

Article Paper Foxing Stains on a Historic Manuscript from the Early Qajar Era: Abiotic or Biotic Foxing?

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Abstract: The aim of this study was to identify the nature and cause of foxing spots in a historical manuscript. This manuscript was a Holy Quran from the beginning of the Qajar period and the end of the 18th century. Samples were incubated for 14 days and were evaluated for the presence of fungal activity. UV fluorescence photography, micro X-ray fluorescence spectroscopy and Fourier transform infrared spectroscopy were also used to investigate the characteristics and causes of foxing spots. The results showed that there was no fungal activity in the foxing spots of this manuscript. Based on the morphology of the stain in UV fluorescence photography, these foxing stains are of the Bullseye type, usually associated with metal ions. μ XRF spectroscopy also showed a high accumulation of iron and copper at the site of these spots. This indicates abiotic foxing in this manuscript. Based on FTIR spectroscopy and peak deconvolution and fitting by Gaussian function, abiotic foxing increases the cellulose oxidation rate. Intensification of cellulose oxidation in foxing stains can be considered as one of the reasons for paper discoloration.

Keywords: historical manuscript conservation; historical paper; foxing; cellulose oxidation; paper biodeterioration

1. Introduction

Paper is one of the most commonly used materials in historical objects. Today, numerous paper-based artifacts can be found in museums, libraries and archives. However, paper is a material that easily undergoes chemical, mechanical and biodegradation, especially if not stored under controlled environmental conditions. The reddish-brown, brown or yellowish irregular spots are one important aspect of deterioration observed on historical papers [1]. The formation of these stains, called foxing, has been reported on historical paper materials for decades [2]. These stains can be seen on many types of historical papers produced after the 16th century [3]. In the foxing area, the paper becomes weaker, friable, brittle and more acidic than the unaffected part [4]. Foxing spots spread on paper over time, which can be monitored using photography and image processing techniques [5]. In addition to paper materials, foxing stains are also common in textile collections [6].

The first use of the term foxing dates back to 1848 and originated from the rusty red color of fox [7]. The term foxing refers to roundish small stains of yellowish or reddishbrown color, which are found on paper or other cellulosic fiber materials. This vague definition sometimes makes it difficult to identify foxing [8]. In other words, spots with different formation mechanisms, but similar appearance, can be classified as foxing.

The study of foxing has been one of the topics of interest due to its complexity, destructiveness and related unknown mechanism. Research about foxing has been established since the 1930s [8]. This damage has been reported on old books, postage stamps, archive documents, the printed photo, drawings, pigment-coated papers, etc. [1–3,9,10]. Studies on foxing have been carried out from various points of view, but to date, its exact mechanism is not known to researchers [11].



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Nonetheless, various hypotheses have been put forward about the reason for the formation of foxing in historical papers which are as follows:

- Foxing stains are the result of an activity of microorganisms, fungi [2,8,12,13] and bacteria [10], which is called the biotic theory [8,12,14].
- Foxing stains are the result of the presence of metal deposits such as iron and copper and chemical reactions such as oxidation, which is called the abiotic theory [8,12,14,15].
- According to reports, the simultaneous presence and activity of microorganisms and metal inclusions is another reason for the formation of foxing spots [8].
- In addition, the accumulation of moisture in parts of the paper increases the automatic oxidation of cellulose, which causes discoloration in the dry/wet interface at the paper surface [4,8].

Therefore, the study of foxing stains, due to their complex nature, is still a key issue in the preservation of historical papers. Thus, the aim of this study was to identify the cause and nature of foxing stains and the degree of degradation caused by them in the cellulose structure. For this purpose, a manuscript of the Quran related back to the early Qajar period was studied. Certainly, diagnosing the nature of the foxing spot and the damage it has caused can be helpful in choosing a suitable treatment method.

2. Materials and Methods

2.1. Sample Description

The investigation was focused on a historical manuscript of the Holy Quran belonging to a private collection. In some pages of this manuscript, a seal with the date 1210 HA (1795–1796 AD) can be seen. This date shows that this manuscript belongs to the first years of the Qajar era. Brown-orange spots, foxing, can be seen on some pages of this manuscript. Additionally, a page shows black spots that may be caused by the activity of microorganisms (Figure 1).



Figure 1. (**A**) The manuscript of the Holy Quran from the late 18th century and the types of stains studied: (**B**) foxing spots, (**C**) fungal colony.

2.2. Isolation and Identification of Fungi

A sample of black spots and five samples of foxed spots were studied to investigate the possibility of the presence of fungi. Fungi were cultivated in sabouraud dextrose agar (SDA; Merck, Germany) medium which was autoclaved at 121 °C for 15 min to sterilize. Samples were incubated for 14 days at 28 \pm 2 °C and 70 \pm 5% RH. Cultivated fungi were characterized according to their macroscopic and microscopic properties, using an Olympus BX51 microscope.

2.3. µ-XRF Spectroscopy

The qualitative concentration of the elements in the two foxed and adjacent unfoxed areas was investigated using a micro XRF. An Unisantis 104 X-ray micro fluorescence spectrometer was used for XRF experiments. The excitation settings are 25 kV and 300 mA/120 s.

2.4. FTIR Spectroscopy

FTIR analysis was carried out using a Nicolet 680Plus FTIR spectrometer (Jasco, Japan) with KBr pellets. One sample of paper without foxing and two samples with foxing stains (S1 and S2) were examined. All Spectra were collected in the range of 400–4000 cm⁻¹ at 4 cm⁻¹ resolution with 32 numbers of scan. After modifying the baseline in the range of 850 to 1800 cm⁻¹, the Gaussian function was used to peaks deconvolution and fitting.

2.5. UV Fluorescence Photography

The images were taken with a Nikon D750 DSLR camera equipped with an AF Nikkor 50 mm f/1.8D lens. UV fluorescence (UVF) photography was performed in a 45° setup by two xenon flash light with B + W UV-Pass filter, used as a radiation source, and a Baader UV/IR cut filter mounted on top of the camera lens.

3. Results and Discussion

First, the presence of fungus activity in foxing spots was investigated using cultivation. After the incubation period, no fungal growth was observed in the foxing spots. In fact, it can be said that fungi are not the reason for the formation of foxing stains in this manuscript. However, in one of the manuscript sheets (Figure 1C), which was without foxing spots, the presence and growth of the fungus was observed. This fungus' morphological features with fast-growing and deeply cottony texture of the colony having a white to gray color and dark pigment sporangium are shown in Figure 2a. Microscopic features show transparent and non-septate mycelium. The non-branched sporangiophores are long and terminate whit columella surrounded by a dark, round sporangium (Figure 2b). These are the characteristics of the genus Rhizopus [16].

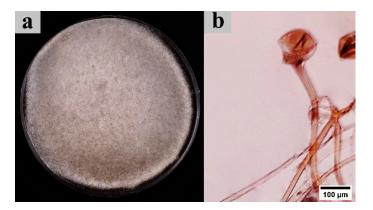


Figure 2. (a) Fungal colony grown on SDA, (b) microscopic image.

As mentioned, no fungal activity was observed in the foxing stains. Therefore, these stains should be considered abiotic foxing. One sign of foxing stains is their fluorescence under UV light [8,17,18]. The color of foxing spots of the manuscript, under reflected light, appear to be orange-brown. Similar to the report of Francisca et al. (2020) [17], under UV radiation, there is a round, well-delimited white fluorescent halo surrounding a non-fluorescent darker center. What is clear is that the halo extends beyond the visible stain (Figure 3). Cain and Miller (1984) have classified these spots as Bullseye [19]. This type of foxing always has metal cores, which do not fluoresce and appear dark [20].

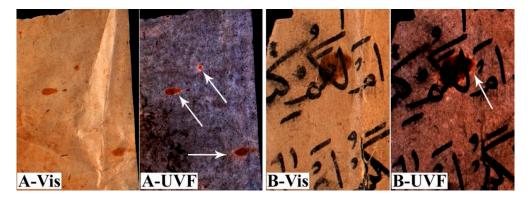


Figure 3. Photographic imaging of two foxing spot on two different papers (**A** and **B**) of manuscript; Vis: visible reflected light, UVF: Ultraviolet fluorescence.

The qualitative concentration of metal elements on the foxed spots of the paper, and adjacent unfoxed areas, were checked by μ XRF spectroscopy. The μ XRF spectra are presented in Figure 4. These spectra show a high accumulation of iron, copper and calcium in the foxing spots compared to the adjacent section. In previous reports, the presence of iron, copper and calcium was documented in foxing spots [15,20–22]. These elements in the paper are believed to result from the papermaking process [22]. Metal contamination may also come from airborne dust. One of the main known sources of these metals in foxing stains is water [14]. Therefore, due to the shape and scattering of the foxings in this manuscript, the source of these metals is likely to be dirty water droplets. Meanwhile, the presence of copper and iron has been confirmed in almost all samples with abiotic foxing spots. Metal ions, iron (II) and copper (I) ions in particular, act as a catalyst and accelerator for cellulose free radical oxidative reaction in cellulose [4,23]. Furthermore, it seems that copper ions, due to their higher catalytic effect, are more capable in foxing formation than iron ions [8,23].

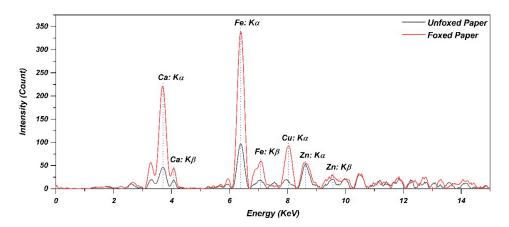


Figure 4. XRF spectra of paper with and without foxing stains that show increased levels of iron, copper and calcium in the foxed area.

The oxidation rate at foxing stains was evaluated using FTIR spectroscopy. Figure 5 presents the Gaussian peak deconvolution of the FTIR spectra in the range of 850–1800 cm⁻¹. Spectra changes under the influence of foxing show the increase in the absorbance peaks at 1700–1760 cm⁻¹. This region corresponds to stretching vibrations of C=O groups and is known as the carbonyl index in the FTIR spectrum. The formation and intensity increase of these peaks can be considered as the oxidation index of cellulose in paper [24,25].

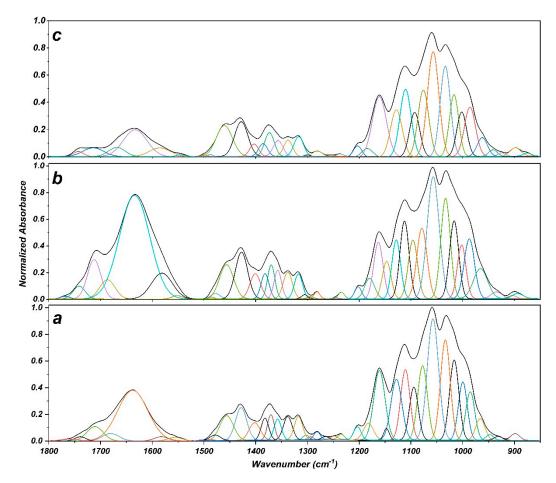


Figure 5. Gaussian peak deconvolution of FTIR spectra of foxed (**b**: S1, **c**: S2) and unfoxed (**a**) papers, baseline corrected in the region of 850–1800 cm⁻¹.

According to Lojewska et al. (2005) [26], cellulose oxidation index defined as the ratio of the intensity of absorbance bands of about 1720 to 1630 cm⁻¹. Therefore, the ratio of these peaks was calculated based on the deconvoluted peaks presented in Table 1. The results show that the intensity ratio of peaks is increased in the foxing spots (Figure 6). In other words, it indicates more oxidation of cellulose in the foxed paper compared to the unfoxed paper.

In the FTIR spectra, the absorbance bands around 1710 and 1740 cm⁻¹ correspond to C=O stretching vibrations in ketone and aldehyde groups, respectively [27]. The results show that foxing degradation in both samples has increased the C=O intensity ratio. Both S1 and S2 samples show an increase in both ketones and aldehydes. However, ketones and aldehydes have formed more in the S1 and S2 samples, respectively.

According to a previous study, cellulose oxidation is involved with C-OH and C-O-C vibrations, for secondary alcohol at C2 and C3, primary alcohol at C6, the C-O-C of glucopyranose ring and oxygen bridge [24]. In other words, the oxidation of cellulose leads to a decrease in the absorption peaks at 1035, 1160 and 1325 cm⁻¹ which corresponds to the C-O-C glucopyranose ring stretching vibration [28], stretching vibration of the glycosidic oxygen bridge (C-O-C) [29] and C-OH stretching vibration [24], respectively.

Sample	Fitting _ Parameters	Deconvoluted Peaks						
		1035	1160	1335	1635	1710	1740	1460
Unfoxed paper	Center	1033.4	1160.9	1338.1	1637.7	1710.7	1738.8	1456.1
	Height	0.754	0.523	0.175	0.381	0.108	0.032	0.187
	Area	17.58	14.14	3.72	27.43	4.29	0.85	6.07
	FWHH *	21.92	25.41	19.95	67.66	37.13	24.94	30.53
Foxed paper (S1)	Center	1033.0	1162.9	1338.1	1634	1712.2	1741.2	1456.4
	Height	0.754	0.425	0.2	0.778	0.295	0.097	0.257
	Area	17.17	9.60	4.37	56.99	9.72	2.82	8.75
	FWHH	21.39	21.24	20.51	68.80	30.95	27.43	31.94
Foxed paper (S2)	Center	1033.9	1161.2	1337.7	1635.1	1716.1	1742.8	1461
	Height	0.716	0.487	0.132	0.232	0.064	0.041	0.247
	Area	19.09	16.48	3.28	18.15	3.11	1.05	9.51
	FWHH	25.05	31.78	23.29	73.41	45.55	24.31	36.15

Table 1. The results of peak deconvolution and fitting Gaussian function of the spectra.

* Full Width at the Half-Height.

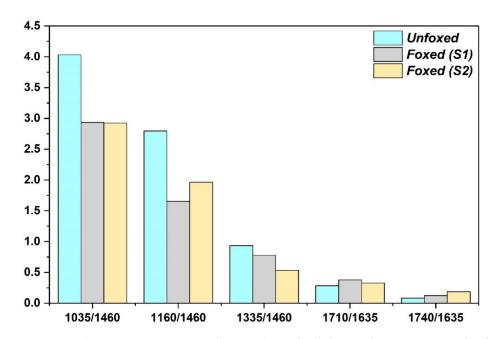


Figure 6. Peaks intensity ratios as oxidation indices of cellulose in the FTIR spectra of unfoxed and foxed papers.

The decreasing rate at 1035, 1160 and 1325 cm⁻¹ has been evaluated and reported in Figure 6 with respect to the intensity corresponding to 1460 cm⁻¹ as the internal standard peak. The internal standard peak was selected based on a previous study [24]. According to the results, foxing-induced oxidation has resulted in the breakdown of glycosidic bond, glucopyranose ring-opening and oxidation of C-OH groups. The breakdown of glycosidic bond has occurred more in the S1 than in the S2. This has led to the formation of more ketone groups in this sample. Conversely, in the S2 sample, more oxidation of the C-OH has led to the formation of more aldehydes.

As mentioned, cellulose oxidation increases carbonyl groups. Carbonyl groups are precursors of chromophores that cause cellulose fibers to become yellow [30]. Previous studies have shown that ketones are the main cause of the yellowing of cellulosic pulps [31]. In addition, carboxyls can also be formed during the oxidation processes of cellulose [30]. However, carboxyls alone do not play a significant role in discoloration, but their concomitant presence with ketones and aldehydes exacerbates yellowing [30]. Therefore, oxidation of cellulose and formation of carbonyls can be considered as one of the main reasons for the formation of yellow-brown spots in foxings (Figure 7).

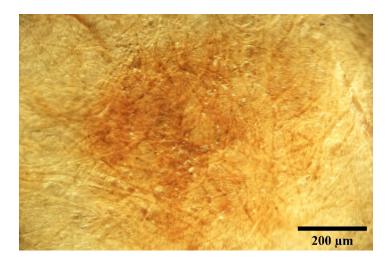


Figure 7. Discoloration of paper fibers to yellow-brown in foxing stains as a result of cellulose oxidation.

4. Conclusions

In this study, the foxing spots in a manuscript from the late 18th century were examined. Based on the results of this study, no fungal activity was observed on the foxing spots in this manuscript. The presence of a Rhizopus fungus was detected on only one page of the manuscript, and not in any of the foxing stains. Therefore, it can be concluded that these spots are abiotic foxing.

UV fluorescence examination also showed a bright halo around the foxing spots, which is known as Bullseye. It has been reported that this type of foxing is usually caused by metal ions. μ -XRF spectroscopy confirmed that iron and copper had accumulated in the foxing area. The accumulation of these metals intensifies the oxidation of cellulose by catalyzing radical reactions. In addition, cellulose oxidation in foxing stains was investigated using FTIR spectroscopy. The results show that the ratio of the intensity of bands about 1710 and 1740 cm⁻¹ to that at 1635 cm⁻¹ can increase in foxed area. In other words, ketone and aldehyde groups have increased as a result of cellulose oxidation due to foxing. Therefore, it is an appropriate indicator to study the oxidation intensity in abiotic foxing spots. The results show that the cellulose oxidation in foxing stains has been involved with the glucopyranose ring opening, oxygen bridges breaking and also hydroxyl groups. Cellulose oxidation can be considered as one of the main reasons for the color changing of paper to yellow-brown at foxing stains.

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