmbH, Hamburg, Germany for some financial support to buy the isotopically labelled standard.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Bogan, K.L.; Brenner, C. Nicotinic acid, nicotinamide, and nicotinamide riboside: A molecular evaluation of NAD<sup>+</sup> precursor vitamins in human nutrition. *Ann. Rev. Nutr.* **2008**, *28*, 115–130, doi:10.1146/annurev.nutr.28.061807.155443.
2. Sauve, A.A. NAD<sup>+</sup> and vitamin B<sub>3</sub>: From metabolism to therapies. *J. Pharmacol. Exp. Therap.* **2008**, *324*, 883–893, doi:10.1124/jpet.107.120758.
3. Scientific Opinion on Dietary Reference Values for niacin. *EFSA J.* **2014**, *12*, doi:10.2903/j.efsa.2014.3759.
4. Horwitt, M.K.; Harper, A.E.; Henderson, L.M. Niacin-tryptophan relationships for evaluating niacin equivalents. *Am. J. Clin. Nutr.* **1981**, *34*, 423–427.
5. Deutscher Kaffeeverband e.V. *Moderne Zubereitung von Kaffee in der Gastronomie*; 2nd ed.; jungvornweg: Dresden, Germany, 2016.
6. MacDougall, D.; Crummett, W.B.; Amore, F.J.; Cox, G.V.; Crosby, D.G.; Estes, F.L.; Lal, J.; Langner, R.R.; McClelland, N.I.; Phillips, W.F.; et al. Guidelines for data acquisition and data quality evaluation in environmental chemistry. *Anal. Chem.* **1980**, *52*, 2242–2249, doi:10.1021/ac50064a004.
7. Lang, R.; Yagar, E.F.; Eggers, R.; Hofmann, T. Quantitative investigation of trigonelline, nicotinic acid, and nicotinamide in foods, urine, and plasma by means of LC-MS/MS and stable isotope dilution analysis. *J. Agric. Food Chem.* **2008**, *56*, 11114–11121, doi:10.1021/jf802838s.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).