

Microwave Assisted Esterification of Aryl/Alkyl Acids Catalyzed by *N*-Fluorobenzenesulfonimide

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Table S1. The central composite design and experimental results.

	X₁	X₂	X₃	Y
Space Type	Temperature (°C)	Time (min)	Amount of Catalyst (%)	Conversion
Center	120	15	7	71
Center	120	15	7	72
Center	120	15	7	73
Center	120	15	7	72
Center	120	15	7	71
Axial	120	30	7	100
Axial	70	15	7	1
Axial	120	15	10	81
Axial	120	15	3	38
Axial	120	0	7	0
Axial	150	15	7	100
Factorial	150	30	4	100
Factorial	90	0	4	0
Factorial	90	30	10	11
Factorial	150	0	10	0
Center	120	15	7	71

Table S2. Coefficients for quadratic equation.

	Intercept	X₁	X₂	X₃	X₁ X₂	X₁ X₃	X₂ X₃	X₁²	X₂²	X₃²
Conversion	73.4907	31.7281	50	14.8122	37.0622	22.25	9.4781	-7.5670	-26.6217	-9.9866
p-values		< 0.0001	< 0.0001	0.0014	< 0.0001	0.0009	0.0238	0.0046	< 0.0001	0.0026

Design-Expert® Software
Factor Coding: Actual

Conversion (%)
● Design Points
0 100

X1 = A: Temperature
X2 = B: Time

Actual Factor
C: Amount of Catalyst = 7

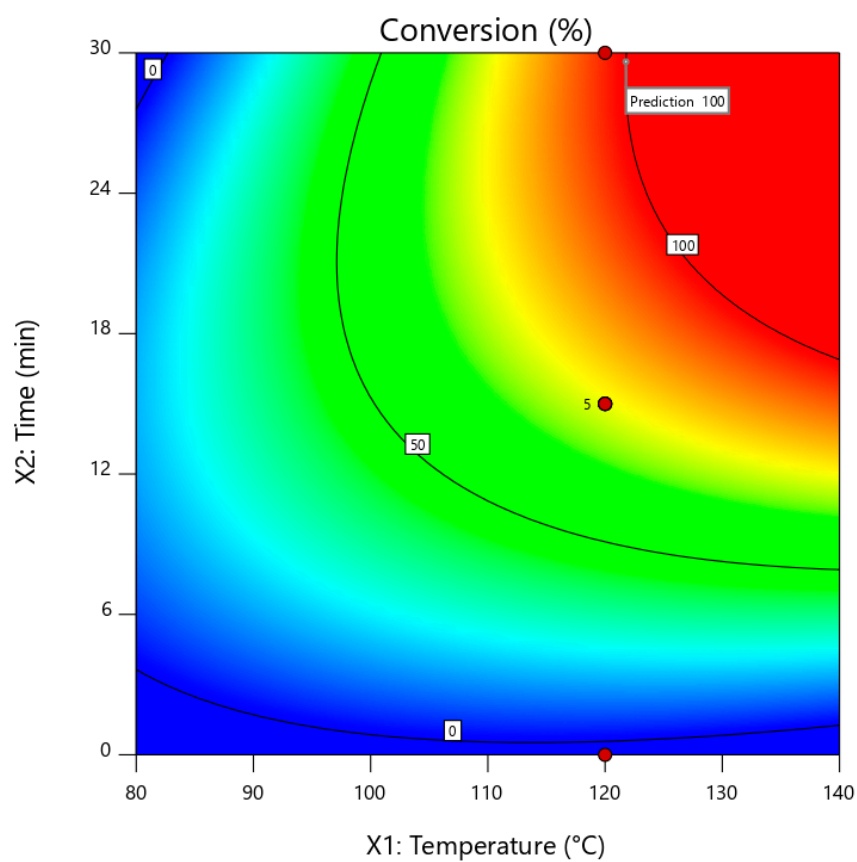


Figure S1. 2D plot obtained after RSM analysis and numerical optimization.

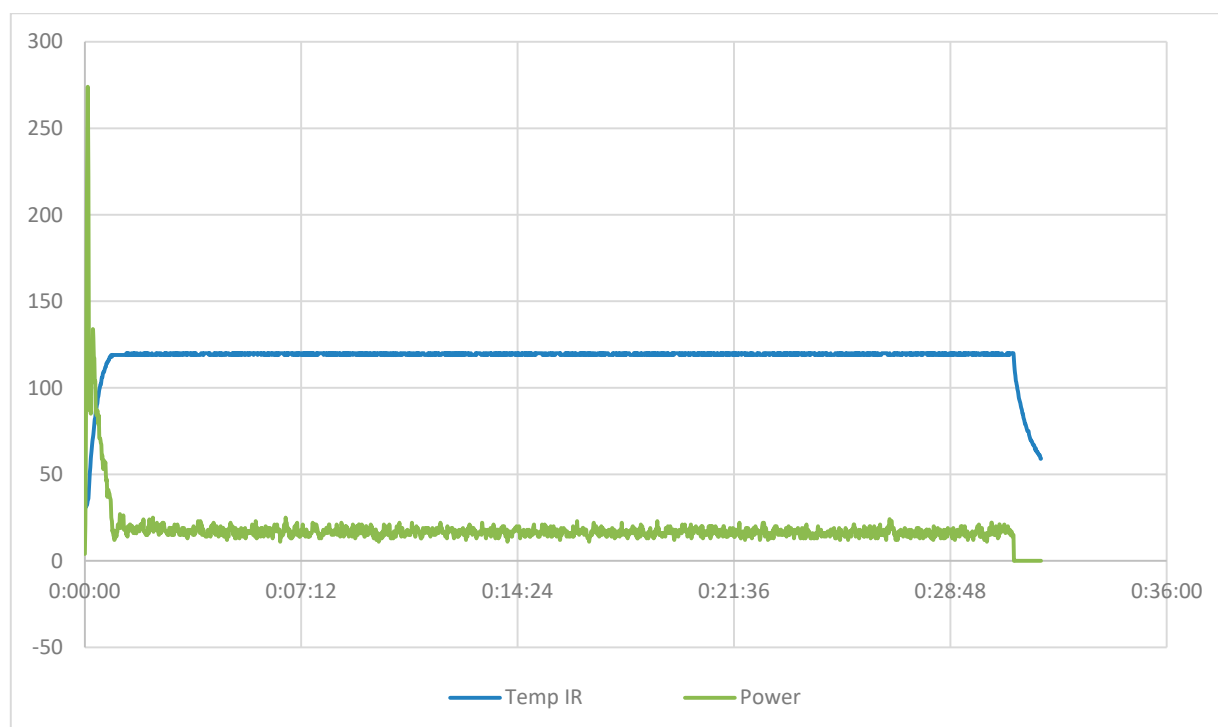


Figure S2. Energy input (gray line – power [W]) and temperature (blue line – in °C) for esterification under optimal reaction conditions and MW irradiation.

List S1. NMR spectral data of synthesized esters.

Methyl benzoate - 1a [1]

^1H NMR (400 MHz, CDCl_3) δ 8.05 – 8.03 (m, 2H), 7.55 (t, J = 7.4 Hz, 1H), 7.43 (t, J = 7.7 Hz, 2H), 3.91 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 167.1, 132.9, 130.2, 129.6, 128.3, 52.1.

Ethyl benzoate – 1b [1]

^1H NMR (400 MHz, CDCl_3) δ 8.06 – 8.04 (m, 2H), 7.55 (d, J = 7.4 Hz, 1H), 7.44 (t, J = 7.6 Hz, 2H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 7.1 Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.7, 132.8, 130.6, 129.6, 128.3, 60.9, 14.3.

Butyl benzoate – 1c [2]

^1H NMR (400 MHz, CDCl_3) δ 8.05 – 8.03 (m, 2H), 7.54 (t, J = 7.4 Hz, 1H), 7.43 (t, J = 7.6 Hz, 2H), 4.32 (t, J = 6.6 Hz, 2H), 1.81 – 1.73 (m, 2H), 1.47 (dd, J = 15.0, 7.5 Hz, 2H), 0.98 (t, J = 7.4 Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.7, 132.8, 130.6, 129.5, 128.3, 64.8, 30.8, 19.3, 13.8.

Methyl stearate – 2a [3]

^1H NMR (400 MHz, CDCl_3) δ 3.66 (s, 3H), 2.30 (t, J = 7.6 Hz, 2H), 1.62 (dd, J = 14.3, 7.1 Hz, 2H), 1.28-1.26 (m, 28H), 0.87 (t, J = 6.8 Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 174.3, 51.4, 34.1, 31.9, 29.7, 29.7, 29.7, 29.7, 29.6, 29.5, 29.4, 29.3, 29.2, 25.0, 22.7, 14.1.

Ethyl stearate – 2b [4]

^1H NMR (400 MHz, CDCl_3) δ 4.12 (q, J = 7.1 Hz, 2H), 2.28 (t, J = 7.6 Hz, 2H), 1.63 – 1.60 (m, 2H), 1.29 – 1.26 (m, 30H), 0.87 (t, J = 6.8 Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 173.9, 60.2, 34.4, 32.0, 29.8, 29.7, 29.7, 29.5, 29.4, 29.3, 29.2, 25.0, 22.7, 14.3, 14.1.

Butyl stearate – 2c [5]

^1H NMR (400 MHz, CDCl_3) δ 4.07 (t, J = 6.7 Hz, 2H), 2.29 (t, J = 7.5 Hz, 2H), 1.64 – 1.59 (m, 4H), 1.44 – 1.34 (m, 2H), 1.28-1.26 (m, 28H), 0.91 (dt, J = 13.6, 7.2 Hz, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 174.0, 64.1, 34.4, 31.9, 30.7, 29.7, 29.7, 29.6, 29.5, 29.4, 29.3, 29.2, 25.1, 22.7, 19.2, 14.1, 13.7.

Isopropyl stearate – 2d [6]

^1H NMR (400 MHz, CDCl_3) δ 5.00 (dt, J = 12.5, 6.2 Hz, 1H), 2.25 (t, J = 7.5 Hz, 2H), 1.62 – 1.59 (m, 2H), 1.26 – 1.23 (m, 34H), 0.88 (t, J = 6.7 Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 173.4, 67.3, 34.7, 31.9, 29.8, 29.7, 29.7, 29.6, 29.5, 29.4, 29.3, 29.2, 25.1, 22.7, 21.9, 14.1.

Methyl 4-hidroxybenzoate – 3a [7]

^1H NMR (400 MHz, CDCl_3) δ 7.95 (d, J = 8.8 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 6.25 (s, 1H), 3.89 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 167.2, 160.1, 131.9, 122.5, 115.3, 52.0.

Methyl 3-hidroxybenzoate – 4a [8]

^1H NMR (400 MHz, CDCl_3) δ 7.61 (dd, J = 7.0, 5.1 Hz, 2H), 7.31 (t, J = 7.9 Hz, 1H), 7.10 – 7.07 (m, 1H), 6.35 (s, 1H), 3.92 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 167.5, 156.0, 131.2, 129.8, 121.8, 120.4, 116.4, 52.4.

Methyl 3-methoxybenzoate – 5a [9]

^1H NMR (400 MHz, CDCl_3) δ 7.63 (d, J = 7.7 Hz, 1H), 7.56 – 7.55 (m, 1H), 7.33 (t, J = 8.0 Hz, 1H), 7.11 – 7.08 (m, 1H), 3.91 (s, 3H), 3.84 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.9, 159.6, 131.5, 129.4, 122.0, 119.5, 114.0, 55.4, 52.1.

Methyl 4-methylbenzoate – 6a [10]

^1H NMR (400 MHz, CDCl_3) δ 7.93 (d, J = 8.2 Hz, 2H), 7.23 (d, J = 8.0 Hz, 2H), 3.89 (s, 3H), 2.40 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 167.2, 143.5, 129.6, 129.1, 127.5, 51.9, 21.6.

Methyl 4-chlorobenzoate – 7a [11]

^1H NMR (400 MHz, CDCl_3) δ 7.96 (d, J = 8.7 Hz, 2H), 7.40 (d, J = 8.7 Hz, 2H), 3.90 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.2, 139.4, 131.0, 128.7, 52.2.

Methyl 4-formylbenzoate – 8a [12]

^1H NMR (400 MHz, CDCl_3) δ 10.10 (s, 1H), 8.19 (d, J = 8.2 Hz, 2H), 7.95 (d, J = 8.5 Hz, 2H), 3.96 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 191.7, 166.1, 139.2, 130.2, 129.5, 52.6.

Methyl 4-nitrobenzoate – 9a [1]

^1H NMR (400 MHz, CDCl_3) δ 8.29 (d, J = 8.9 Hz, 2H), 8.22 (d, J = 8.9 Hz, 2H), 3.98 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.2, 150.6, 130.7, 123.6, 52.8.

Methyl 2-nitrobenzoate – 10a [13]

^1H NMR (400 MHz, CDCl_3) δ 7.91 (dd, J = 7.8, 1.1 Hz, 1H), 7.76 – 7.74 (m, 1H), 7.70 – 7.62 (m, 2H), 3.93 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.8, 132.9, 131.8, 129.9, 127.6, 123.9, 53.2.

Dimethyl phthalate – 11a [14]

^1H NMR (400 MHz, CDCl_3) δ 7.73 (dd, J = 5.7, 3.3 Hz, 2H), 7.54 (dd, J = 5.7, 3.3 Hz, 2H), 3.91 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 168.1, 131.9, 131.1, 128.9, 52.7.

Methyl oleate – 12a [3]

¹H NMR (400 MHz, CDCl₃) δ 5.35 (dt, *J* = 8.5, 5.9 Hz, 2H), 3.66 (s, 3H), 2.30 (t, *J* = 7.5 Hz, 2H), 2.06 – 1.99 (m, 4H), 1.64 – 1.60 (m, 2H), 1.31 – 1.26 (m, 20H), 0.88 (dd, *J* = 8.7, 4.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 130.0, 129.8, 51.4, 34.1, 31.9, 29.8, 29.7, 29.7, 29.6, 29.4, 29.3, 29.2, 29.2, 29.1, 27.3, 27.2, 25.0, 22.7, 14.1.

Methyl 4-methoxyphenylacetate – 13a [15]

¹H NMR (400 MHz, CDCl₃) δ 7.18 (d, *J* = 7.7 Hz, 2H), 6.84 (d, *J* = 7.3 Hz, 2H), 3.75 (s, 3H), 3.65 (s, 3H), 3.54 (s, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 172.3, 158.7, 130.3, 126.1, 114.0, 55.2, 51.9, 40.2.

Methyl 4-chlorophenylacetate – 14a [16]

¹H NMR (400 MHz, CDCl₃) δ 7.29 – 7.27 (m, 2H), 7.20 (d, *J* = 8.3 Hz, 2H), 3.68 (s, 3H), 3.58 (s, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 171.6, 133.1, 132.5, 130.7, 128.7, 52.1, 40.4.

Methyl 4-nitrophenylacetate – 15a [17]

¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.7 Hz, 2H), 7.47 (d, *J* = 8.6 Hz, 2H), 3.76 (s, 2H), 3.72 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 170.6, 147.2, 141.4, 130.4, 123.7, 52.4, 40.7.

2-(4-Isobutyl-phenyl)-propionic acid methyl ester (Ibuprofen methyl ester) – 16a [18]

¹H NMR (400 MHz, CDCl₃) δ 7.19 (d, *J* = 8.0 Hz, 2H), 7.08 (d, *J* = 8.0 Hz, 2H), 3.68 (q, *J* = 7.2 Hz, 1H), 3.62 (s, 3H), 2.44 (d, *J* = 7.2 Hz, 2H), 1.84 (dt, *J* = 13.5, 6.8 Hz, 1H), 1.47 (d, *J* = 7.2 Hz, 3H), 0.89 (d, *J* = 6.6 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 175.1, 140.5, 137.8, 129.3, 127.1, 51.9, 45.1, 45.0, 30.2, 22.4, 18.6.

Methyl cinnamate – 17a [19]

¹H NMR (400 MHz, CDCl₃) δ 7.69 (d, *J* = 16.0 Hz, 1H), 7.51 – 7.49 (m, 2H), 7.36 – 7.35 (m, 3H), 6.43 (d, *J* = 16.0 Hz, 1H), 3.78 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 167.4, 144.8, 134.4, 130.3, 128.9, 128.1, 117.8, 51.6.

Methyl 1H-indol-2-carboxylate – 18a [20]

¹H NMR (400 MHz, CDCl₃) δ 9.09 (s, 1H), 7.69 (d, *J* = 8.1 Hz, 1H), 7.44 – 7.42 (m, 1H), 7.34 – 7.30 (m, 1H), 7.23 (d, *J* = 1.1 Hz, 1H), 7.17 – 7.13 (m, 1H), 3.95 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 162.5, 136.9, 127.5, 127.1, 125.4, 122.6, 120.8, 111.9, 108.8, 52.0.

Methyl cyclohexylcarboxylate – 19a [21]

¹H NMR (400 MHz, CDCl₃) δ 3.66 (s, 3H), 2.30 (ddd, *J* = 14.9, 7.5, 3.6 Hz, 1H), 1.89 (d, *J* = 13.3 Hz, 2H), 1.75 (dd, *J* = 9.3, 6.1 Hz, 2H), 1.64 (dd, *J* = 9.7, 4.0 Hz, 1H), 1.48 – 1.39 (m, 2H), 1.26 – 1.20 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 176.6, 51.4, 43.2, 29.1, 25.8, 25.5.

Dimethyl succinate – 21a [18]

^1H NMR (400 MHz, CDCl_3) δ 3.70 (d, J = 1.4 Hz, 6H), 2.64 (d, J = 1.3 Hz, 4H).

^{13}C NMR (101 MHz, CDCl_3) δ 172.8, 51.9, 28.9.

Dimethyl phenylsuccinate – 22a [22]

^1H NMR (400 MHz, CDCl_3) δ 7.29 (dd, J = 15.5, 6.4 Hz, 5H), 4.09 (dd, J = 10.0, 5.2 Hz, 1H), 3.67 (s, 6H), 3.21 (dd, J = 16.9, 10.1 Hz, 1H), 2.67 (dd, J = 17.0, 5.2 Hz, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 173.4, 171.9, 137.7, 128.9, 127.7, 52.3, 51.8, 47.1, 37.6.

Dimethyl itaconate – 23a [23]

^1H NMR (400 MHz, CDCl_3) δ 6.34 (s, 1H), 5.73 (d, J = 0.9 Hz, 1H), 3.77 (s, 3H), 3.70 (s, 3H), 3.35 (s, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.2, 166.7, 133.7, 128.6, 52.2, 52.1, 37.5.

Dimethyl malate – 24a [24]

^1H NMR (400 MHz, CDCl_3) δ 4.54 – 4.51 (m, 1H), 3.81 (s, 3H), 3.72 (s, 3H), 3.06 (s, 1H), 2.84 (qd, J = 16.4, 5.2 Hz, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 173.7, 171.0, 67.3, 52.9, 52.1, 38.5.

Dimethyl tartarate – 25a [25]

^1H NMR (400 MHz, CDCl_3) δ 4.57 (s, 2H), 3.86 (s, 6H), 2.79 (s, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 172.0, 72.1, 53.1.

Dimethyl fumarate – 26a [18]

^1H NMR (400 MHz, CDCl_3) δ 6.87 (s, 2H), 3.82 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.4, 133.4, 52.3.

Dimethyl maleate – 27a [18]

^1H NMR (400 MHz, CDCl_3) δ 6.28 – 6.27 (m, 2H), 3.80 (dd, J = 3.5, 1.1 Hz, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.7, 129.8, 52.2.

Trimethyl citrate – 28a [26]

^1H NMR (400 MHz, CDCl_3) δ 4.15 (s, 1H), 3.83 (s, 3H), 3.69 (s, 6H), 2.86 (dd, J = 40.4, 15.6 Hz, 4H).

^{13}C NMR (101 MHz, CDCl_3) δ 173.8, 170.3, 73.3, 53.2, 52.0, 43.1.

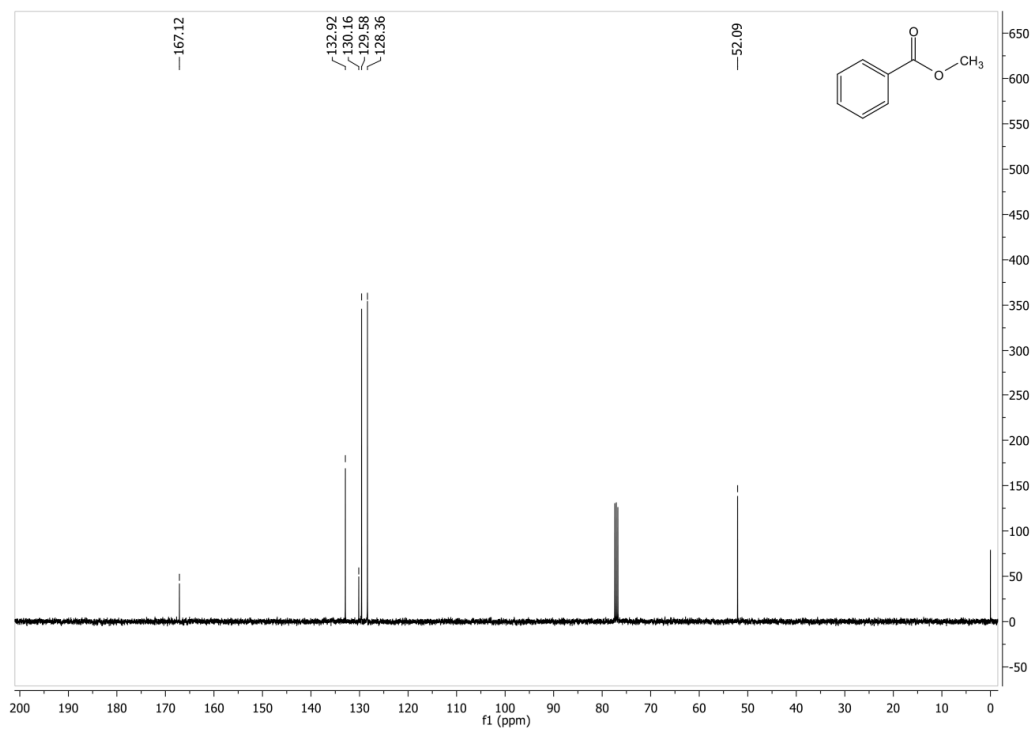
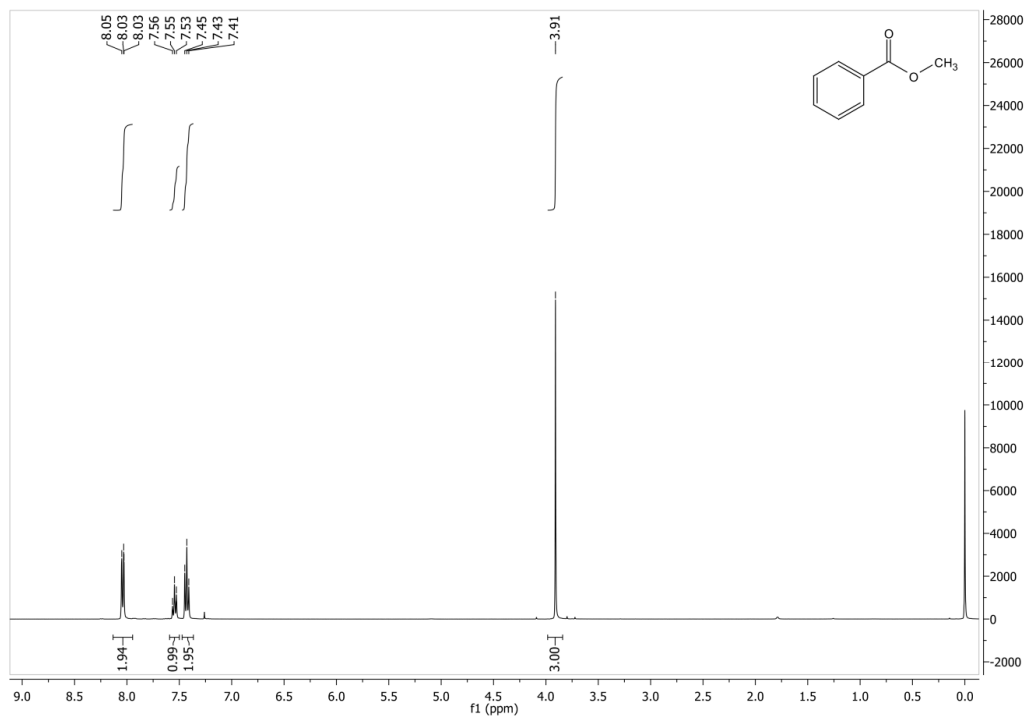
Methyl palmitate – 30a [3]

^1H NMR (400 MHz, CDCl_3) δ 3.66 (s, 3H), 2.30 (t, J = 7.5 Hz, 2H), 1.63 – 1.60 (m, 2H), 1.26 (s, 24H), 0.88 (t, J = 6.3 Hz, 3H).

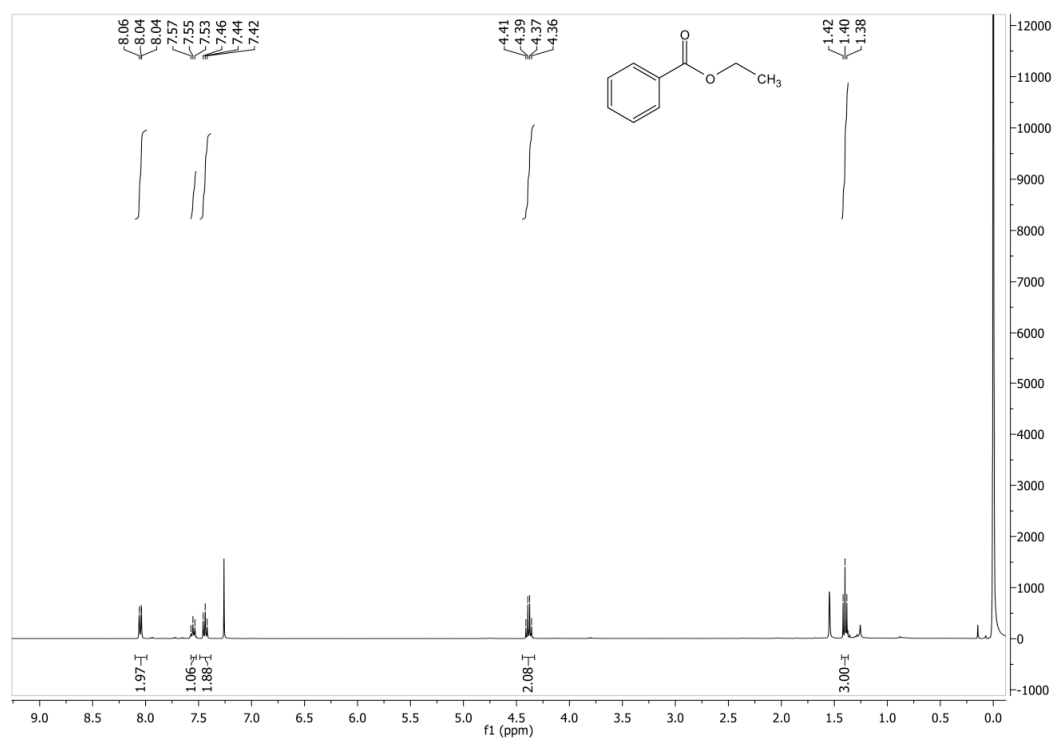
^{13}C NMR (101 MHz, CDCl_3) δ 174.3, 51.4, 34.1, 31.9, 29.7, 29.6, 29.5, 29.4, 29.3, 29.2, 25.0, 22.7, 14.1.

List S2. ^1H and ^{13}C NMR spectra of synthesized esters.

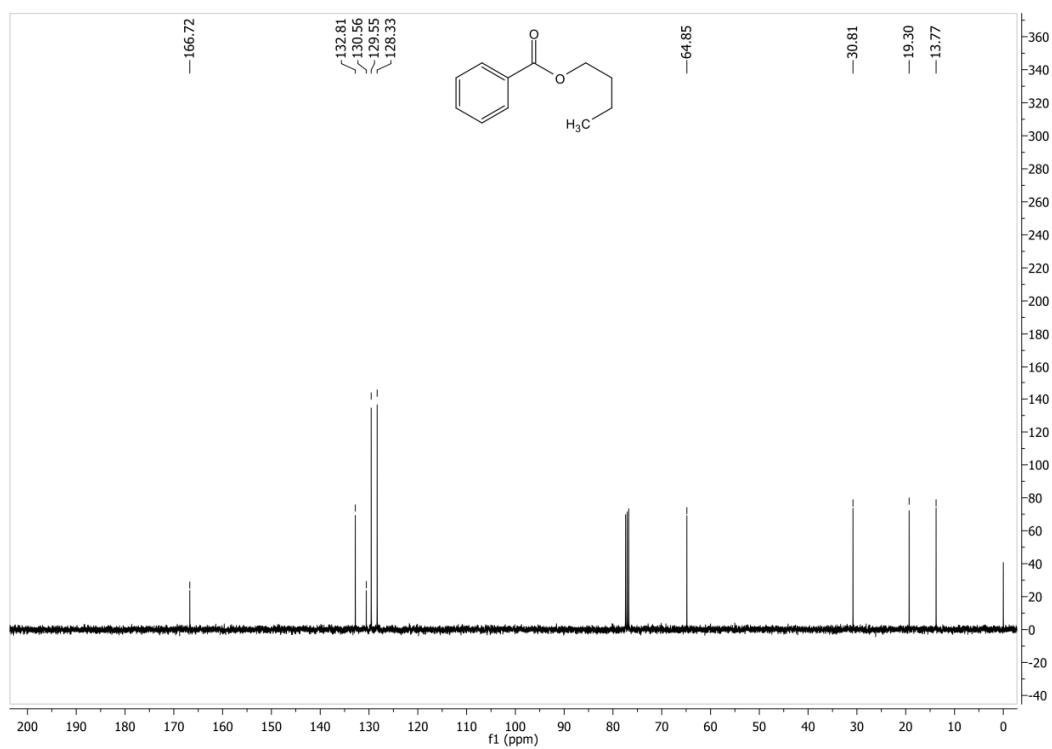
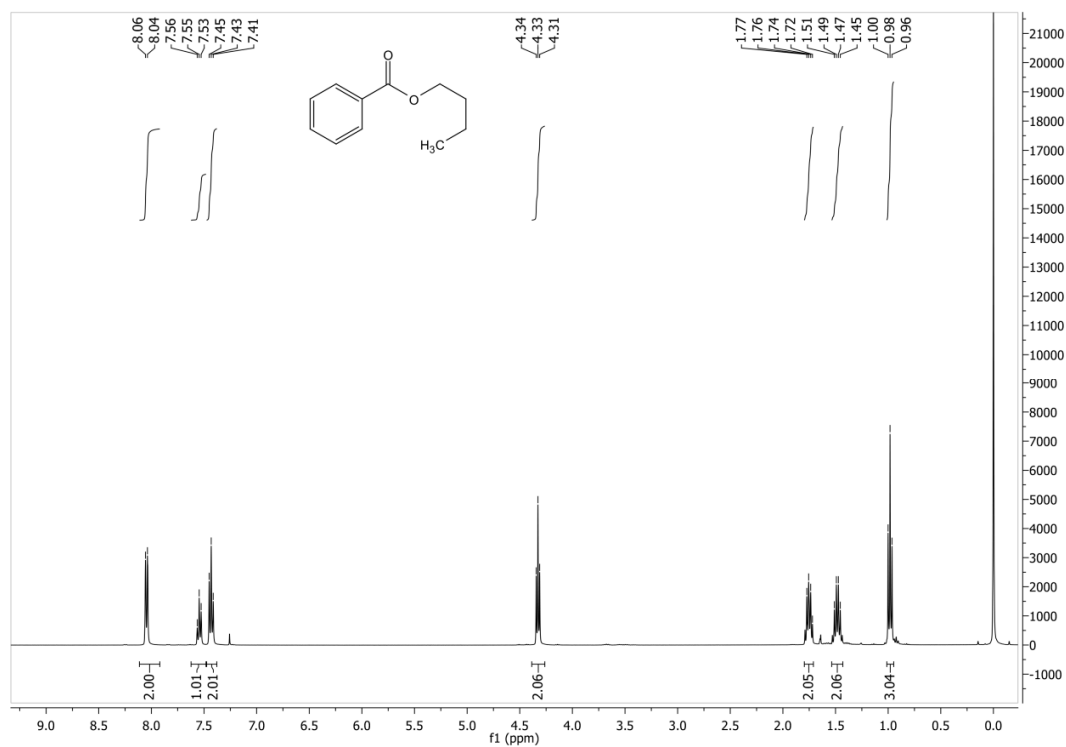
Methyl benzoate – 1a



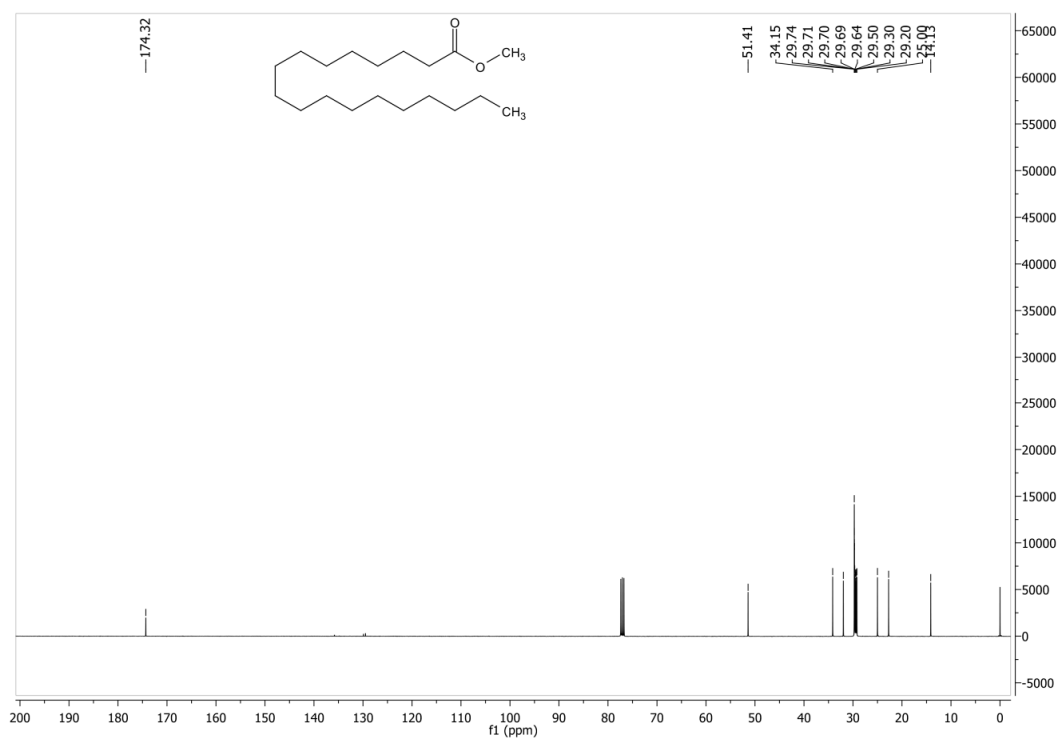
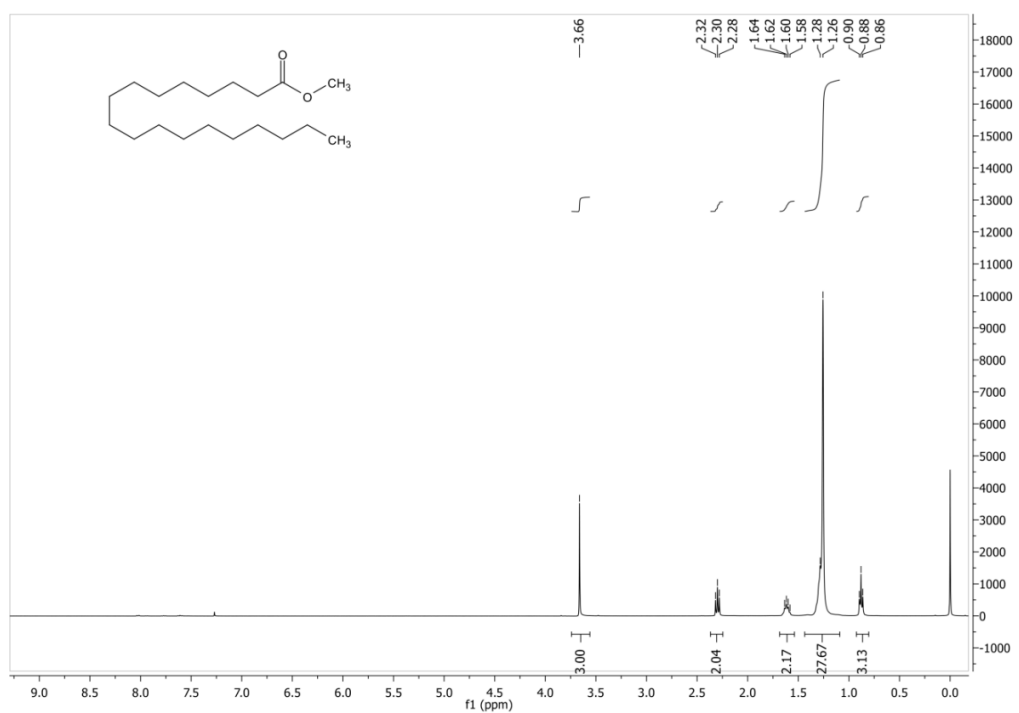
Ethyl benzoate – 1b



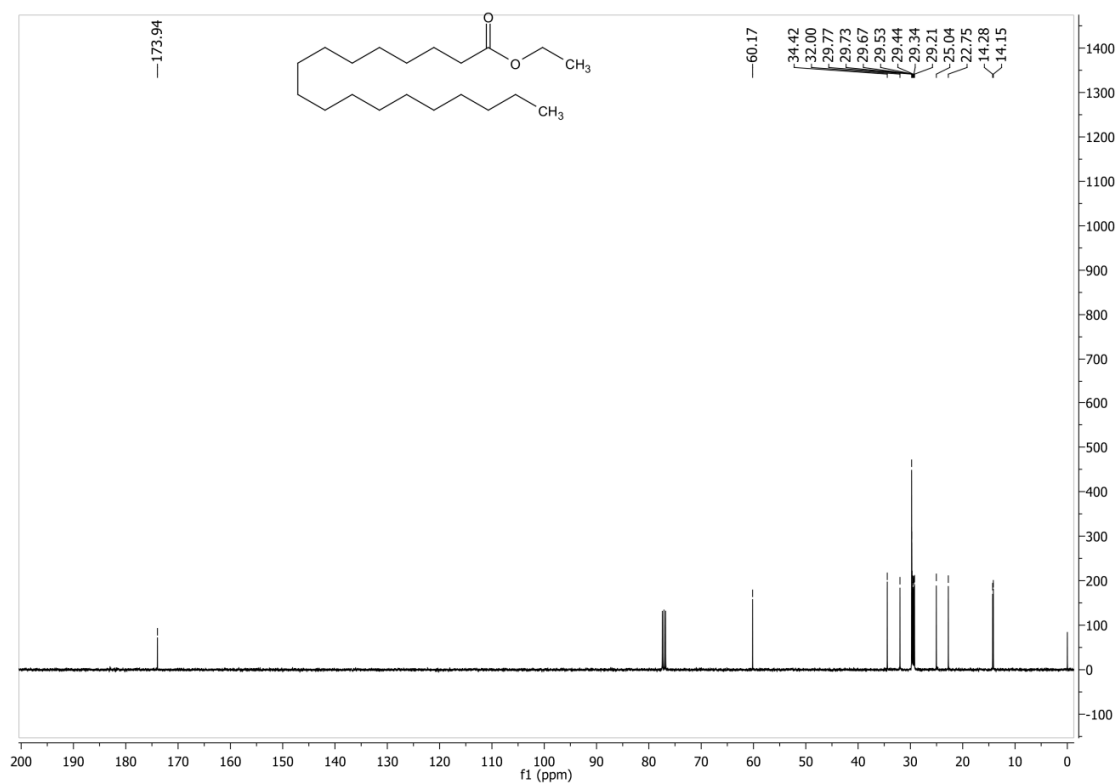
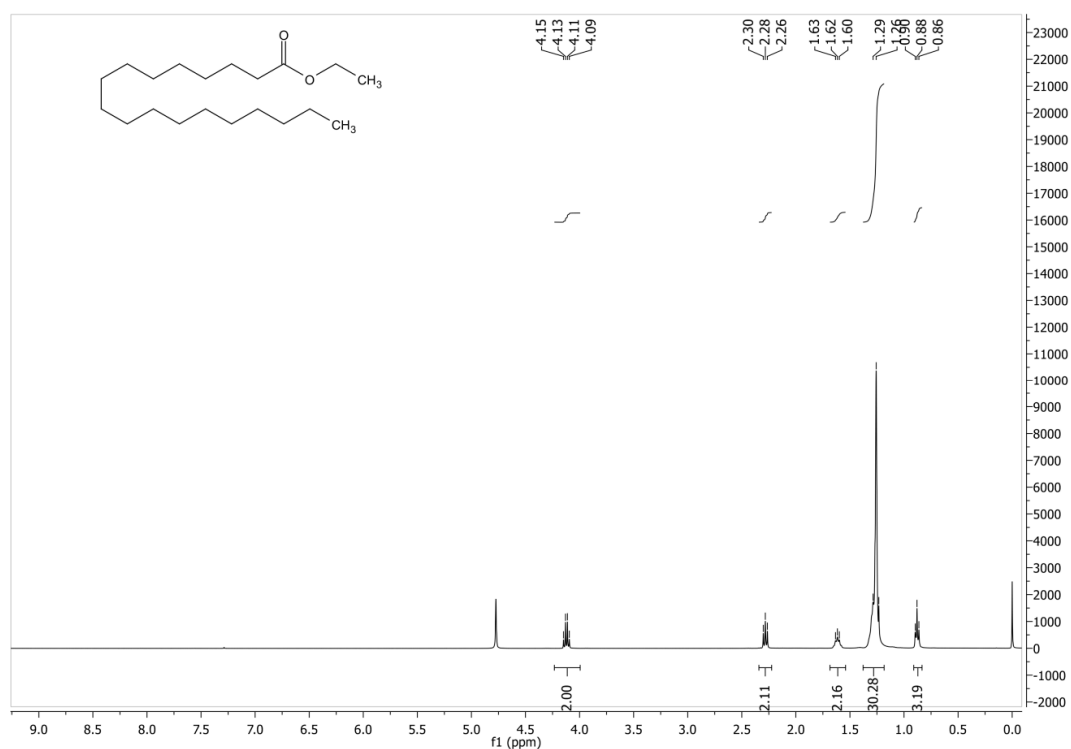
Butyl benzoate – 1c



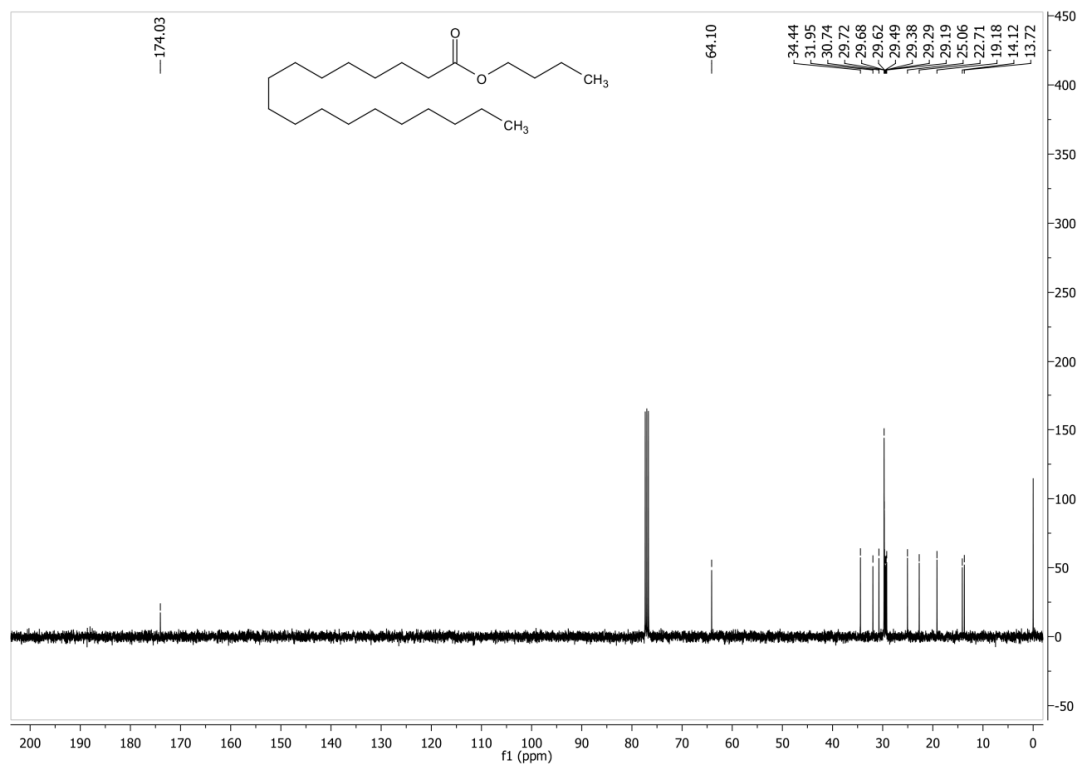
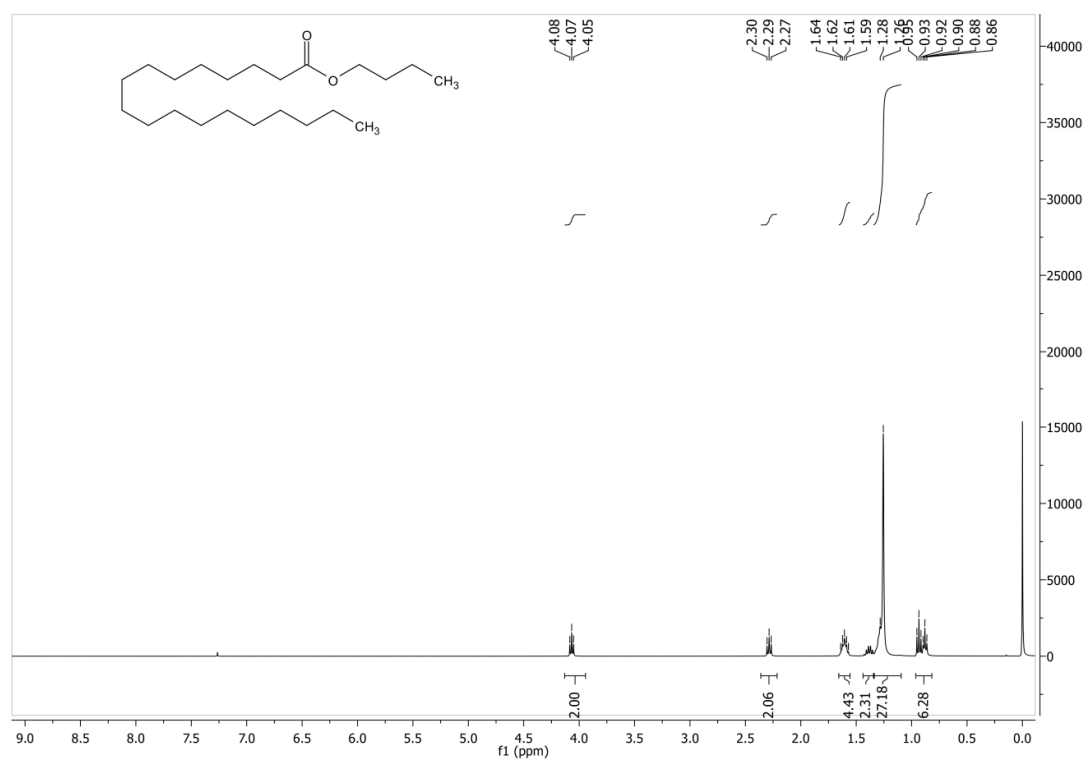
Methyl stearate – 2a



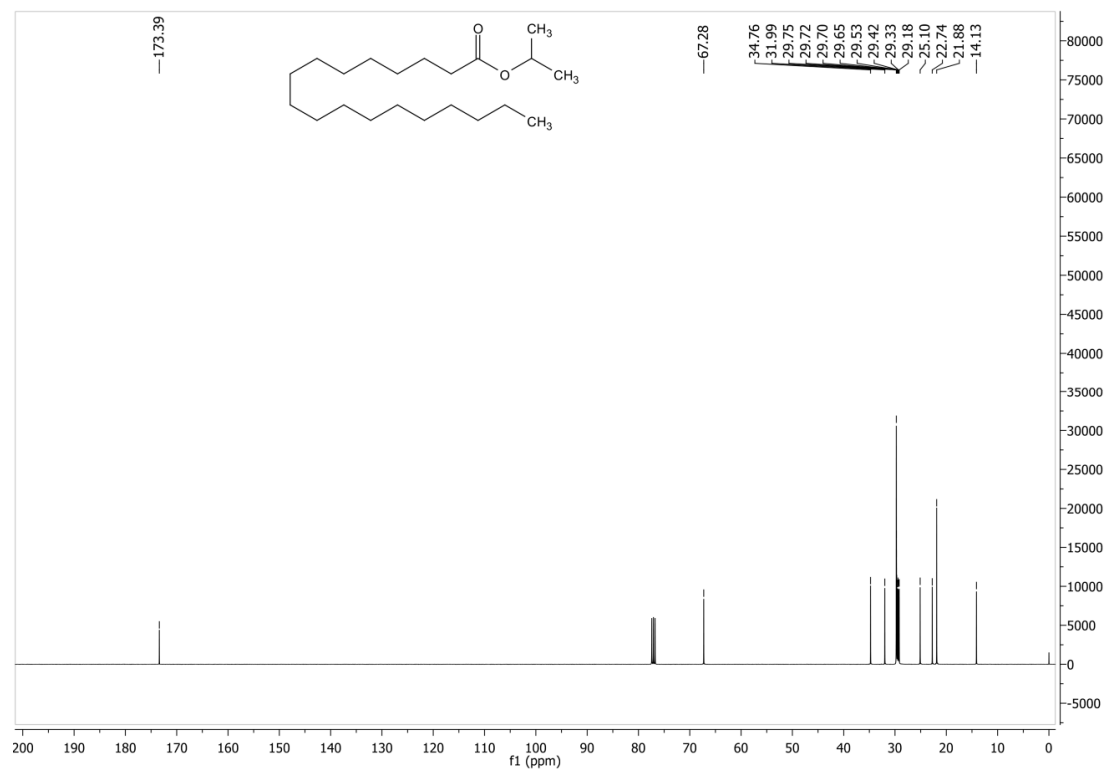
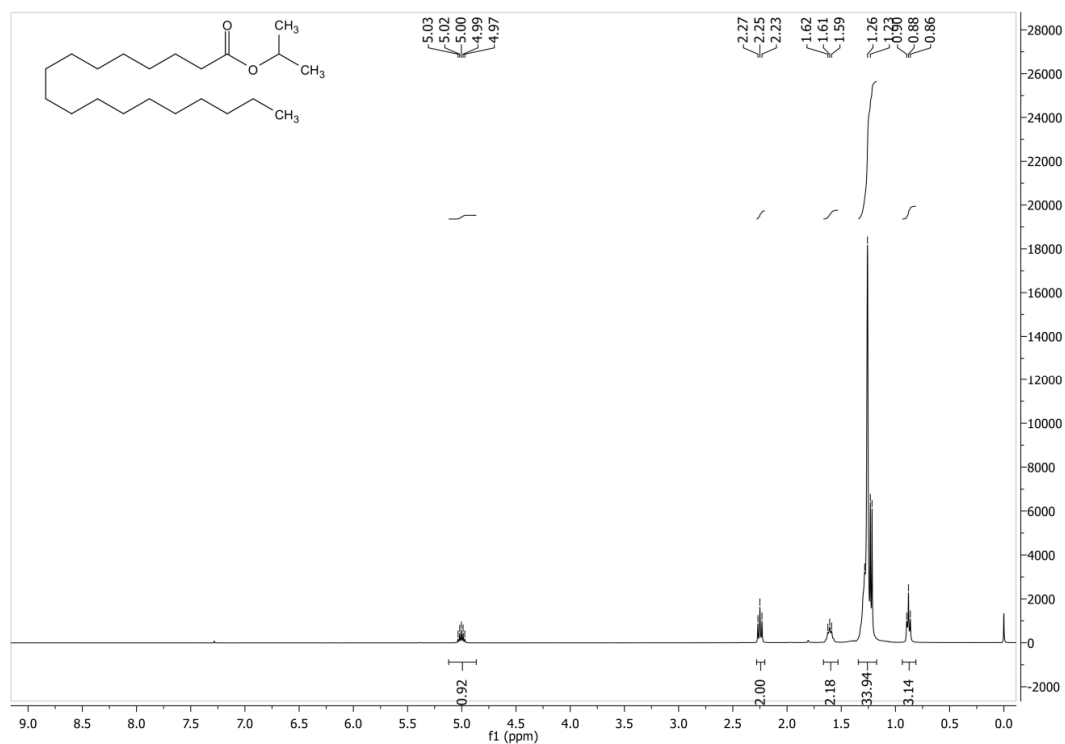
Ethyl stearate – 2b



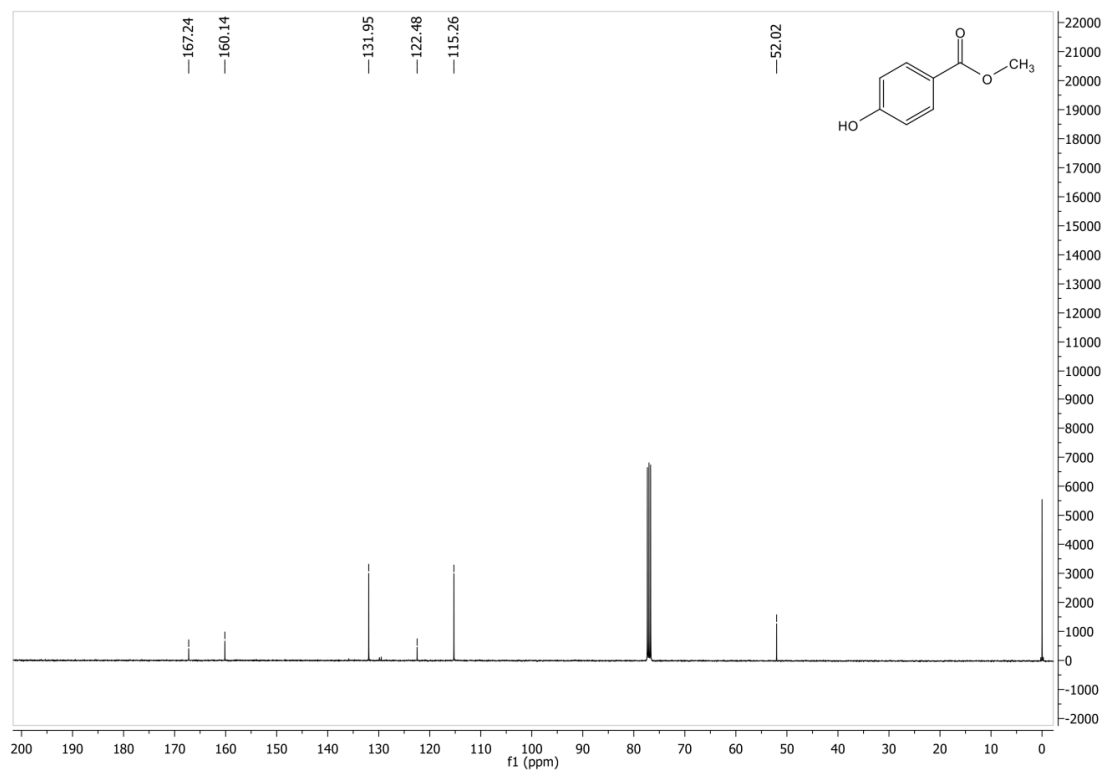
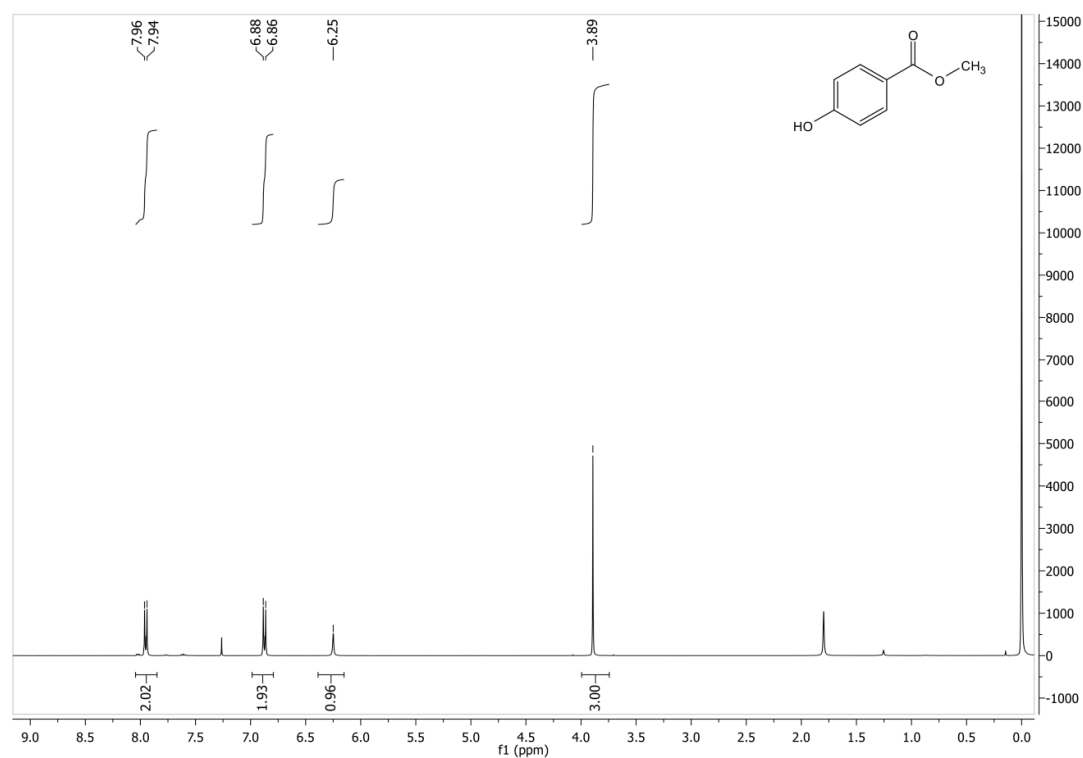
Butyl stearate – 2c



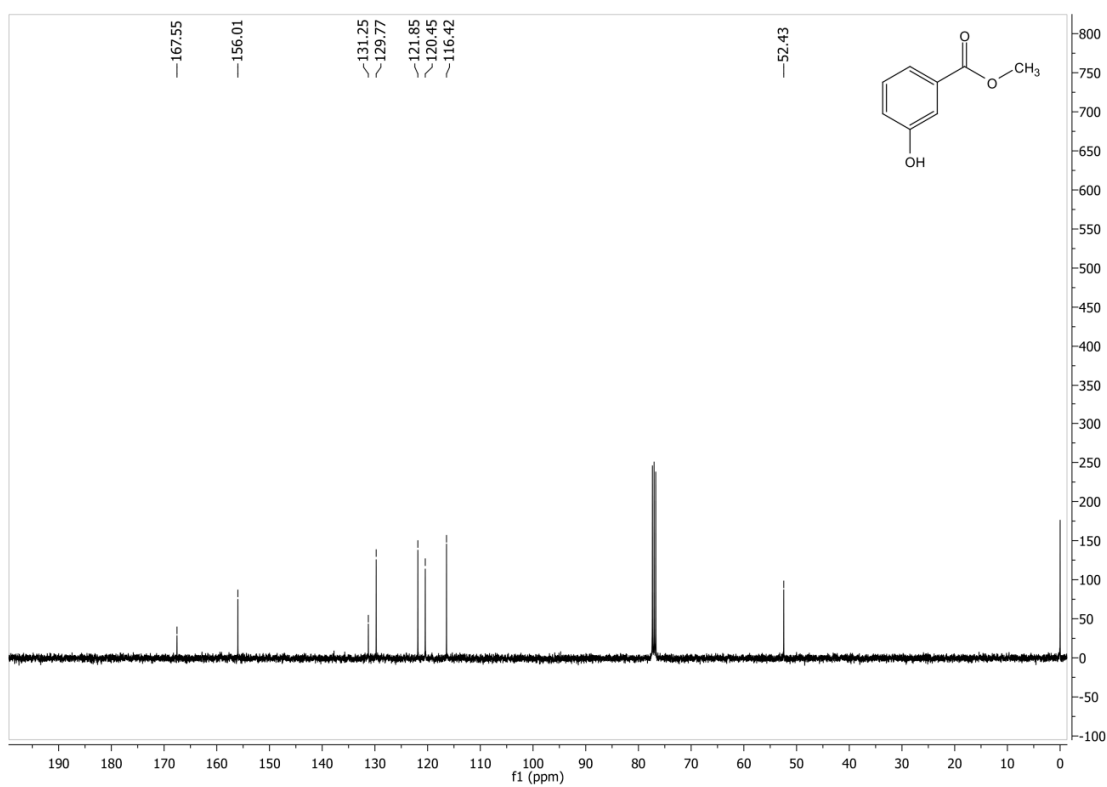
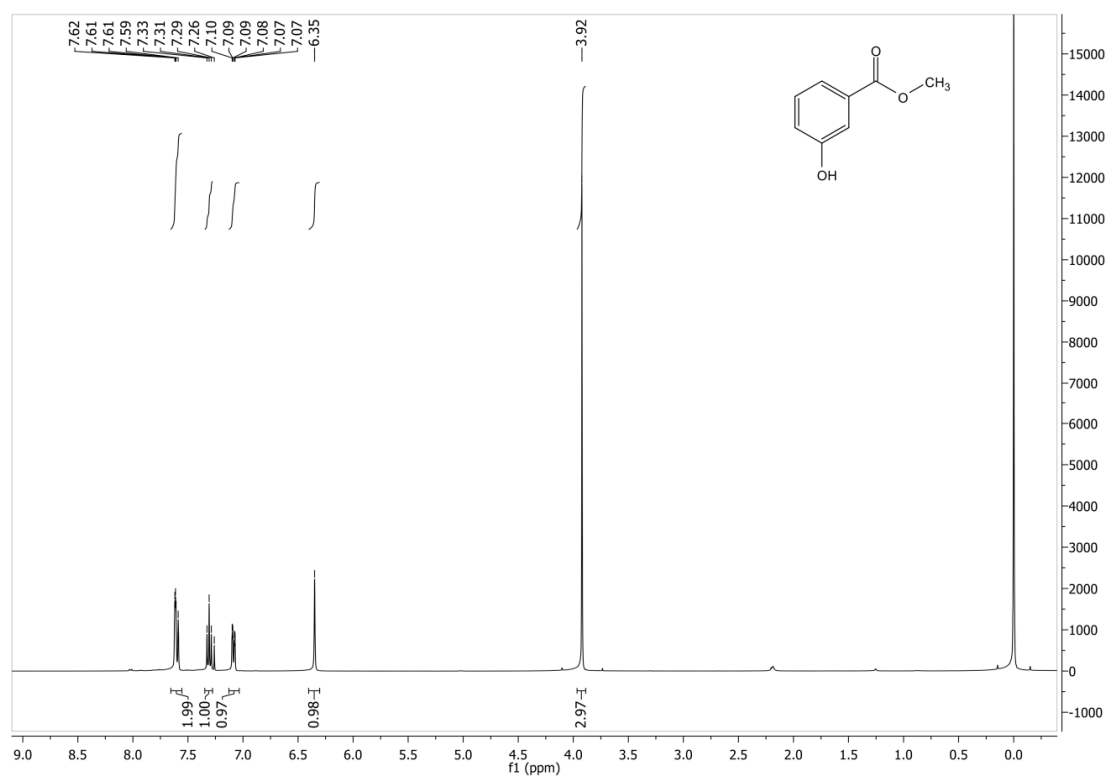
Isopropyl stearate – 2d



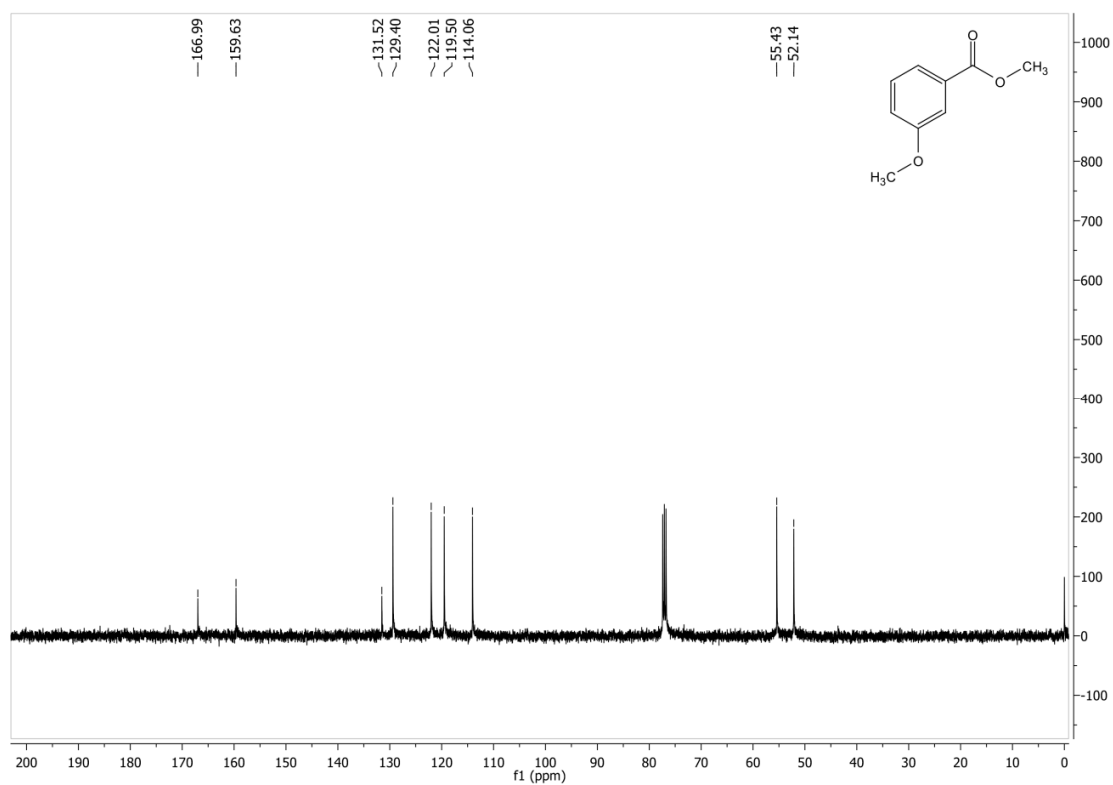
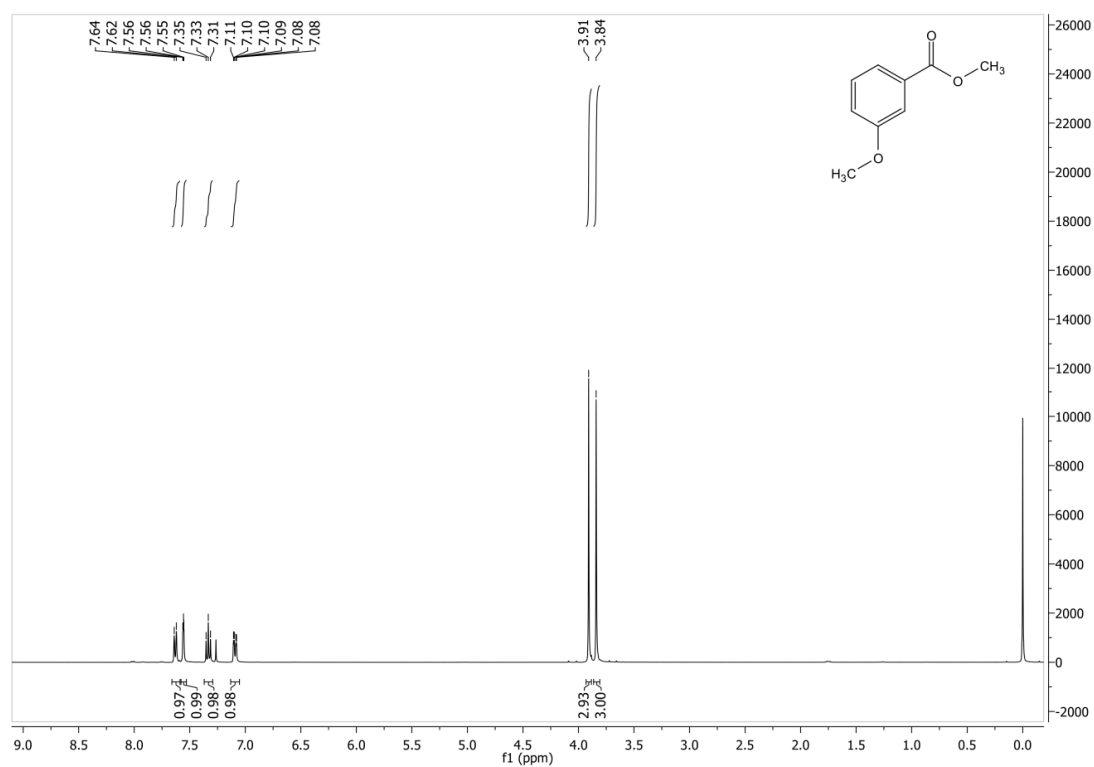
Methyl 4-hidroxybenzoate – 3a



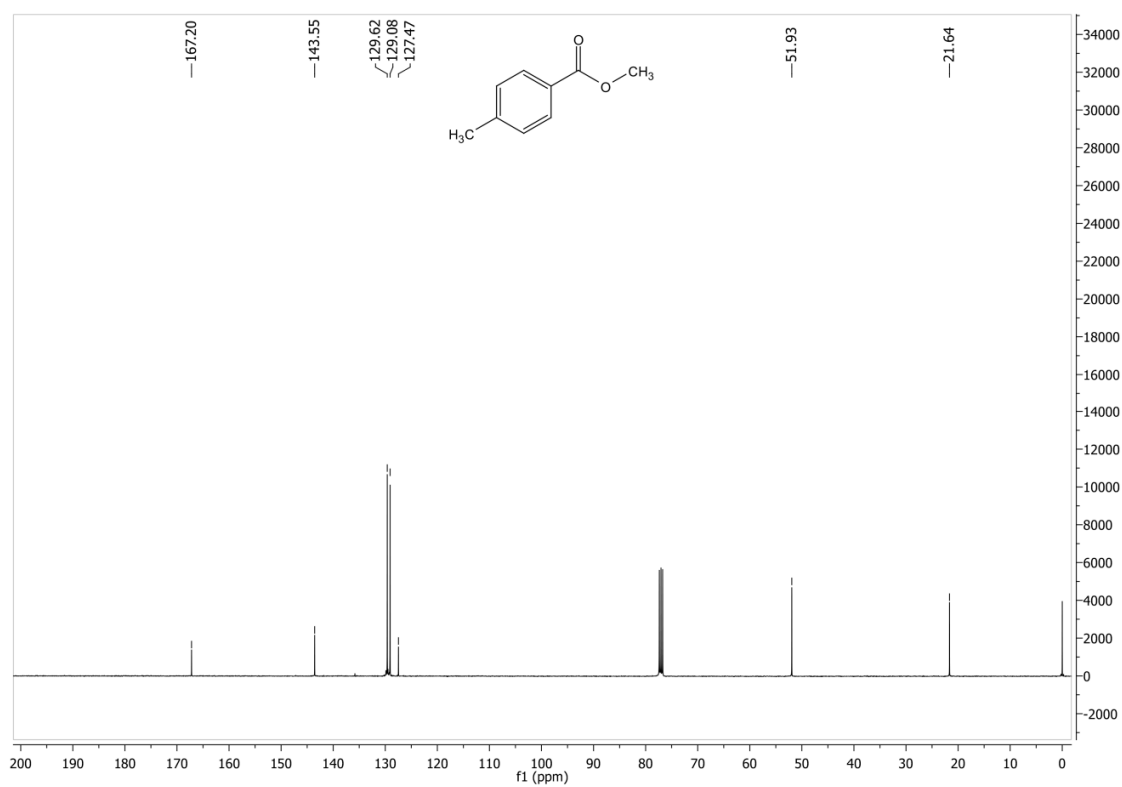
Methyl 3-hidroxybenzoate – 4a



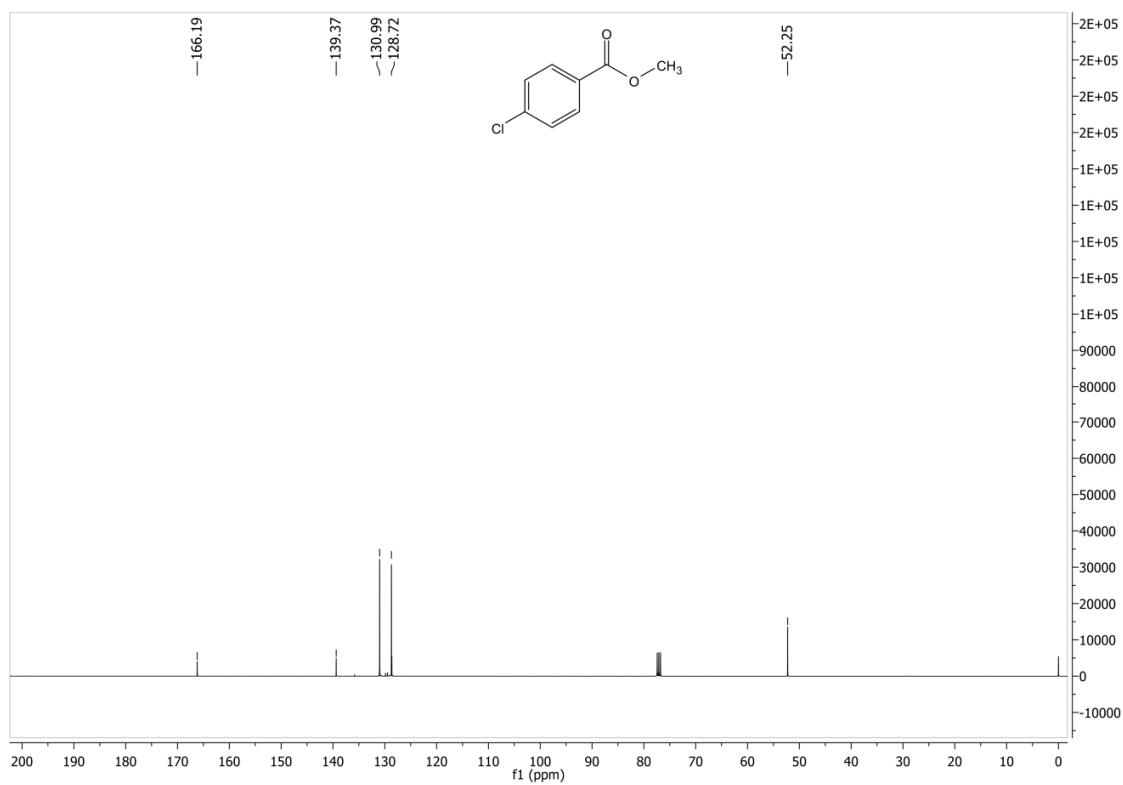
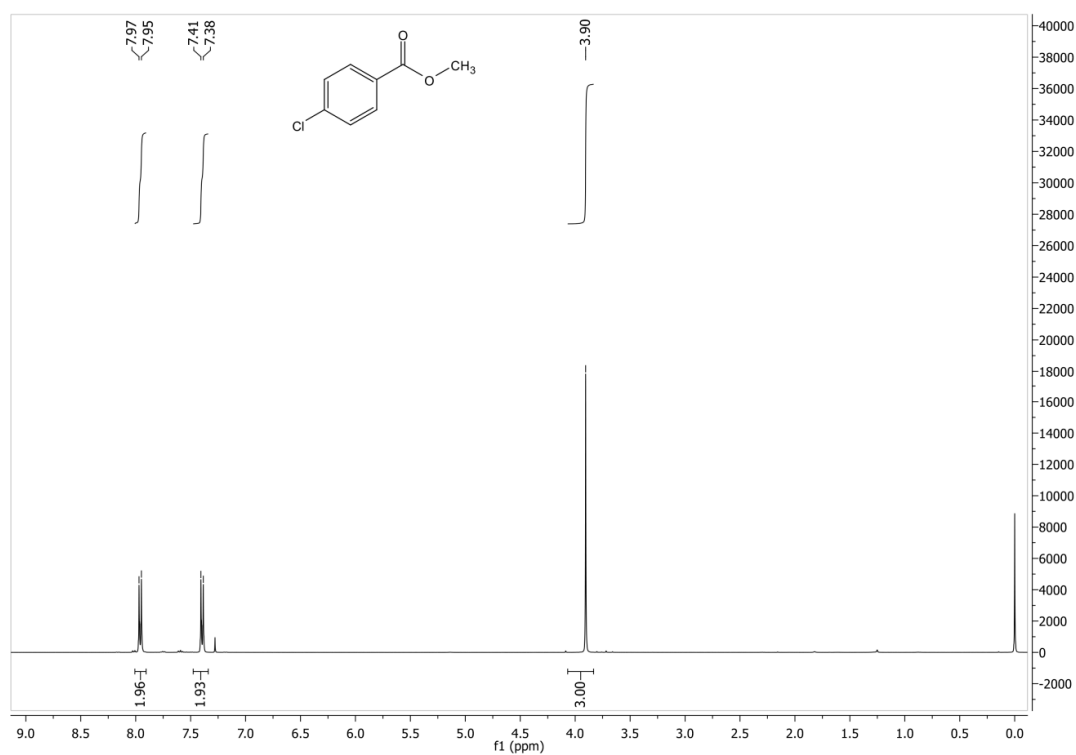
Methyl 3-methoxybenzoate – 5a



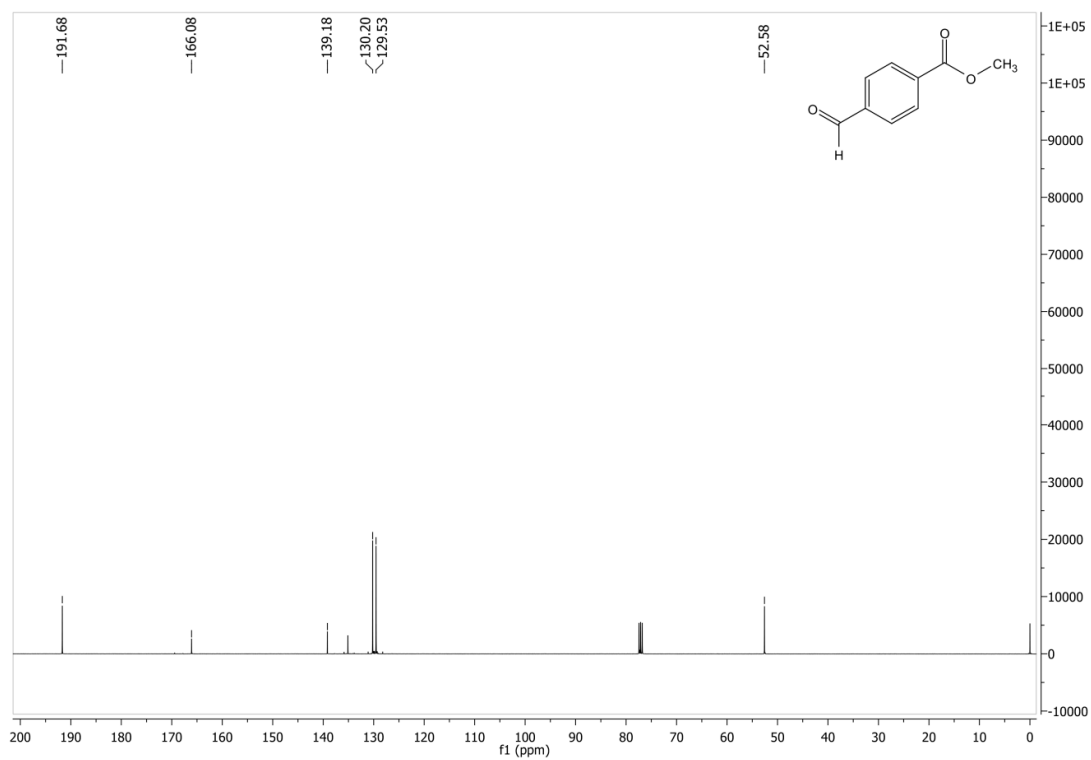
Methyl 4-methylbenzoate – 6a



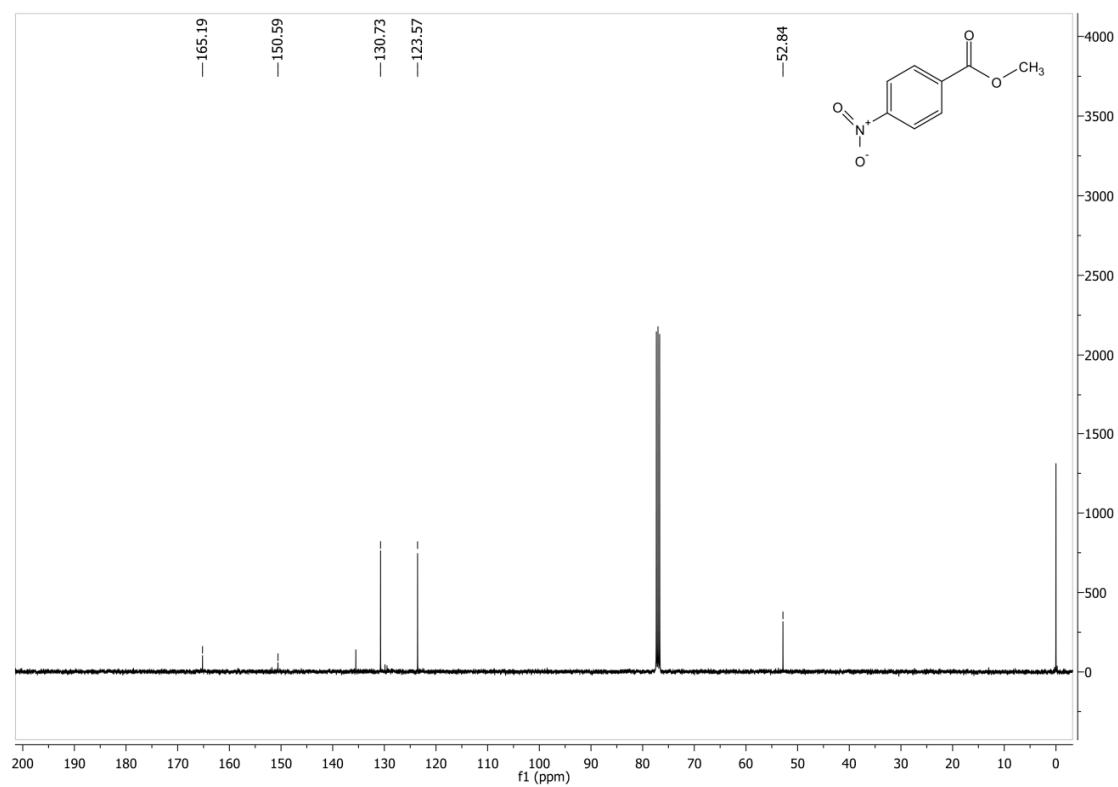
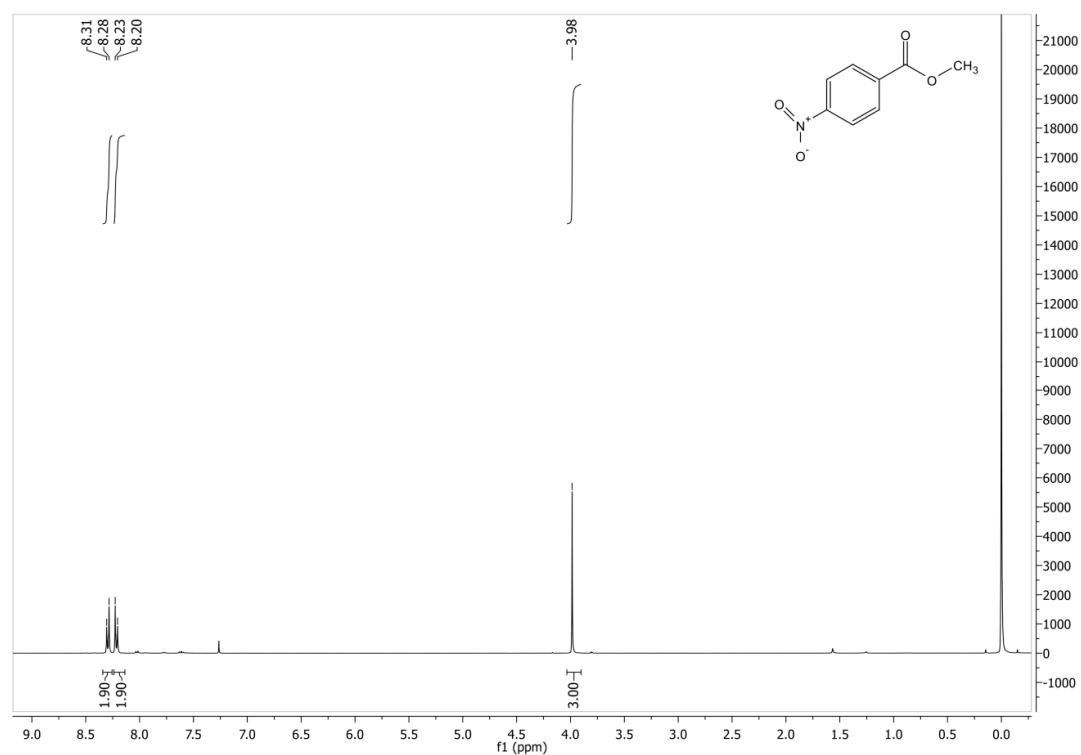
Methyl 4-chlorobenzoate – 7a



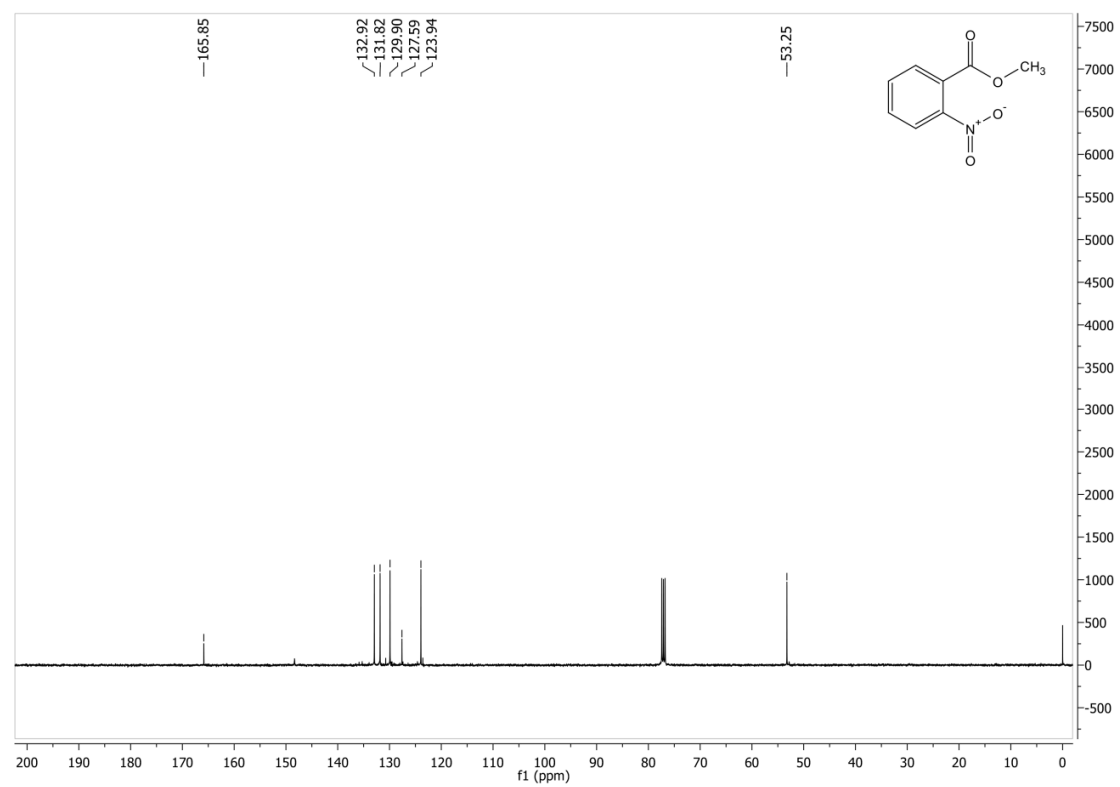
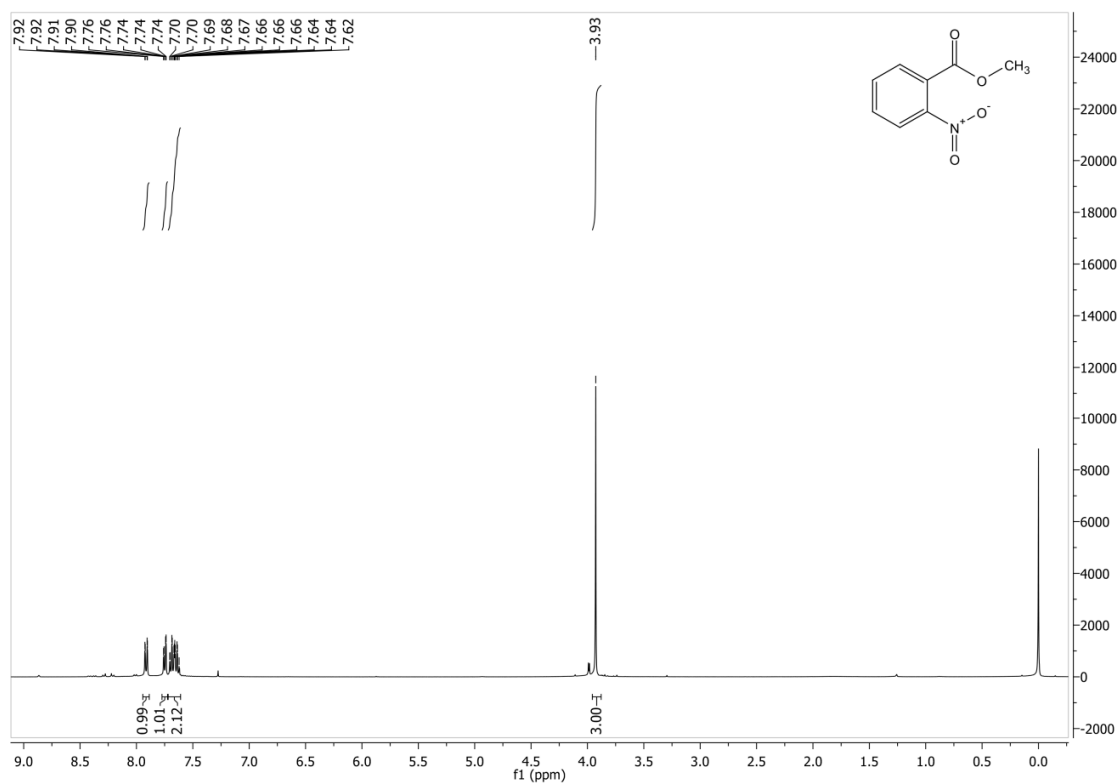
Methyl 4-formylbenzoate – 8a



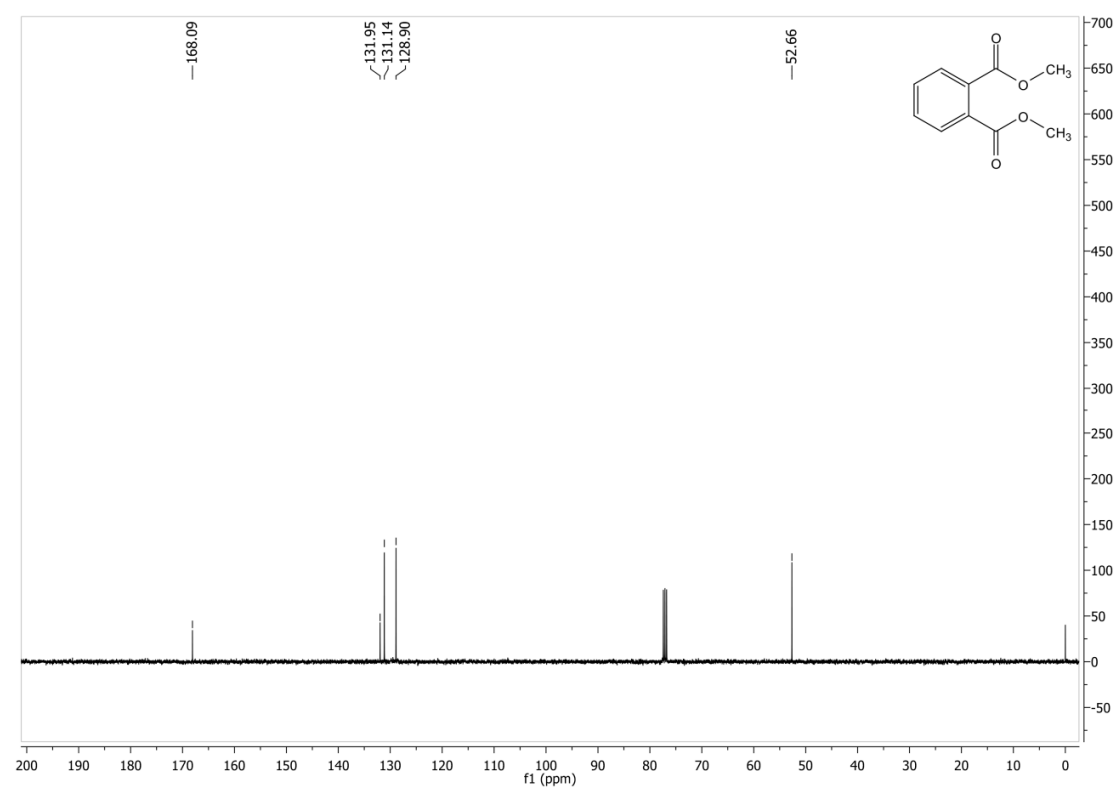
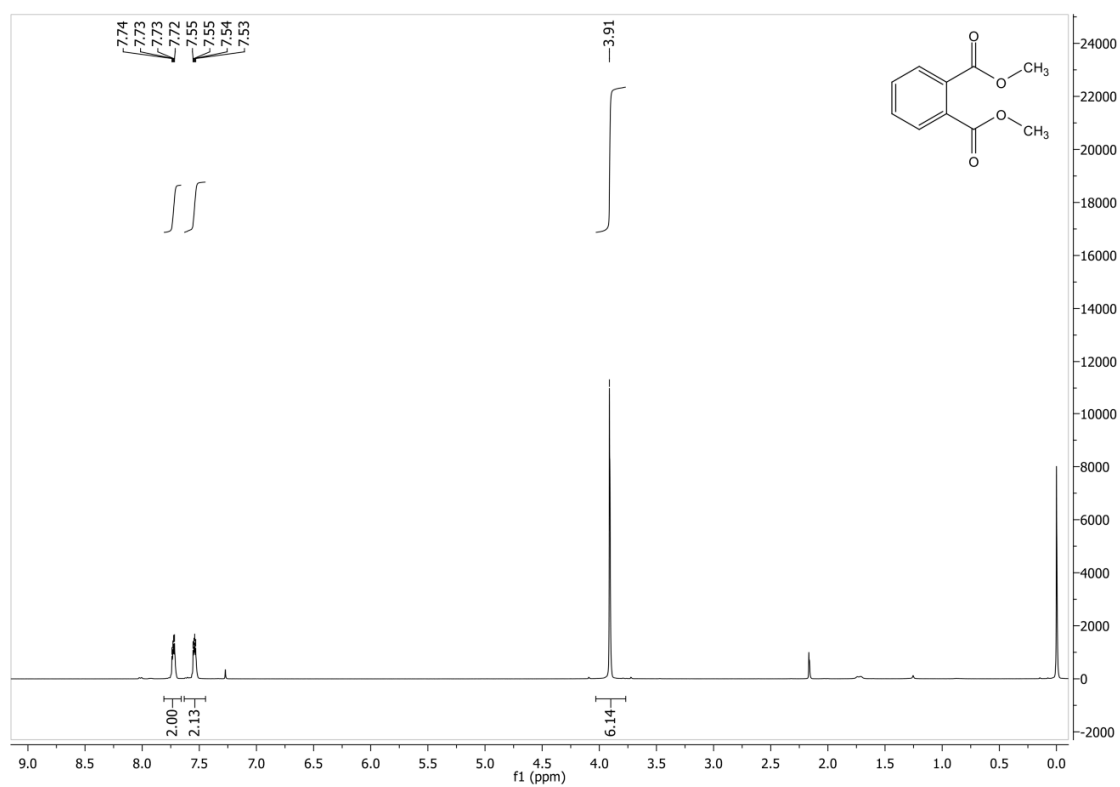
Methyl 4-nitrobenzoate – 9a



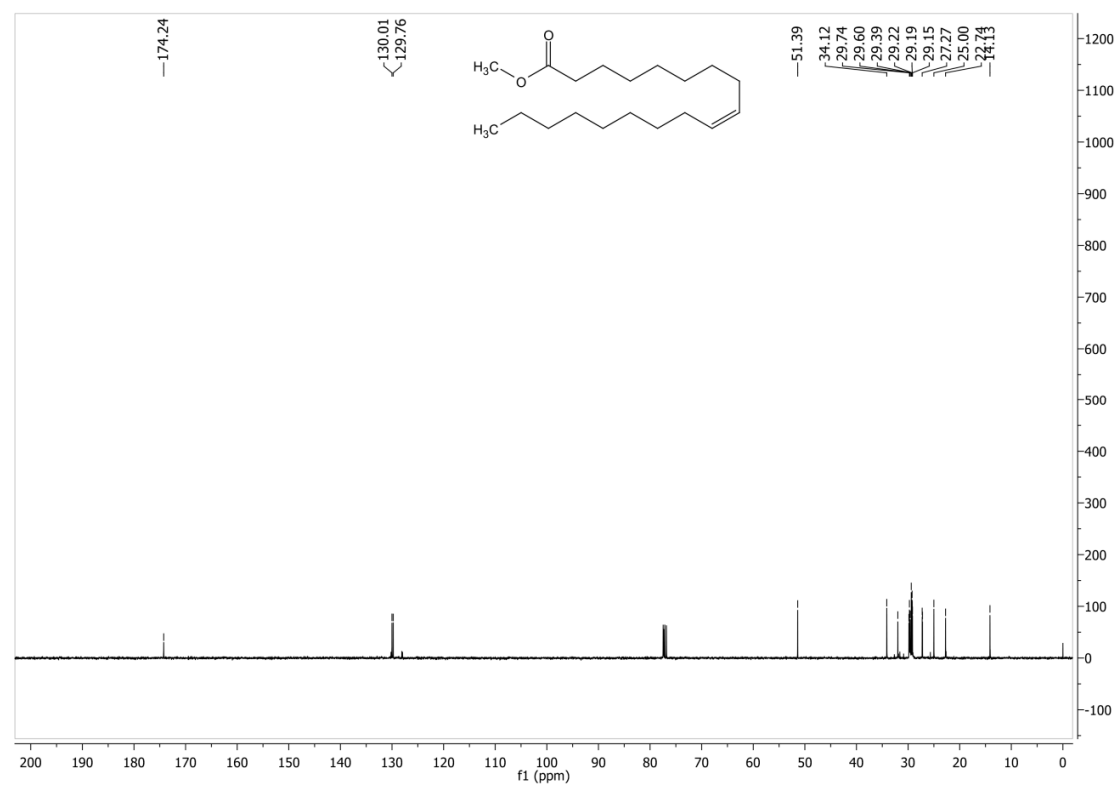
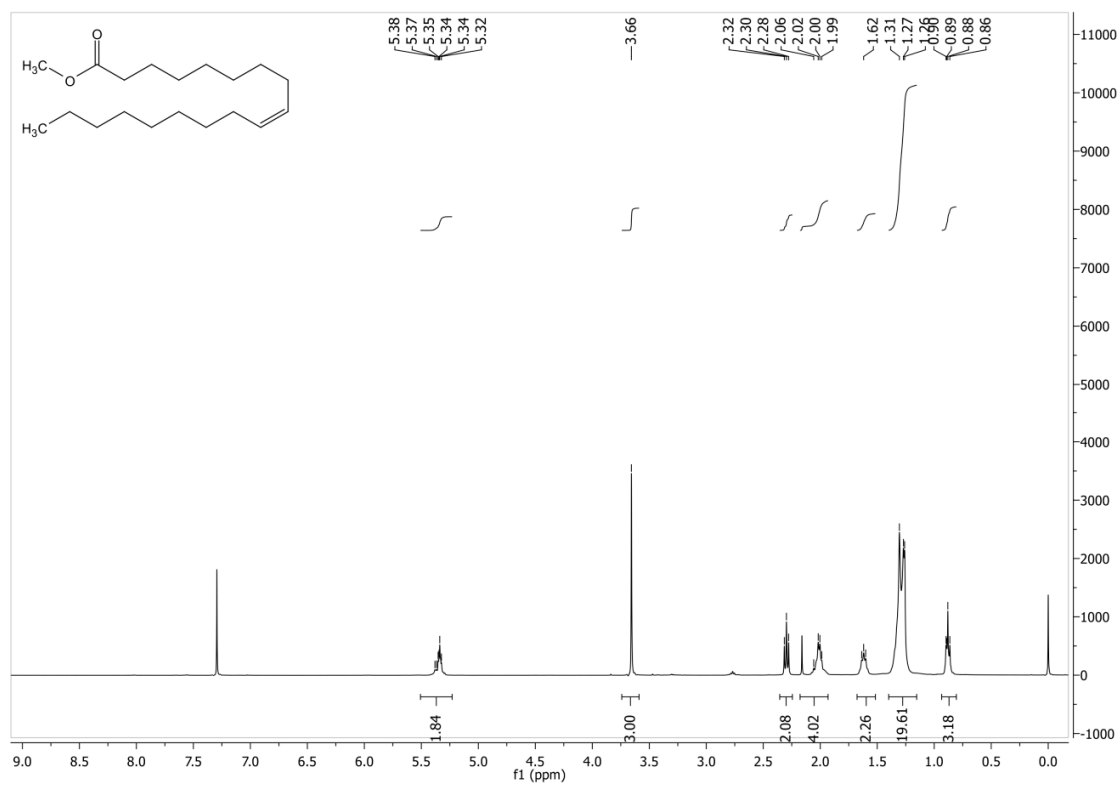
Methyl 2-nitrobenzoate – 10a



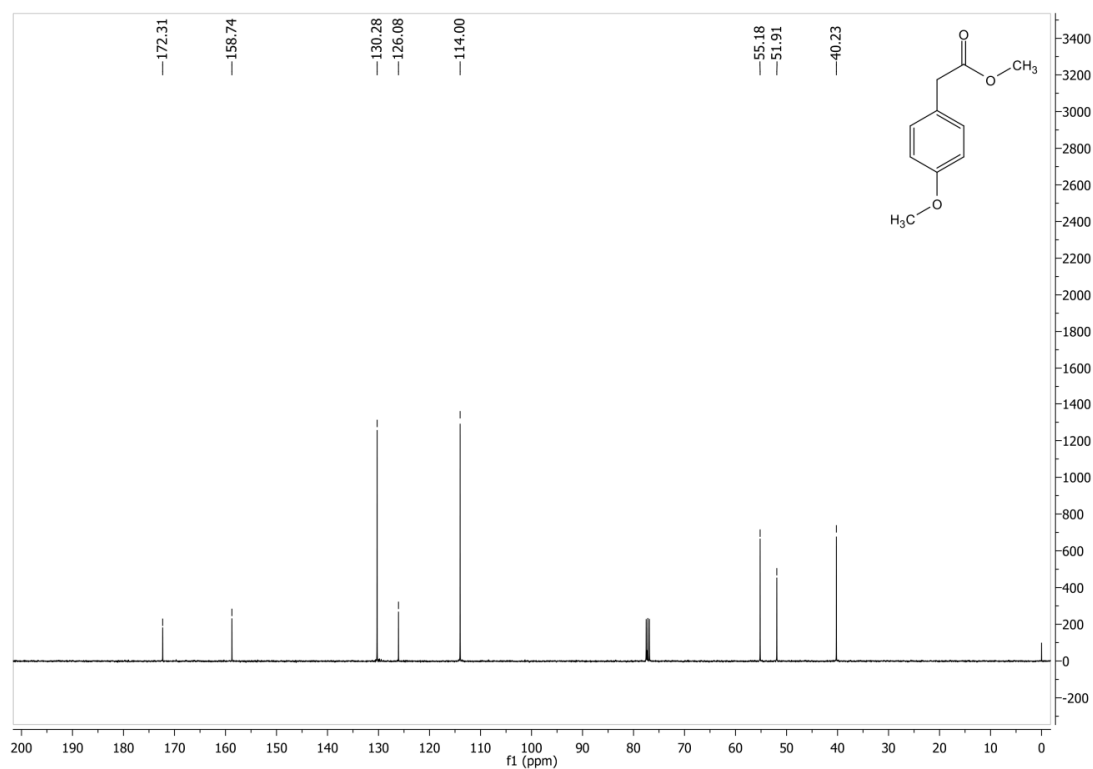
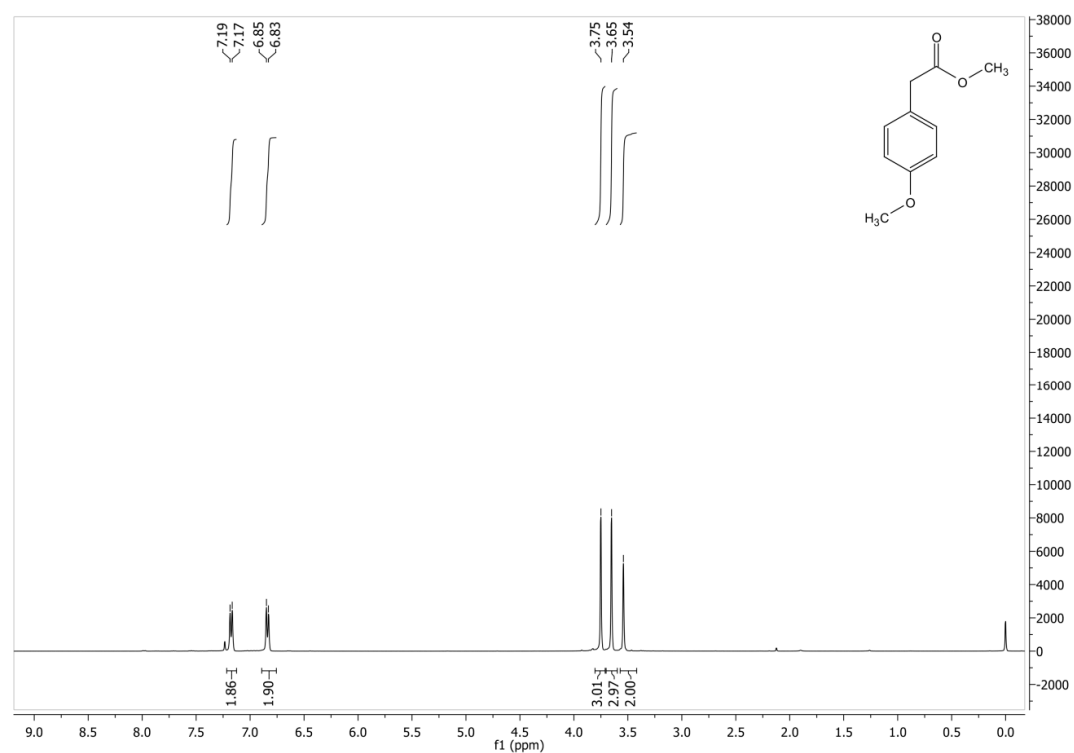
Dimethyl phthalate – 11a



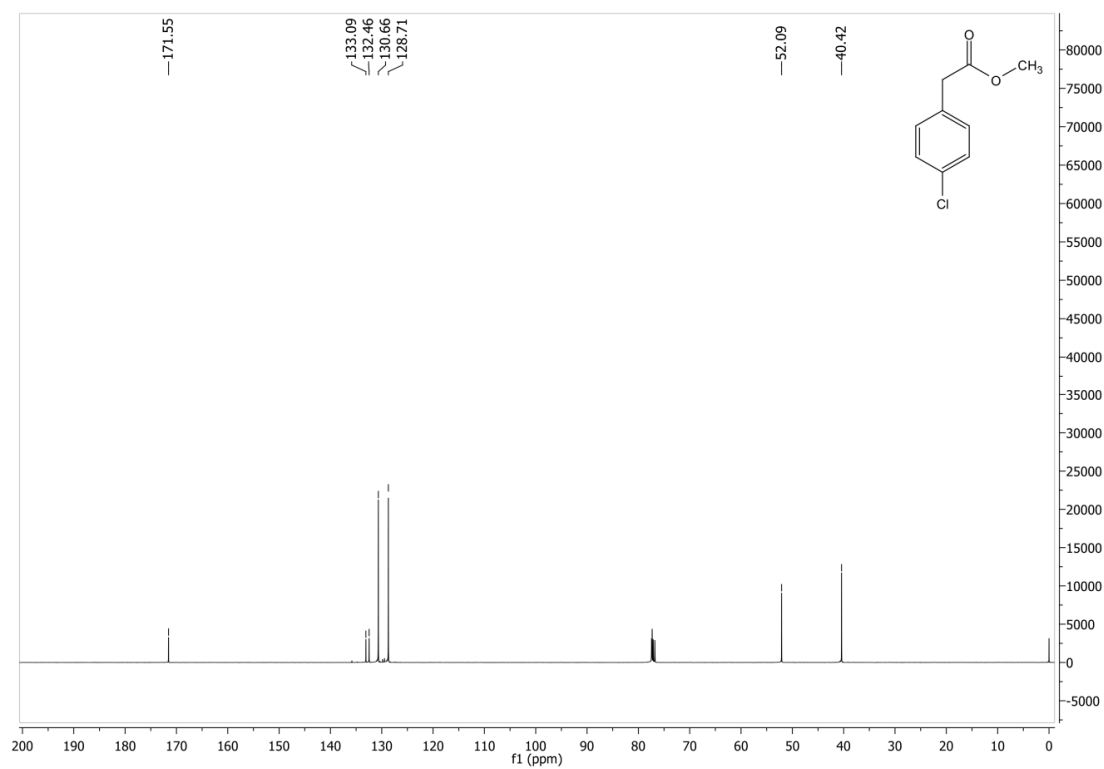
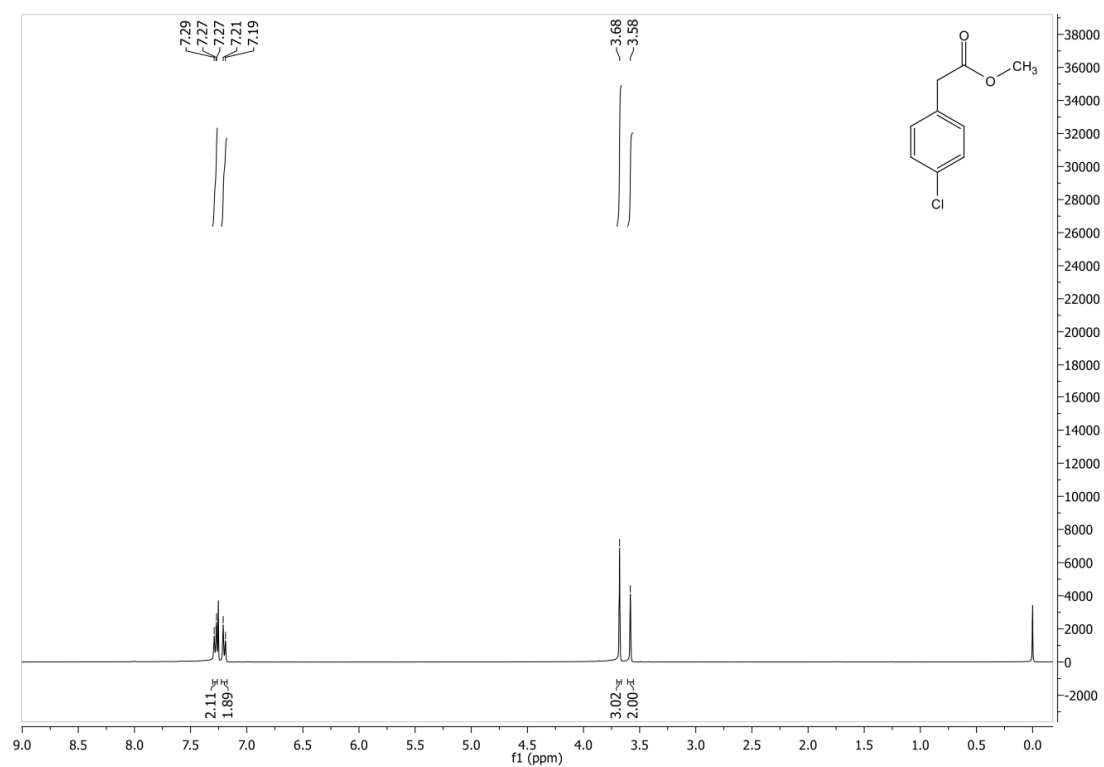
Methyl oleate – 12a



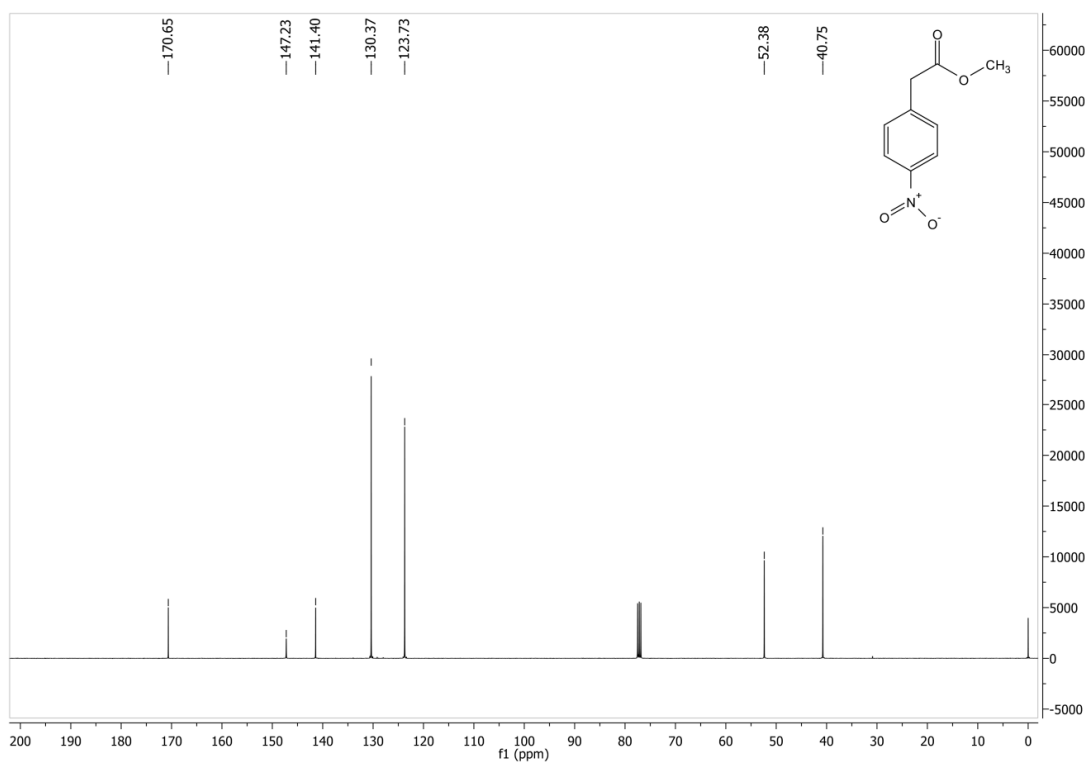
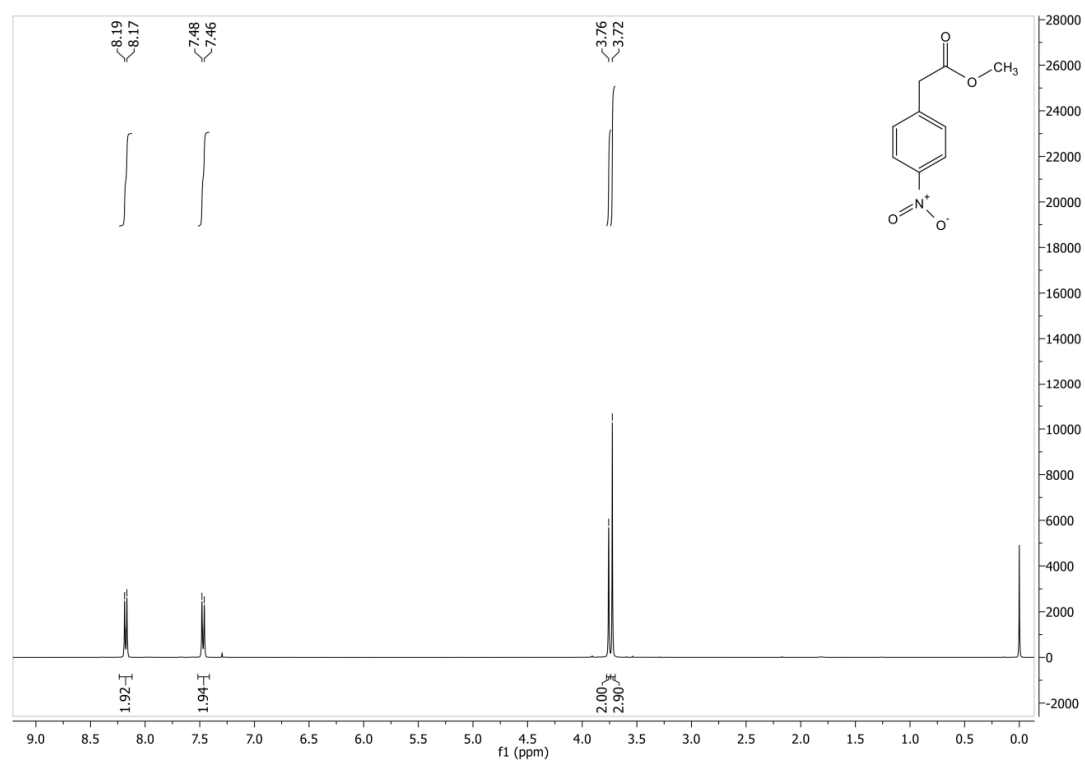
Methyl 4-methoxyphenylacetate – 13a



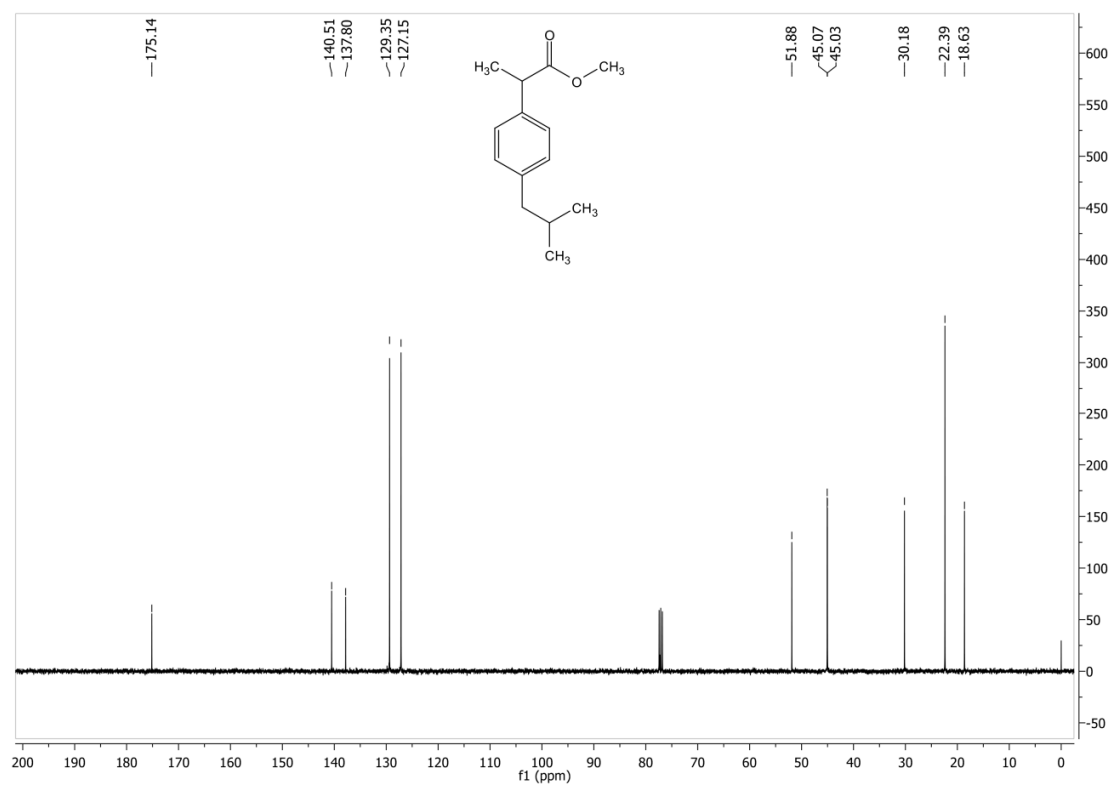
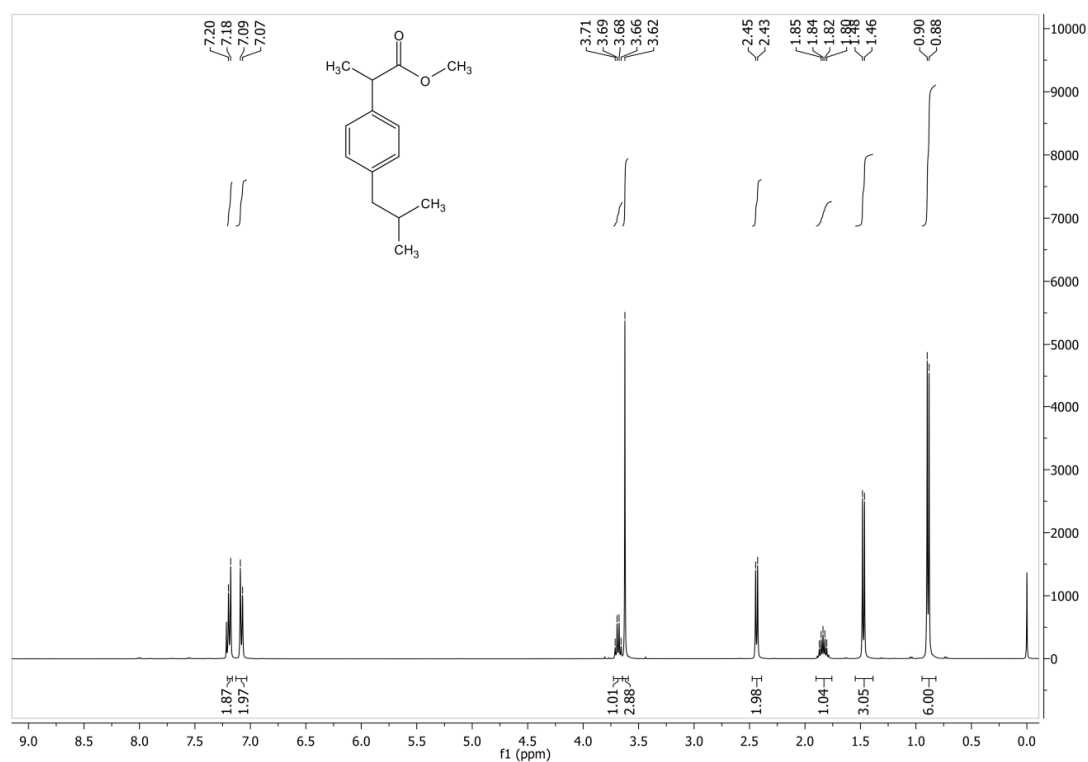
Methyl 4-chlorophenylacetate – 14a



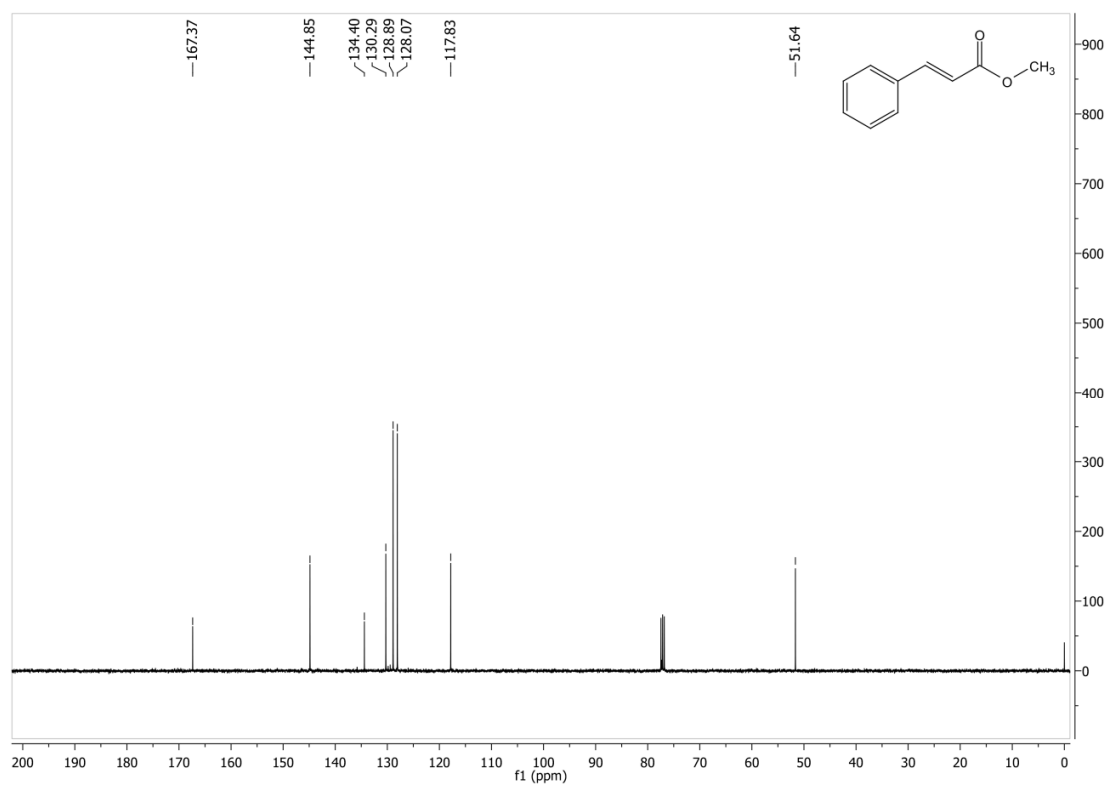
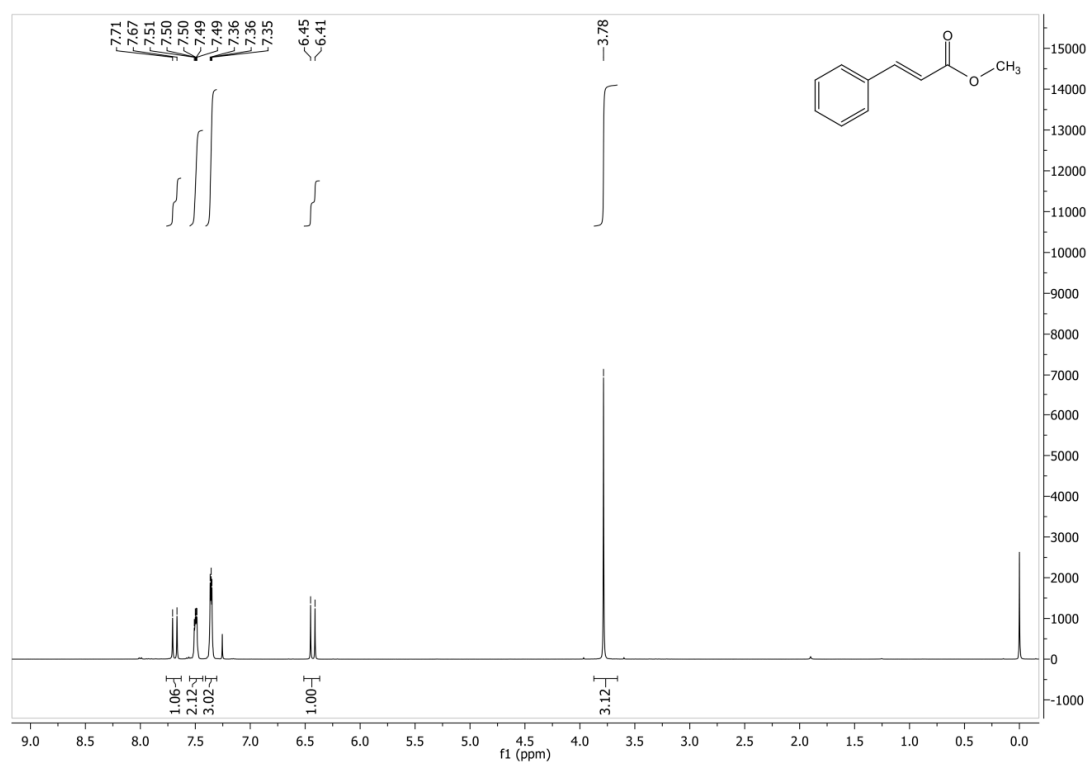
Methyl 4-nitrophenylacetate – 15a



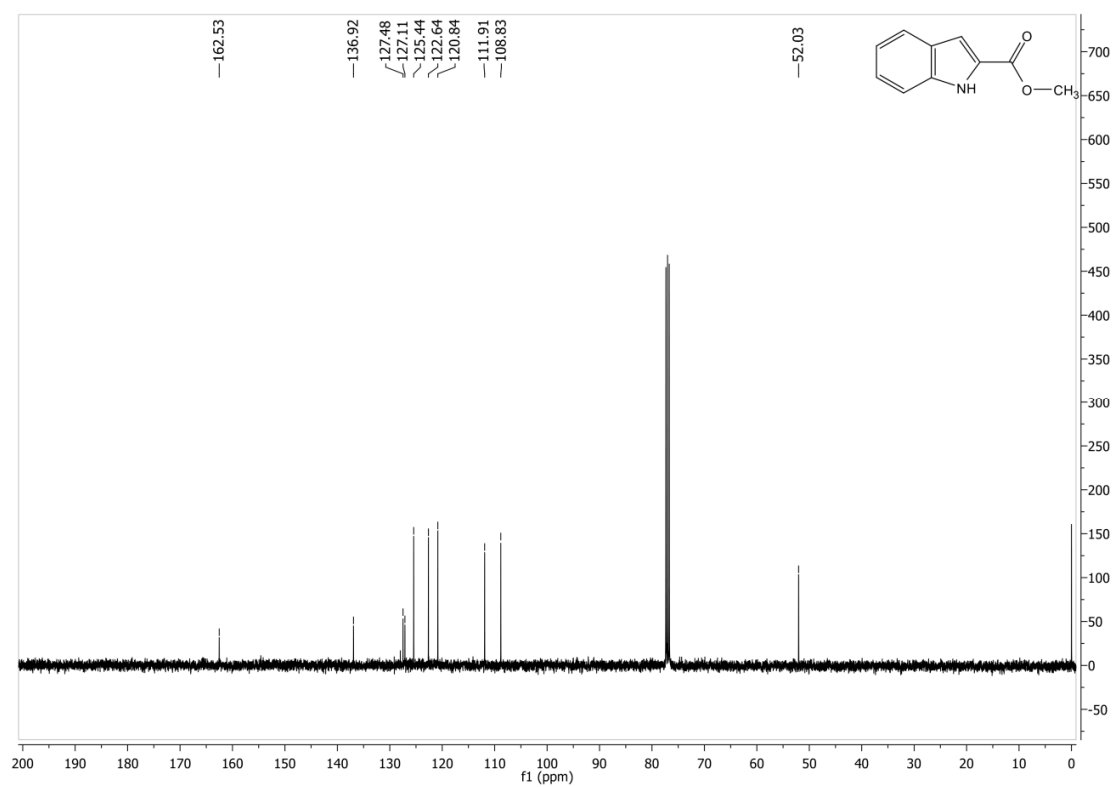
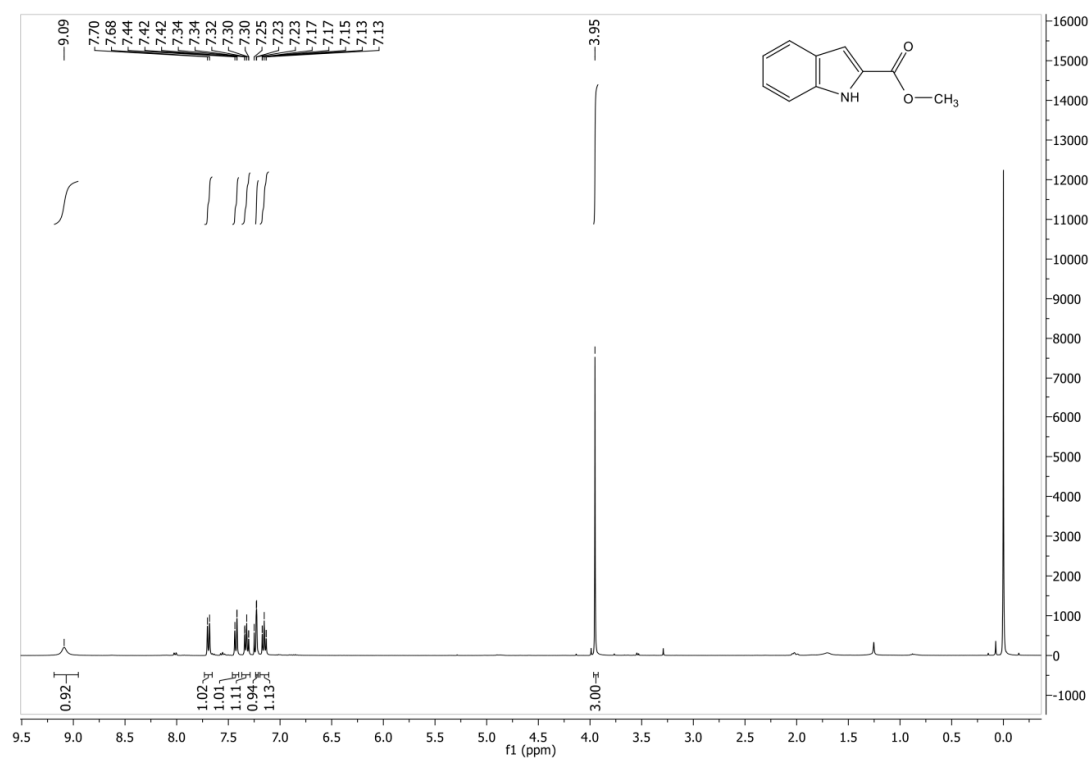
Ibuprofen methyl ester – 16a



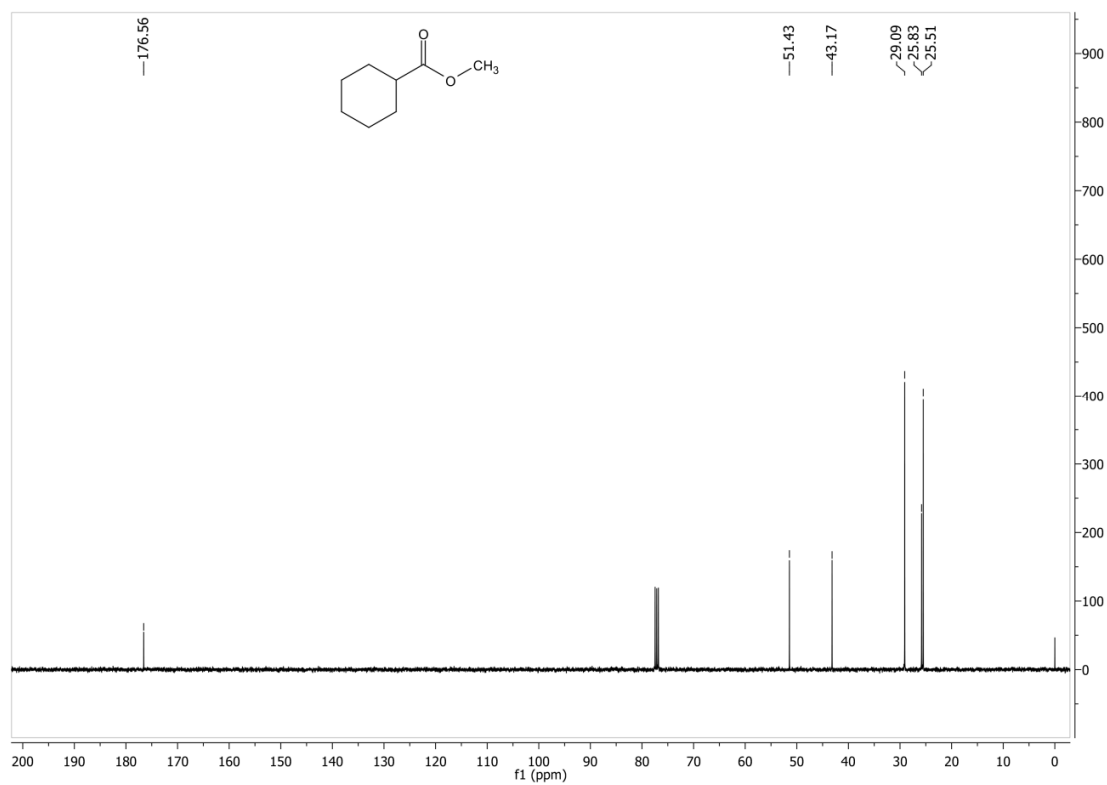
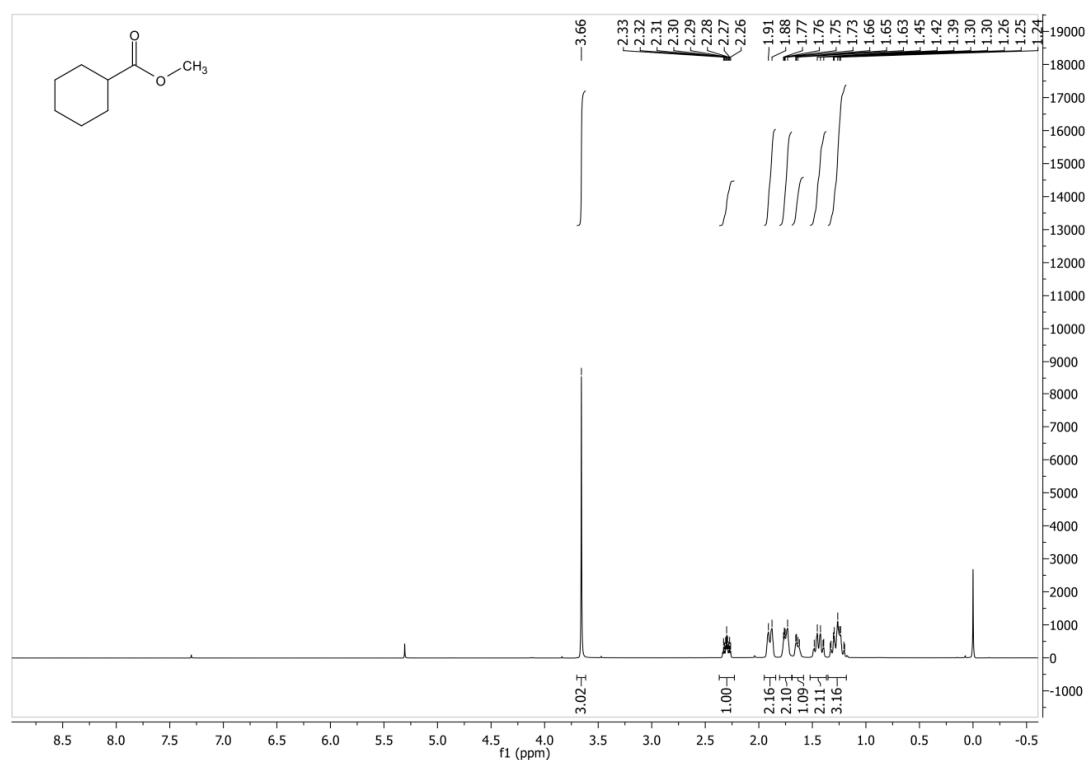
Methyl cinnamate – 17a



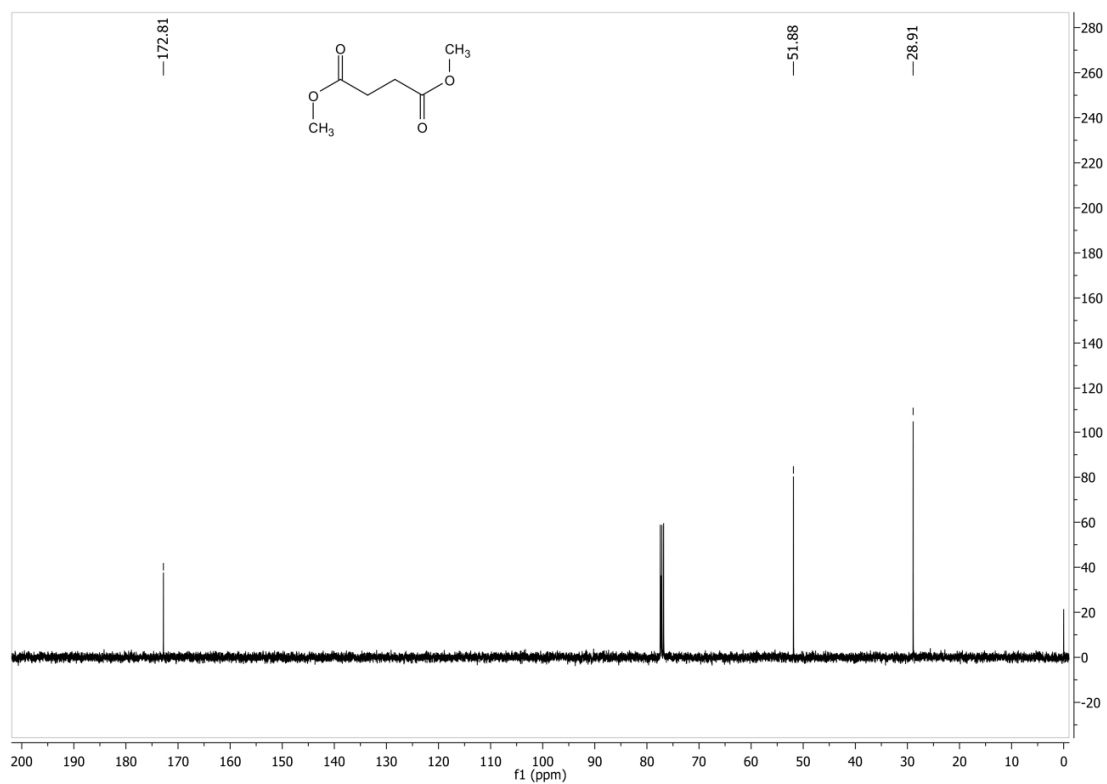
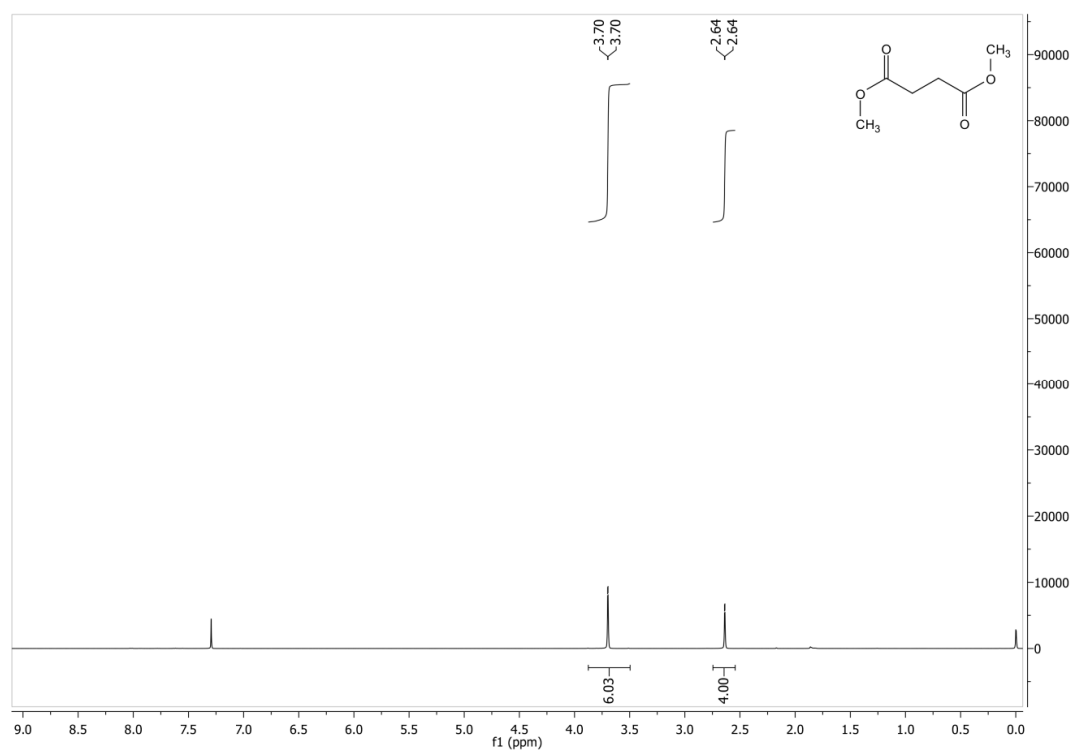
Methyl indol-2-carboxylate – 18a



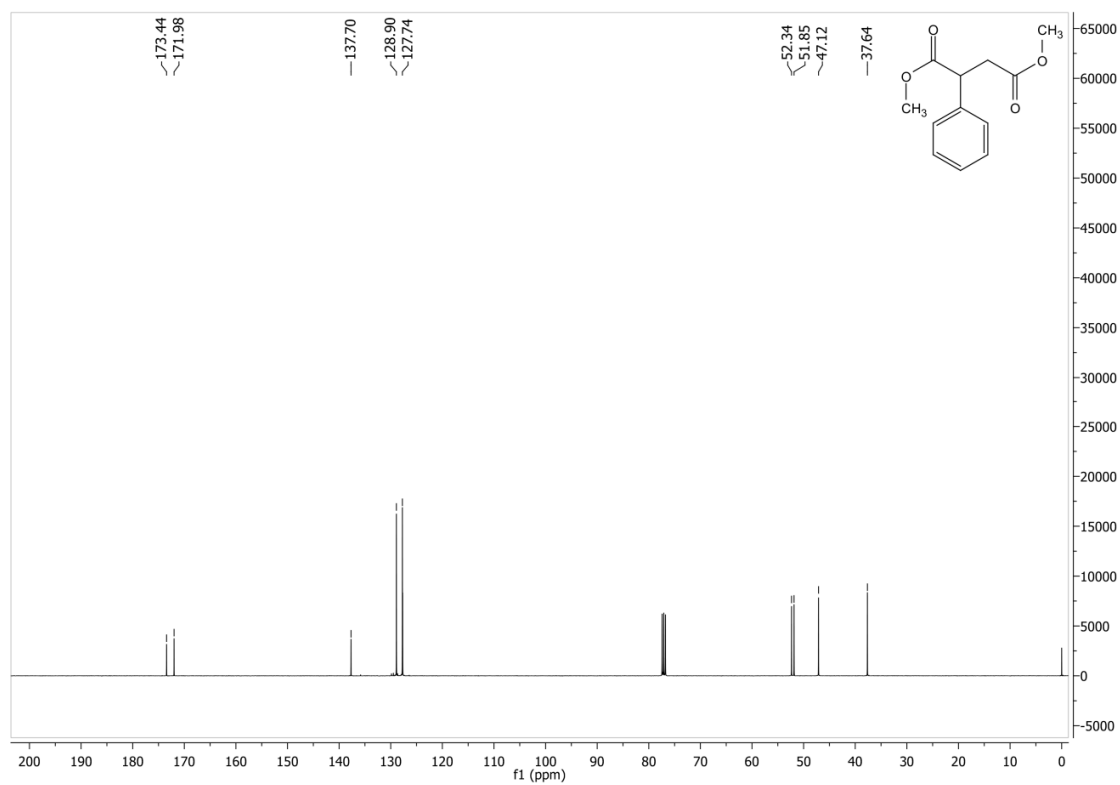
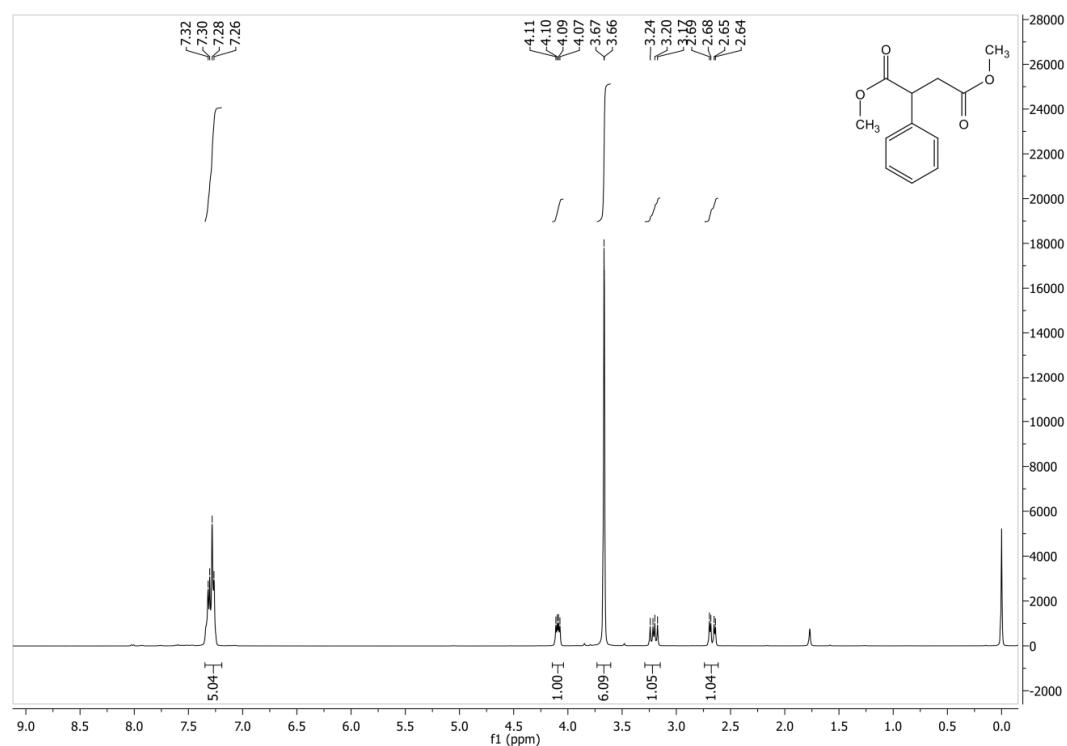
Methyl cyclohexylcarboxylate – 19a



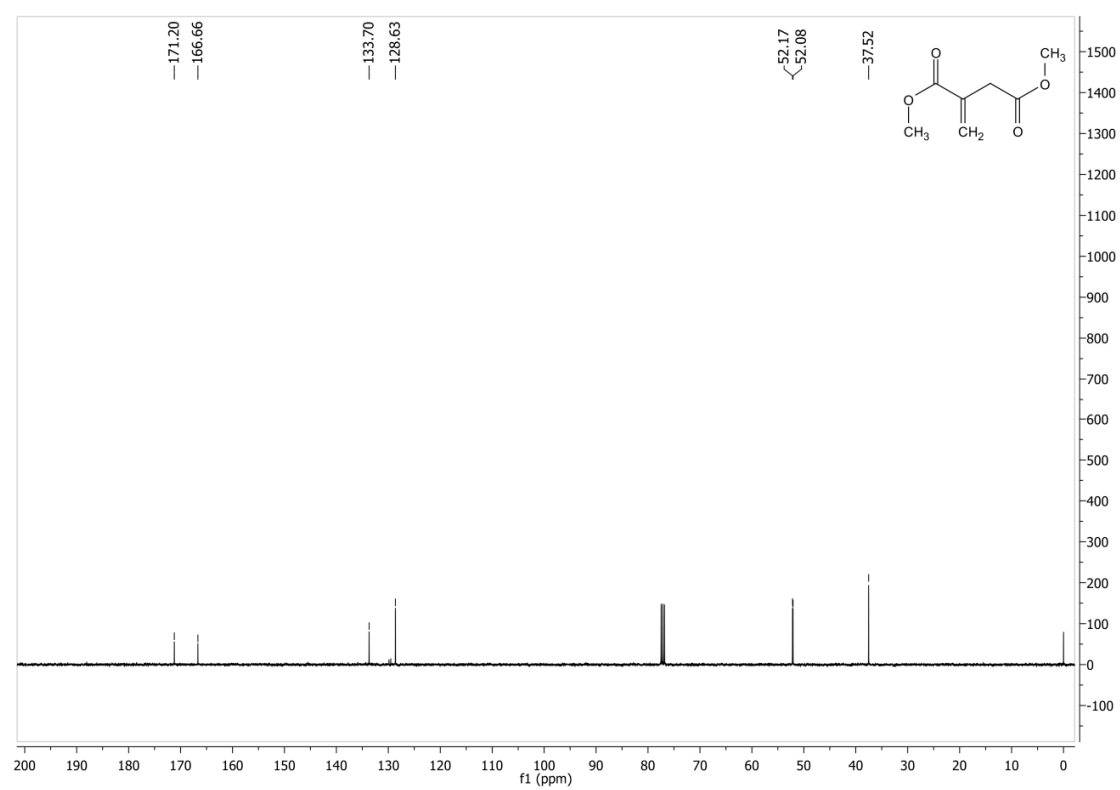
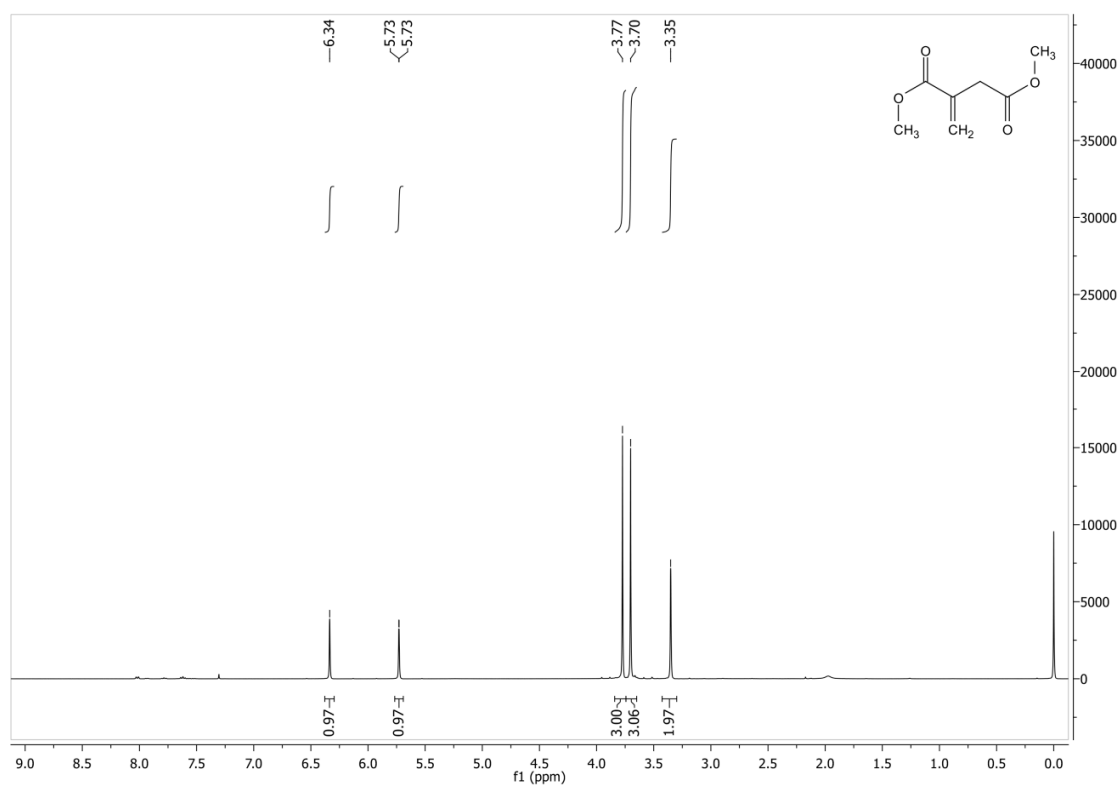
Dimethyl succinate – 21a



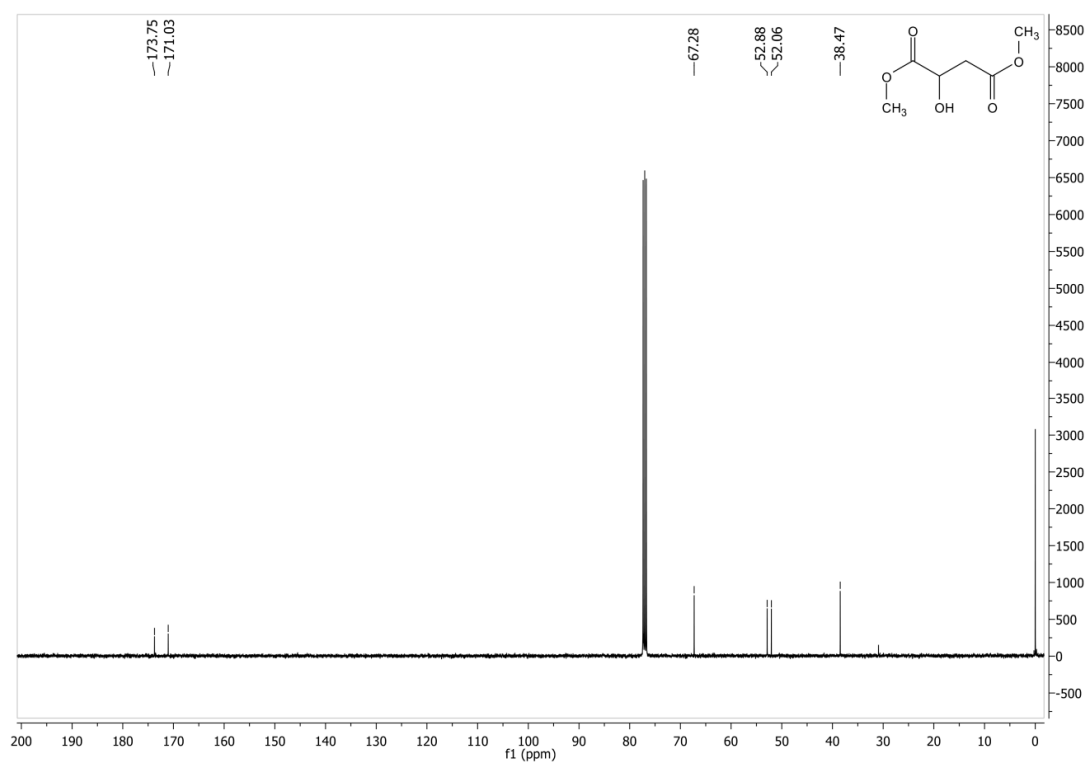
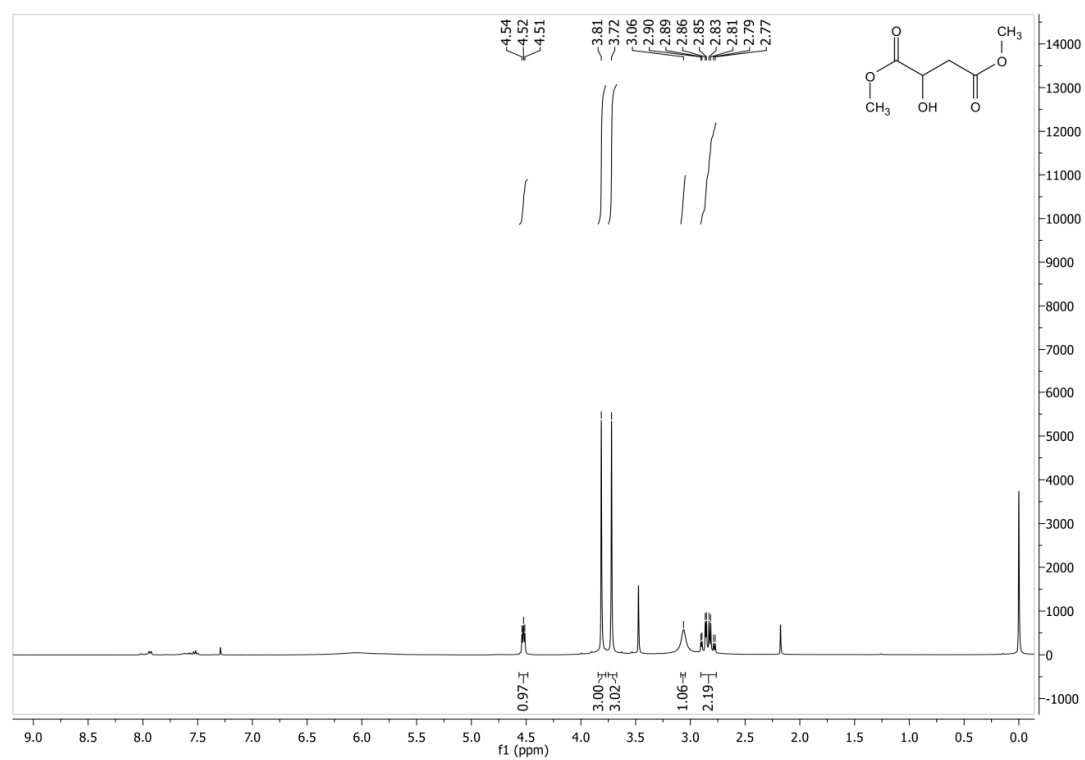
Dimethyl phenylsuccinate – 22a



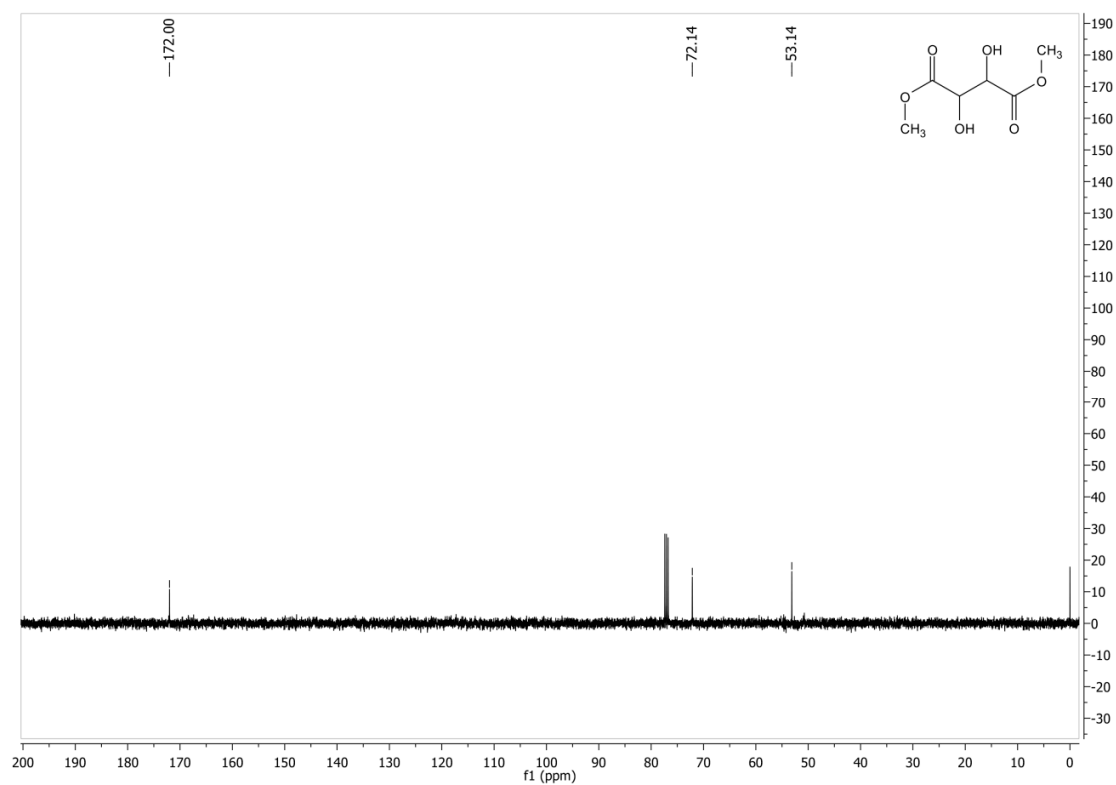
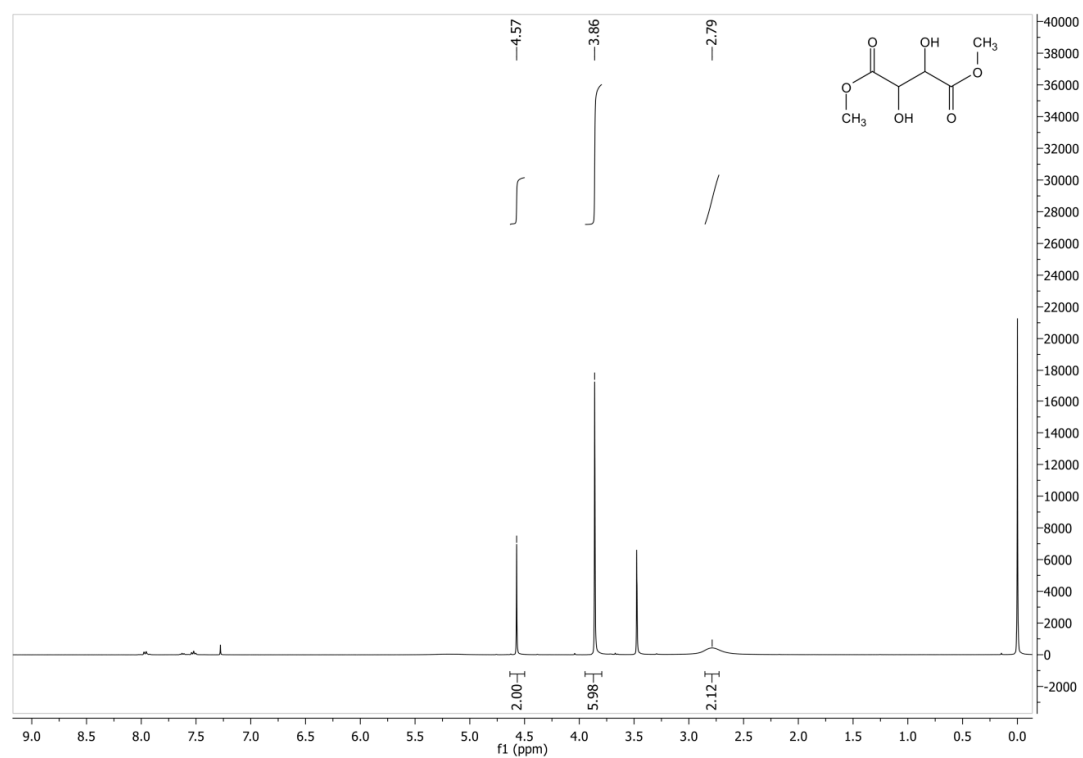
Dimethyl itaconate – 23a



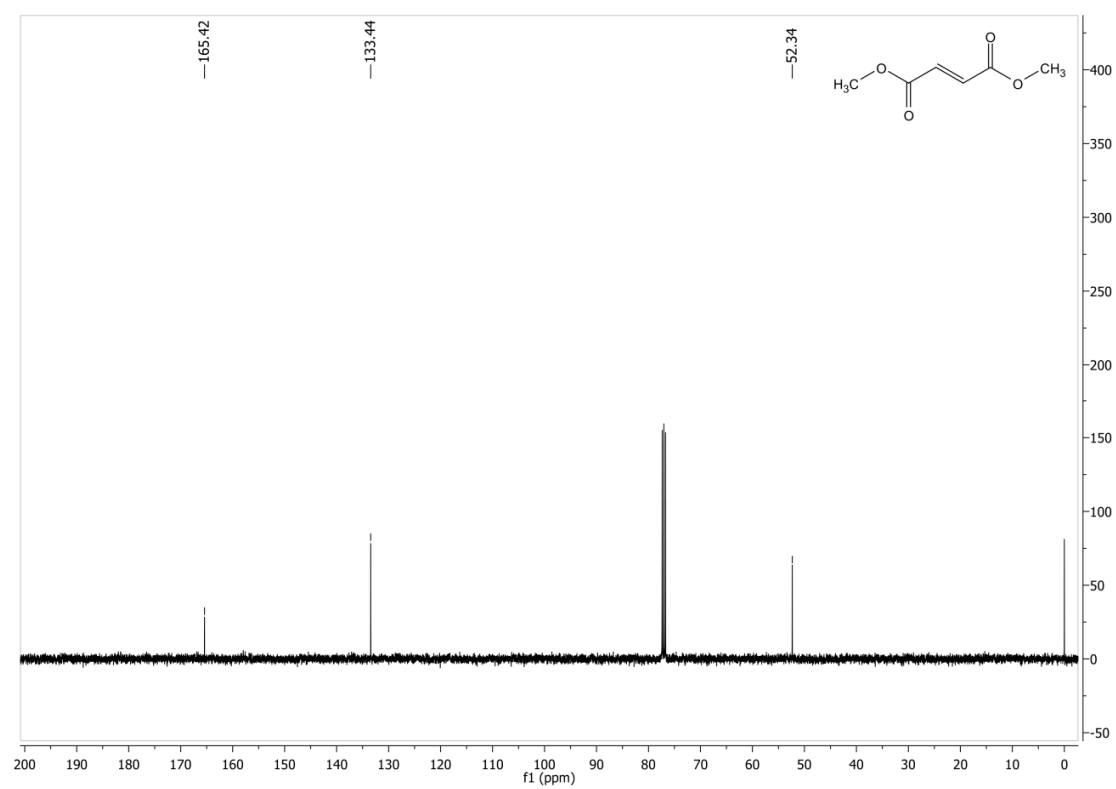
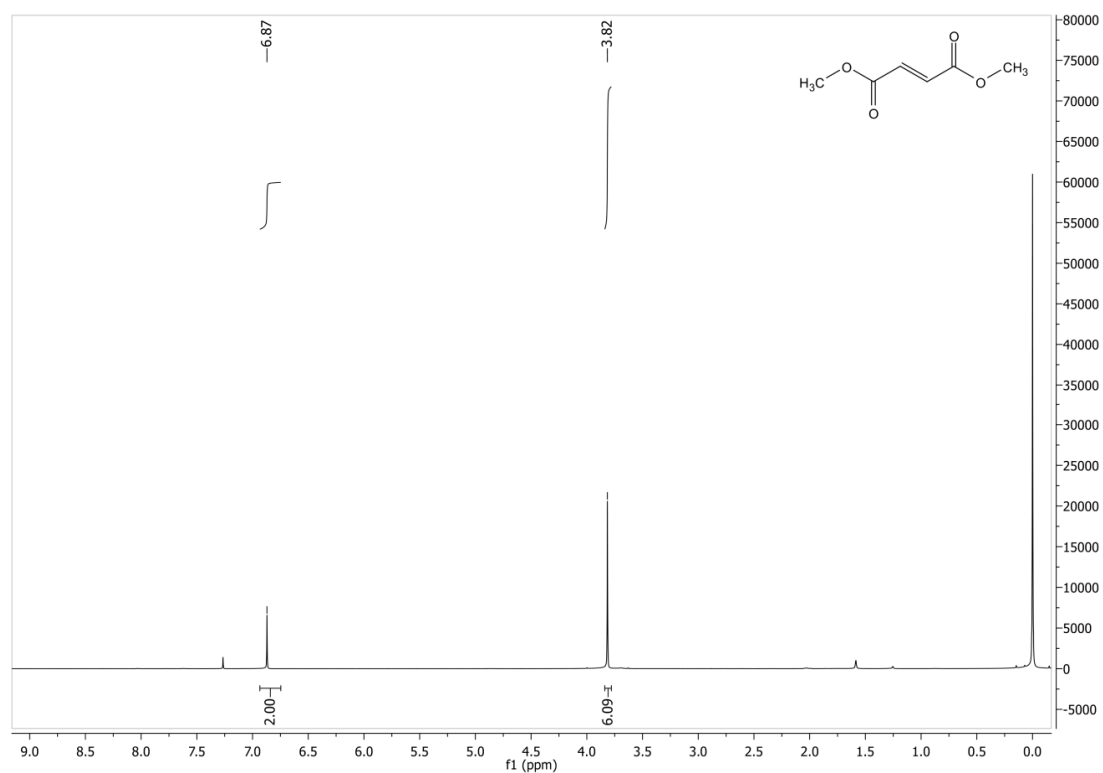
Dimethyl malate – 24a



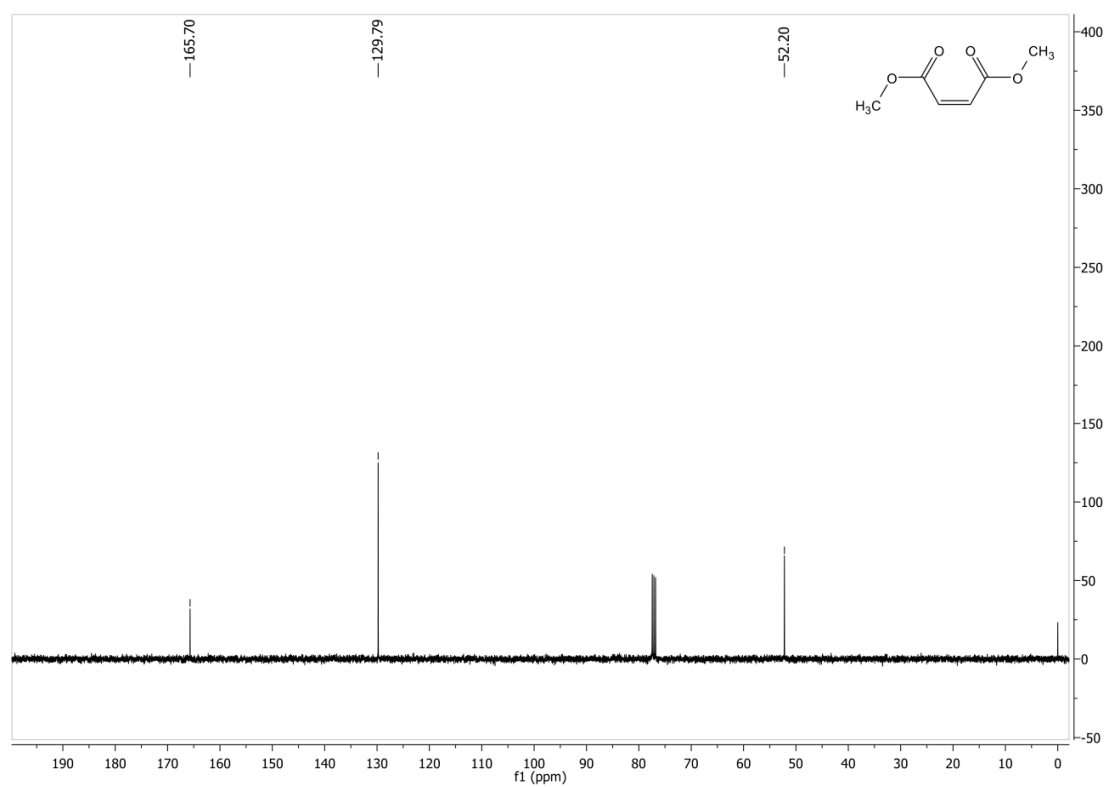
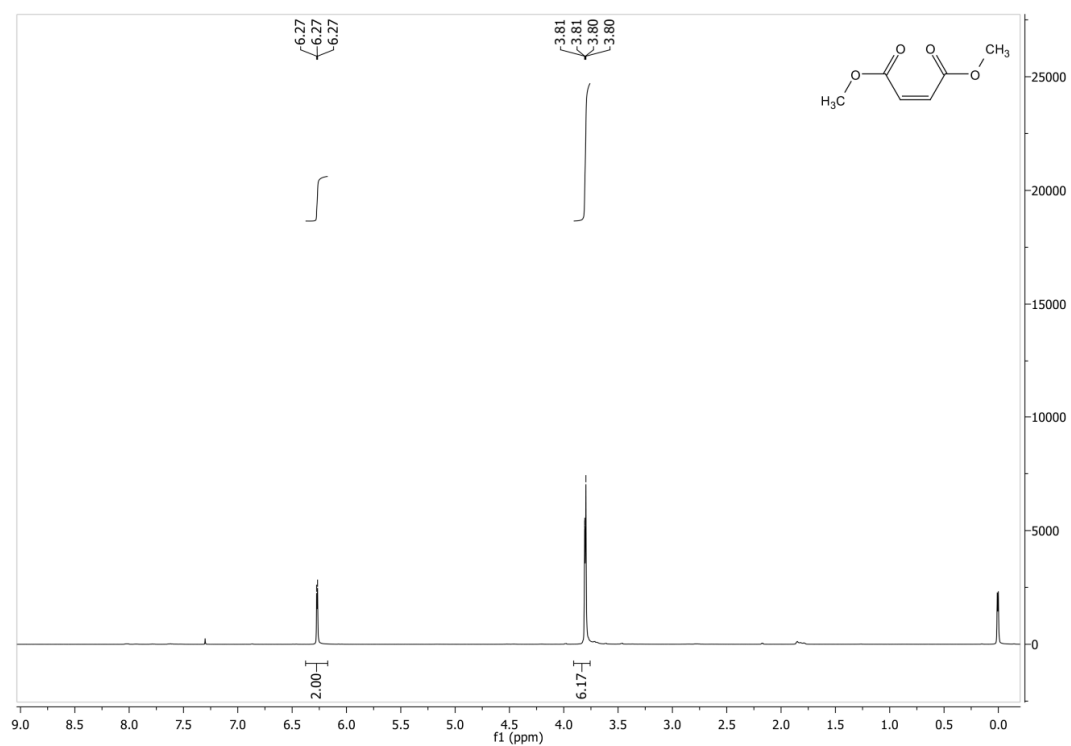
Dimethyl tartarate – 25a



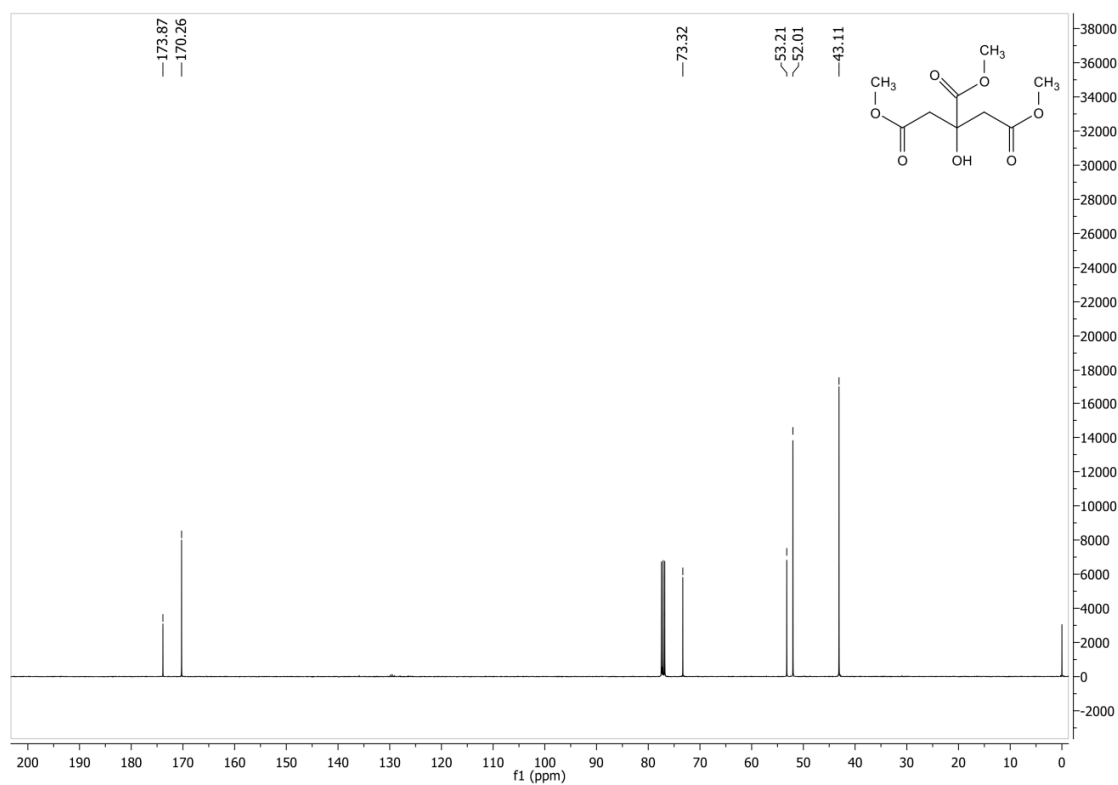
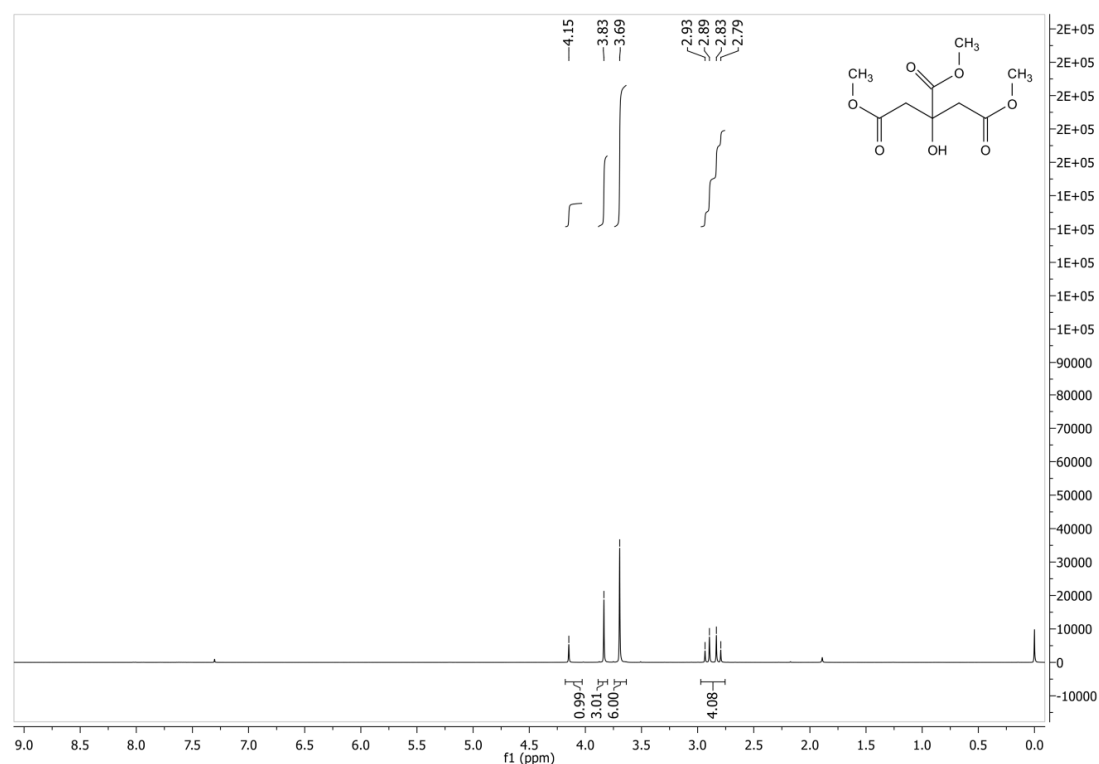
Dimethyl fumarate – 26a



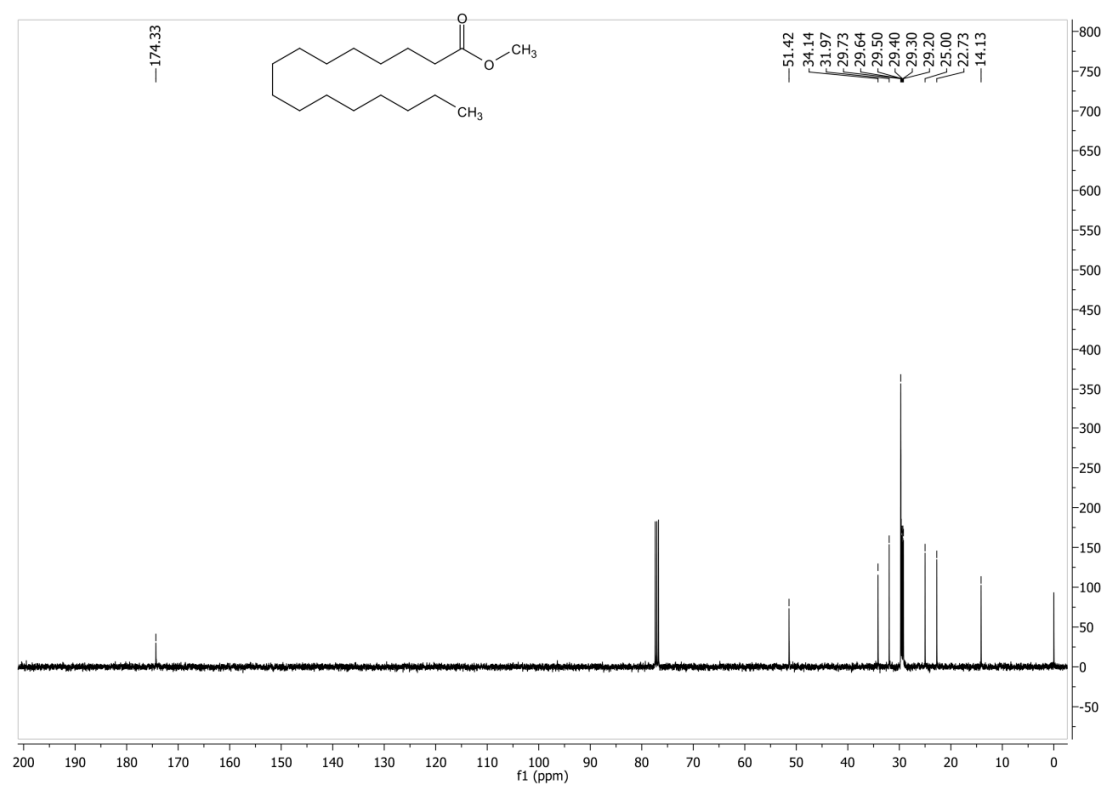
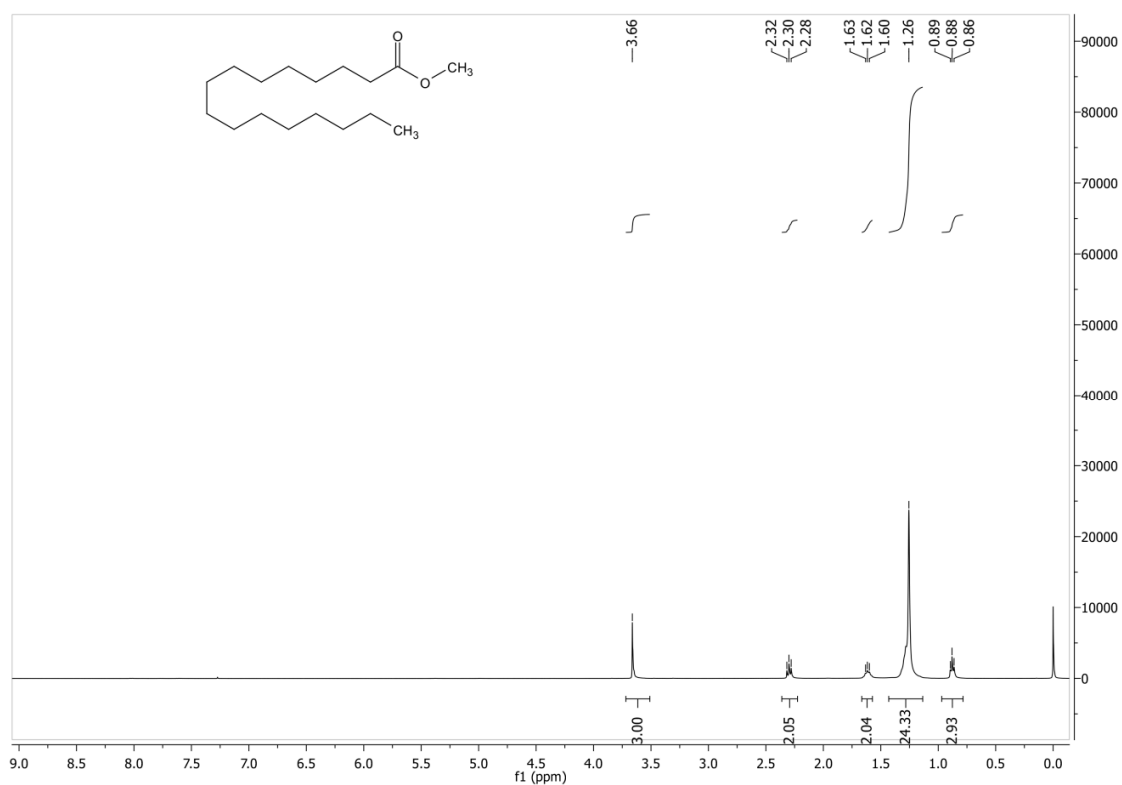
Dimethyl maleate – 27a



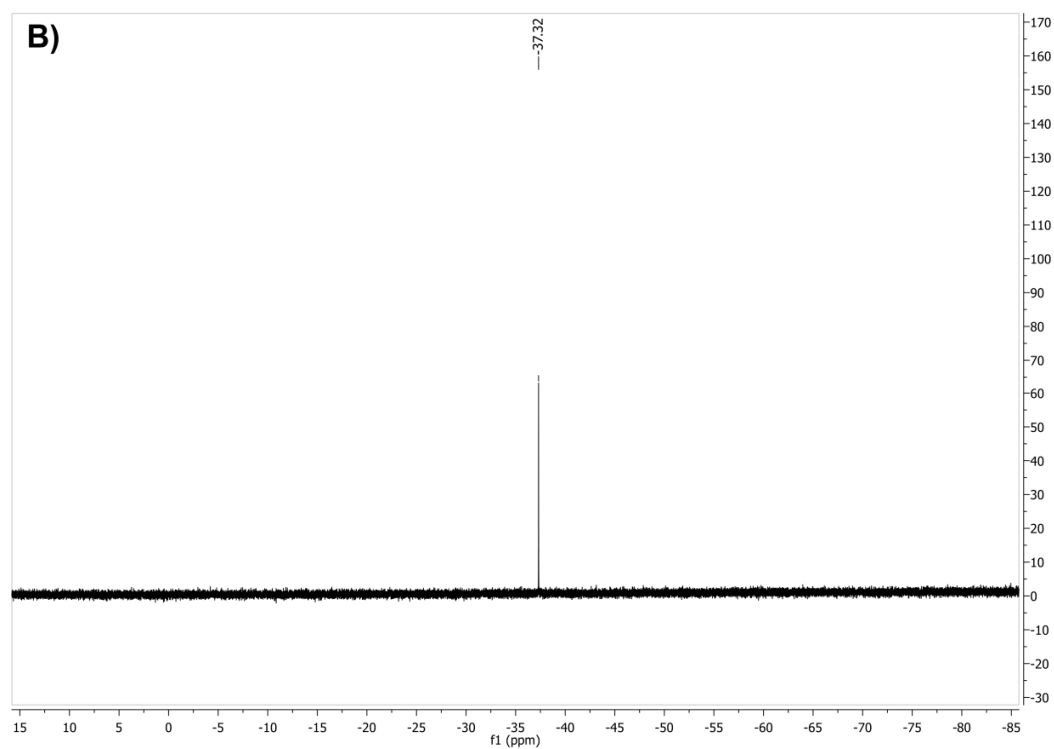
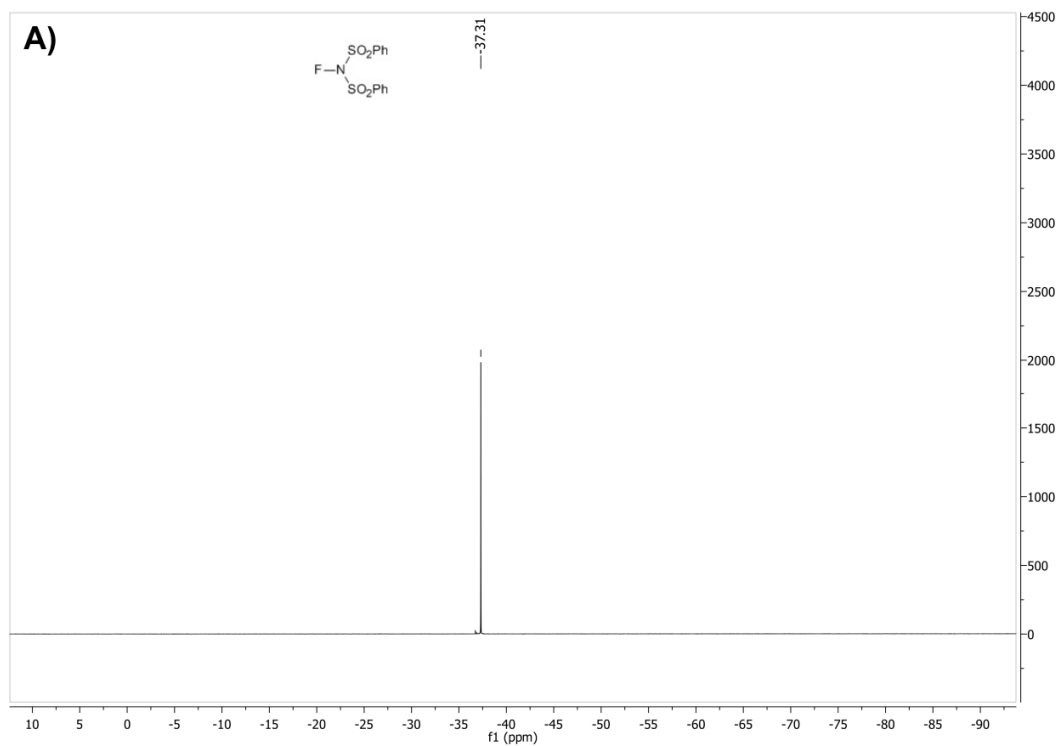
Trimethyl citrate – 28a



Methyl palmitate – 30a



List S3. ^{19}F NMR spectra of NFSi (**A**) and crude reaction mixture of cinnamic acid, MeOH and NFSi (**B**).



References

- Chen, Z.; Wen, Y.; Fu, Y.; Chen, H.; Ye, M.; Luo, G., Graphene oxide: An efficient acid catalyst for the construction of esters from acids and alcohols. *Synlett* **2017**, 28, 981–985.
- Whittaker, A.M.; Dong, V.M. Nickel-Catalyzed Dehydrogenative Cross-Coupling: Direct Transformation of Aldehydes into Esters and Amides. *Angewandte Chemie International Edition* **2015**, 54, 1312–1315.
- Minakawa, M.; Baek, H.; Yamada, Y.M.; Han, J.W.; Uozumi, Y. Direct dehydrative esterification of alcohols and carboxylic acids with a macroporous polymeric acid catalyst. *Organic letters* **2013**, 15, 5798–5801.
- Wang, Y.; Huang, Z.; Leng, X.; Zhu, H.; Liu, G.; Huang, Z. Transfer hydrogenation of alkenes using ethanol catalyzed by a NCP pincer iridium complex: scope and mechanism. *Journal of the American Chemical Society* **2018**, 140, 4417–4429.
- Hosseini-Sarvari, M.; Sodagar, E. Esterification of free fatty acids (Biodiesel) using nano sulfated-titania as catalyst in solvent-free conditions. *Comptes rendus chimie* **2013**, 16, 229–238.
- Sanna, V.; Mariani, A.; Caria, G.; Sechi, M. Synthesis and evaluation of different fatty acid esters formulated into Precirol® ATO-based lipid nanoparticles as vehicles for topical delivery. *Chemical and Pharmaceutical Bulletin* **2009**, 57, 680–684.
- Yang, H.; Li, Y.; Jiang, M.; Wang, J.; Fu, H. General copper-catalyzed transformations of functional groups from arylboronic acids in water. *Chemistry—A European Journal* **2011**, 17, 5652–5660.
- Bastos, E.L.; Ciscato, L.F.M.L.; Weiss, D.; Beckert, R.; Baader, W.J. Comparison of convenient alternative synthetic approaches to 4-[(3-tert-butylidimethylsilyloxy) phenyl]-4-methoxyspiro [1, 2-dioxetane-3, 2'-adamantane]. *Synthesis* **2006**, 2006, 1781–1786.
- Cheung, C.W.; Buchwald, S.L. Mild and general palladium-catalyzed synthesis of methyl aryl ethers enabled by the use of a palladacycle precatalyst. *Organic letters* **2013**, 15, 3998–4001.
- Jia, J.; Jiang, Q.; Zhao, A.; Xu, B.; Liu, Q.; Luo, W.-P.; Guo, C.-C. Copper-catalyzed O-methylation of carboxylic acids using DMSO as a methyl source. *Synthesis* **2016**, 48, 421–428.
- McNulty, J.; Capretta, A.; Laritchev, V.; Dyck, J.; Robertson, A.J. Dimethylmalonyltrialkylphosphoranes: New general reagents for esterification reactions allowing controlled inversion or retention of configuration on chiral alcohols. *The Journal of Organic Chemistry* **2003**, 68, 1597–1600.
- Yamamoto, Y. The First General and Selective Palladium (II)-Catalyzed Alkoxyacylation of Arylboronates: Interplay among Benzoquinone-Ligated Palladium (0) Complex, Organoboron, and Alcohol Solvent. *Advanced Synthesis & Catalysis* **2010**, 352, 478–492.
- Kiran, Y.; Ikeda, R.; Sakai, N.; Konakahara, T. Single-step conversion of electron-deficient aldehydes into the corresponding esters in aqueous alcohols in the presence of iodine and sodium nitrite. *Synthesis* **2010**, 2010, 276–282.
- Talzi, V. A ¹³C and ¹H NMR analysis of perfumes. *Russian journal of applied chemistry* **2006**, 79, 107–116.
- Revelant, G.; Dunand, S.; Hesse, S.; Kirsch, G. Microwave-assisted synthesis of 5-substituted 2-aminothiophenes starting from arylacetaldehydes. *Synthesis* **2011**, 2011, 2935–2940.
- Yamamoto, N.; Obora, Y.; Ishii, Y. Iridium-catalyzed oxidative methyl esterification of primary alcohols and diols with methanol. *The Journal of organic chemistry* **2011**, 76, 2937–2941.
- Aridoss, G.; Laali, K.K. Ethylammonium nitrate (EAN)/Tf₂O and EAN/TFAA: Ionic liquid based systems for aromatic nitration. *The Journal of Organic Chemistry* **2011**, 76, 8088–8094.
- Dawar, P.; Raju, M.B.; Ramakrishna, R.A. One-pot esterification and Ritter reaction: chemo-and regioselectivity from tert-butyl methyl ether. *Tetrahedron letters* **2011**, 52, 4262–4265.
- Omar, S.; Abu-Reziq, R. Palladium nanoparticles supported on magnetic organic-silica hybrid nanoparticles. *The Journal of Physical Chemistry C* **2014**, 118, 30045–30056.
- Bonnamour, J.; Bolm, C. Iron (II) triflate as a catalyst for the synthesis of indoles by intramolecular C–H amination. *Organic letters* **2011**, 13, 2012–2014.
- Shenvi, R.A.; O'Malley, D.P.; Baran, P.S. Chemoselectivity: the mother of invention in total synthesis. *Accounts of chemical research* **2009**, 42, 530–541.
- Gao, Y.X.; Chang, L.; Shi, H.; Liang, B.; Wongkhan, K.; Chaiyaveij, D.; Batsanov, A.S.; Marder, T.B.; Li, C.C.; Yang, Z. A Thiourea-Oxazoline Library with Axial Chirality: Ligand Synthesis and Studies of the Palladium-Catalyzed Enantioselective Bis (methoxycarbonylation) of Terminal Olefins. *Advanced Synthesis & Catalysis* **2010**, 352, 1955–1966.
- Hall, M.; Stueckler, C.; Hauer, B.; Stuermer, R.; Friedrich, T.; Breuer, M.; Kroutil, W.; Faber, K. Asymmetric bioreduction of activated C=C bonds using *Zymomonas mobilis* NCR enoate reductase and old yellow enzymes OYE 1–3 from yeasts. Wiley Online Library: Hoboken, NJ, USA, 2008.

24. Denmark, S.E.; Ahmad, M. Carbonylative ring opening of terminal epoxides at atmospheric pressure. *The Journal of Organic Chemistry* **2007**, *72*, 9630–9634.
25. Matteson, D.S.; Ray, R.; Rocks, R.R.; Tsai, D.J. Directed chiral synthesis by way of. α -chloro boronic esters. *Organometallics* **1983**, *2*, 1536–1543.
26. Sun, H.-B.; Hua, R.; Yin, Y. $\text{ZrOCl}_2 \cdot 8\text{H}_2\text{O}$: An efficient, cheap and reusable catalyst for the esterification of acrylic acid and other carboxylic acids with equimolar amounts of alcohols. *Molecules* **2006**, *11*, 263–271.