Selenium-Epoxy 'Click' Reaction and Se-Alkylation: Efficient Access to Organo-Selenium and Selenonium Compounds

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General Methods and Materials

Glycidol, glycidyl methacrylate, allyl glycidyl ether, glycidyl propargyl ether. epichlorohydrin, aniline, benzoic acid, phenol, thiophenol, benzeneselenol, lithium hydroxide monohydrate (LiOH), 1,8,-diazabicyclo[5.4.0]undec-7-ene (DBU), triethylamine (TEA), acetic anhydride, 3-bromopropane-1,2-diol, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI), 4-dimethylaminopyridine (DMAP), acetic acid. iodomethane, silver tetrafluoroborate (AgBF₄), red blood cells (sterile defibrinated sheep's blood, RBC), were purchased from commercial sources. NMR spectra were recorded on a Varian NMR system 500 MHz spectrometer, using CDCl₃, DMSO-d₆ and D₂O as the solvents. HRMS was carried out on a JMS-700 from JEOL in EI and FAB mode. FT-IR spectrometer with ATR (attenuated total reflection) was carried out on Agilent Cary 630 FTIR. Hemolysis assay were performed using a microplate reader (Spectra Max 340, Molecular Devices, Sunnyvale, CA USA).

Synthesis of **3** in water: To a solution of freshly obtained benzeneselenol **1** (471 mg, 3.0 mmol) and glycidol **2** (222 mg, 3.0 mmol) in 3 mL water was added TEA, DBU, or LiOH (0.01, 0.03 or 0.05 eq., respectively, per selenol unit) at 0 °C. The cooling was removed and the reaction mixture was stirred at room temperature for time shown in Table 2. After this time, 100 μ L of the reaction mixture was removed and diluted with 1 mL of CDCl₃, dried over sodium sulfate, and studied with the help of NMR spectroscopies.

Synthesis of **3** in chloroform: To a solution of freshly obtained benzeneselenol **1** (471 mg, 3.0 mmol) and glycidol **2** (222 mg, 3.0 mmol) in 3 mL chloroform was added TEA, DBU, or LiOH (0.01, 0.03 or 0.05 eq., respectively, per selenol unit) at 0 °C. The cooling was removed and the reaction mixture was stirred at room temperature for time shown in Table 1. After this time, 100 μ L of the reaction mixture was removed and diluted with 1 mL of CDCl₃, and studied with the help of NMR spectroscopies.

Synthesis of **3** via **6**: To a solution of 3-bromopropane-1,2-diol **6** (1.0 g, 6.45 mmol) and benzeneselenol **1** (2.03 g, 12.9 mmol) in 6 mL MeCN was added TEA (5.22 g, 51.6 mmol) and the reaction mixture was stirred at 75 °C for 6 h. After this time, the reaction mixture was

cooled to room temperature, diluted with DCM and washed with water. The organic solvent was removed under reduced pressure. The crude product was purified by silica gel column chromatography (eluent = DCM/MeOH 95:5) which gave about 1.31 g of **3** as a white powder (yield = 88%). ¹H NMR (500 MHz, DMSO-*d*₆) δ 7.50 – 7.44 (m, 2H), 7.35 – 7.15 (m, 3H), 4.98 (d, *J* = 5.1 Hz, 1H), 4.65 (t, *J* = 5.5 Hz, 1H), 3.74 – 3.56 (m, 1H), 3.44 – 3.33 (m, 2H), 3.17 – 2.85 (m, 2H). ¹³C NMR (126 MHz, DMSO-*D*₆) δ 131.22, 130.94, 129.15, 126.19, 70.88, 65.01, 31.59. ⁷⁷Se NMR (95 MHz, DMSO-*D*₆) δ 262.59. HRMS (EI+) m/z calcd for C₉H₁₂O₂Se [M]⁺: 232.0003, observed: 231.9997. IR (cm⁻¹): 3324, 3194, 3054, 2944, 2925, 1576, 1487, 1474, 1448, 1435, 1412, 1375, 1321, 1295, 1244, 1200, 1181, 1164, 1134, 1095, 1017, 1041, 1026, 1017, 984, 896, 872, 836, 803, 724, 687, 670.

Synthesis of 7: A suspension of compound **3** (500 mg, 2.16 mmol), acetic acid (649 mg, 10.8 mmol), EDCI (2.07 g, 10.8 mmol), and DMAP (132 mg, 1.08 mmol) was stirred at 0 °C in 10 mL DMF for 1 hr. Then the reaction mixture was stirred at room temperature for overnight. After this time, the reaction was diluted with DCM and washed with water, 0.1 M HCl solution, followed by saturated NaHCO₃. The organic solvent was removed under reduced pressure which gave about 552 mg of the product as a colorless oil (yield = 81%). ¹H NMR (500 MHz, DMSO-*d*₆) δ 7.61 – 7.42 (m, 2H), 7.40 – 7.19 (m, 3H), 5.10 (dtd, *J* = 7.0, 6.1, 3.3 Hz, 1H), 4.30 – 4.05 (m, 2H), 3.24 – 3.12 (m, 2H), 1.98 (s, 3H), 1.90 (s, 3H). ¹³C NMR (126 MHz, DMSO-*D*₆) δ 170.22, 169.82, 131.94, 129.57, 129.42, 127.13, 70.76, 64.06, 26.68, 20.69, 20.61. ⁷⁷Se NMR (95 MHz, DMSO-*D*₆) δ 272.87. HRMS (EI+) m/z calcd for C₁₃H₁₆O₄Se [M]⁺: 316.0214, observed: 316.0211. IR (cm⁻¹): 3054, 2950, 1735, 1578, 1477, 1436, 1367, 1220, 1108, 1041, 1021, 998, 961, 943, 872, 836, 803, 736, 691, 669.

Synthesis of **12-15**: To a solution of **8**, **9**, **10**, or **11** (3.0 mmol) and freshly obtained benzeneselenol **1** (471 mg, 3.0 mmol) in 3 mL water was added LiOH (0.03 eq. per selenol unit) at 0 °C. The cooling was removed and the reaction mixture was stirred at room temperature for 15 mins. After this time, 100 μ L of the reaction mixture was added to 1 mL CDCl₃, dried over sodium sulfate, and studied with the help of NMR spectroscopies.

12: ¹H NMR (500 MHz, CDCl₃) δ 7.56 – 7.47 (m, 2H), 7.29 – 7.20 (m, 3H), 4.12 (dd, J = 2.4, 0.9 Hz, 2H), 3.96 – 3.87 (m, 1H), 3.67 – 3.52 (m, 2H), 3.12 – 2.97 (m, 2H), 2.73 (d, J = 4.4 Hz, 1H), 2.43 (t, J = 2.4 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 132.97, 129.56, 129.32, 127.35, 79.43, 75.06, 72.71, 69.37, 58.65, 31.93. ⁷⁷Se NMR (95 MHz, CDCl₃) δ 244.31. HRMS (EI+) m/z calcd for C₁₂H₁₄O₂Se [M]⁺: 270.0159, observed: 270.0156. IR (cm⁻¹): 3408,

3283, 3054, 2901, 2855, 2115, 1578, 1477, 1436, 1356, 1263, 1205, 1095, 1071, 1021, 946, 909, 734, 689, 699.

13: ¹H NMR (500 MHz, CDCl₃) δ 7.55 – 7.47 (m, 2H), 7.29 – 7.19 (m, 3H), 5.85 (ddt, J = 17.3, 10.4, 5.7 Hz, 1H), 5.28 – 5.14 (m, 2H), 3.95 (dt, J = 5.7, 1.5 Hz, 2H), 3.94 – 3.86 (m, 1H), 3.55 – 3.43 (m, 2H), 3.12 – 2.97 (m, 2H), 2.75 (d, J = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 134.43, 132.90, 129.68, 129.27, 127.28, 117.48, 72.90, 72.37, 69.52, 31.97. ⁷⁷Se NMR (95 MHz, CDCl₃) δ 245.11. HRMS (EI+) m/z calcd for C₁₂H₁₆O₂Se [M]⁺: 272.0316, observed: 272.0313. IR (cm⁻¹): 3412, 3067, 2896, 2855, 1647, 1578, 1477, 1436, 1420, 1345, 1265, 1203, 1099, 1071, 1021, 997, 924, 799, 732, 689, 699.

14: ¹H NMR (500 MHz, CDCl₃) δ 7.56 – 7.47 (m, 2H), 7.25 (m, 3H), 6.09 (p, *J* = 1.1 Hz, 1H), 5.57 (p, *J* = 1.6 Hz, 1H), 4.28 – 4.16 (m, 2H), 4.02 – 3.94 (m, 1H), 3.15 – 2.96 (m, 2H), 2.88 – 2.78 (s, 1H), 1.91 (t, *J* = 1.3 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 167.40, 135.95, 133.19, 129.42, 127.63, 126.29, 68.74, 67.29, 32.44, 18.40. ⁷⁷Se NMR (95 MHz, CDCl₃) δ 243.31. HRMS (EI+) m/z calcd for C₁₃H₁₆O₃Se [M]⁺: 300.0265, observed: 300.0262. IR (cm⁻¹): 3431, 3056, 2980, 2950, 2925, 2892, 1701, 1636, 1578, 1477, 1451, 1436, 1420, 1401, 1377, 1317, 1293, 1157, 1101, 1071, 1021, 943, 812, 734, 689, 699.

15: ¹H NMR (500 MHz, CDCl₃) δ 7.58 – 7.48 (m, 2H), 7.27 (m, 3H), 3.97 – 3.88 (m, 1H), 3.70 – 3.60 (m, 2H), 3.16 – 3.02 (m, 2H), 2.76 (d, *J* = 5.1 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 133.18, 129.46, 128.95, 127.69, 70.19, 48.44, 32.44. ⁷⁷Se NMR (95 MHz, CDCl₃) δ 247.43. HRMS (EI+) m/z calcd for C₉H₁₁ClOSe [M]⁺: 249.9664, observed: 249.9661. IR (cm⁻¹): 3390, 3052, 2948, 1654, 1576, 1476, 1436, 1339, 1297, 1261, 1209, 1181, 1157, 1131, 1066, 1034, 1021, 998, 982, 943, 909, 883, 851, 808, 732, 689, 699.

Reaction with aniline, phenol, benzoic acid, or thiophenol: To a solution of glycidol **2** (3.0 mmol), freshly obtained benzeneselenol **1** (3.3 mmol) and any one of the nucleophile (3.3 mmol) in 3 mL water added LiOH (0.03 eq. per selenol units) at 0 °C. The cooling was removed and the reaction mixture was stirred at room temperature for 15 mins. After this time, 100 μ L of the reaction mixture was added in 1 mL CDCl₃, dried over sodium sulfate, and studied with the help of NMR spectroscopy.

Reaction between thiophenol and glycidol in the absence of 1: To a solution of glycidol (222 mg, 3.0 mmol), thiophenol (330 mg, 3 mmol) in 3 mL water added LiOH (2 mg, 0.09 mmol) at 0 °C. The cooling was removed and the reaction mixture was stirred at room temperature for 15 mins. After this time, 100 μ L of the reaction mixture was added in 1 mL CDCl₃, dried over sodium sulfate, and studied with the help of NMR spectroscopy.

Synthesis of 16/17: To a solution of selenide 3 or 7 (100 mg/mL) and AgBF₄ (1.25 eq. per selenide) in MeCN was added iodomethane (4 eq. per selenol unit) and the reaction mixture was stirred under an inert atmosphere in the dark at 50 °C for 24 hr. After this time, AgI was removed by syringe filter and the filtrate was precipitated into diethyl ether thrice and then dried under high vacuum conditions.

16: ¹H NMR (500 MHz, DMSO-*D*₆) δ 7.97 – 7.90 (m, 2H), 7.74 – 7.59 (m, 3H), 5.81 – 5.74 (m, 1H), 5.15 (t, *J* = 5.5 Hz, 0.5H), 5.09 (t, *J* = 5.6 Hz, 0.5H), 3.95 (ddq, *J* = 7.8, 5.9, 4.7 Hz, 0.5H), 3.77 – 3.56 (m, 2.5H), 3.52 – 3.37 (m, 2H), 3.07 (d, *J* = 1.6 Hz, 3H). ¹³C NMR (126 MHz, DMSO-*D*₆) δ 132.62, 132.38, 130.97, 130.72, 130.52, 130.41, 127.72, 126.55, 67.29, 66.95, 64.89, 48.89, 48.60, 23.34, 22.99. ⁷⁷Se NMR (95 MHz, DMSO-*D*₆) δ 377.97. HRMS (FAB+) m/z calcd for C₁₀H₁₅O₂Se⁺ [M]⁺: 247.0237, observed: 247.0235. IR (cm⁻¹): 3507, 3054, 2952, 2884, 1636, 1578, 1582, 1483, 1446, 1416, 1285, 995, 937, 877, 743, 683.

17: ¹H NMR (500 MHz, DMSO- D_6) δ 7.98 (m, 2H), 7.74 – 7.63 (m, 3H), 5.38 – 5.30 (m, 0.5H), 5.06 (ddt, J = 7.4, 5.0, 3.6 Hz, 0.5H), 4.28 – 4.10 (m, 2H), 1 Hz, 2H), 3.92 (dd, J = 12.7, 3.6 Hz, 1H), 3.87 – 3.78 (m, 2H), 3.11 (d, J = 11.9 Hz, 3H), 2.03 (d, J = 8.1 Hz, 3H), 1.82 (d, J = 6.0 Hz, 3H). ¹³C NMR (126 MHz, DMSO- D_6) δ 170.17, 170.15, 170.12, 169.93, 132.89, 132.77, 131.04, 130.83, 130.58, 130.49, 127.04, 126.21, 67.83, 67.33, 63.77, 63.69, 43.61, 43.53, 24.26, 23.83, 20.57, 20.52, 20.37, 20.25. ⁷⁷Se NMR (95 MHz, DMSO- D_6) δ 380.59, 379.13. HRMS (FAB+) m/z calcd for C₁₄H₁₉O₄Se⁺ [M]⁺: 331.0449, observed: 331.0449. (cm⁻¹): 3507, 3050, 2948, 1735, 1645, 1513, 1483, 1446, 1420, 1373, 1231, 1017, 939, 877, 840, 742, 685.

Hemolysis assay

Hemolytic activity of the crosslinked micelles was determined using sheep's red blood cells (RBCs). RBCs were pelletized by centrifuging 1 mL of the blood and then washing the RBCs at 4 times with PBS buffer (pH 7.4) solution. The crosslinked micelles were re-dispersed in PBS buffer. Then, 1mL of samples and 25 μ L of the RBCs were mixed and incubated at room temperature for 2h. After incubation, the mixtures were re-pelletized by centrifuging and 200 μ L of the supernatant of each sample were put into wells in a 96-well plate. The absorbance at 540 nm was measured with a microplate reader. The percent hemolysis was calculated by following equation.

 $I = \{(A_i - A_{negative}) / (A_{positive} - A_{negative})\} \times 100 \%$

(A_i = the absorbance of samples solution, $A_{negative}$ = the absorbance of PBS solution, $A_{positive}$ = the absorbance of DI.)



Figure S1. Crude ¹H-NMR (CDCl₃) from ring opening reaction in chloroform under TEA catalysis.



Figure S2. Crude ¹H-NMR (CDCl₃) from ring opening reaction in chloroform under DBU catalysis.



Figure S3. Crude ¹H-NMR (CDCl₃) from ring opening reaction in chloroform under LiOH catalysis.



Figure S4. Crude ¹H-NMR (CDCl₃) from ring opening reaction in water under TEA catalysis.



Figure S5. Crude ¹H-NMR (CDCl₃) from ring opening reaction in water under DBU catalysis.



Figure S6. Crude ¹H-NMR (CDCl₃) from ring opening reaction in water under LiOH catalysis.



Figure S7. Crude ¹H-NMR (CDCl₃) from non-stoichiometric ring opening reaction in chloroform.



Figure S8. Crude ¹H-NMR (CDCl₃) from non-stoichiometric ring opening reaction in water.



200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 Figure S9. 13 C-NMR (DMSO- d_6) of **3**.









Figure S12. Crude ¹³C-NMR (CDCl₃) of **12**.



Figure S13. Crude ¹H-NMR (CDCl₃) of **13**.



Figure S14. Crude 13 C-NMR (CDCl₃) of **13**. The signal from methanol is shown with an asterisk.



Figure S15. Crude ¹H-NMR (CDCl₃) of 14.





Figure S17. Crude ¹H-NMR (CDCl₃) of **15**.



Figure S19. ¹H-NMR (CDCl₃) of phenol (top) and the reaction mixture (bottom). The signals from phenol remains unchanged.



Figure S20. ¹H-NMR (CDCl₃) of benzoic acid (top) and the reaction mixture (bottom). The signals from benzoic acid remains unchanged.



Figure S21. ¹H-NMR (CDCl₃) of aniline (top) and the reaction mixture (bottom). The signals from aniline remains unchanged.



Figure S22. ¹H-NMR (CDCl₃) of reactants, products, and the crude reaction mixtures in thiol/selenium-based reactions.



Figure S23. ¹H-NMR (DMSO-*d*₆) of **16**. The signals from DMF is shown with an asterisk.



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Figure S30. ⁷⁷Se-NMR (CDCl₃) of 15.