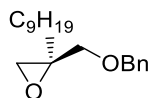
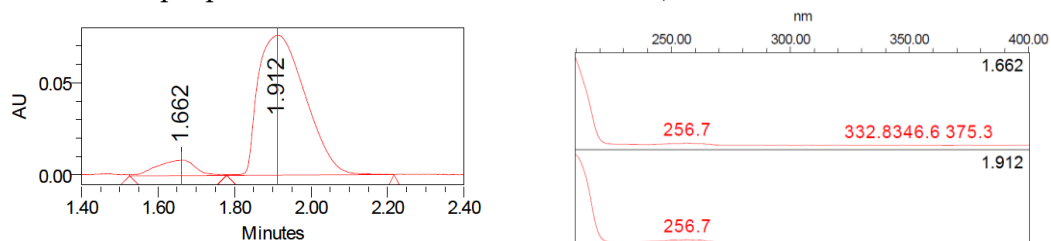


## Supplementary Materials

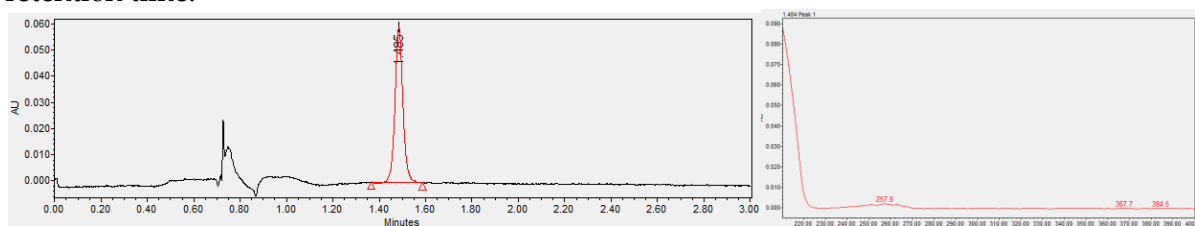


**14**

**Reference Sample:** The chromatogram of **14** was compared to a previously prepared sample of the (*S*)-enantiomer of the same compound. (Conditions: Waters Acquity UPC<sup>2</sup>, Chiracel IB, scCO<sub>2</sub>/isopropanol = 95:5, flow rate = 2 mL min<sup>-1</sup>).

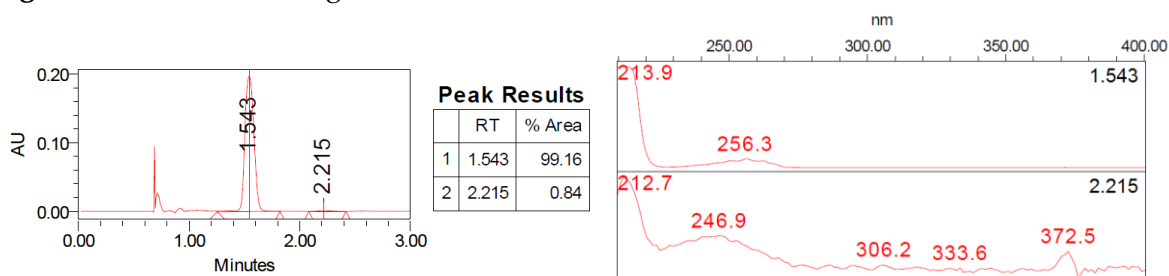


**Intermediate 14:** Only one peak was observed in the chromatogram of **14** shown below indicating an *ee* over 99 %. The product was assigned as the (*R*)-enantiomer due to the retention time.

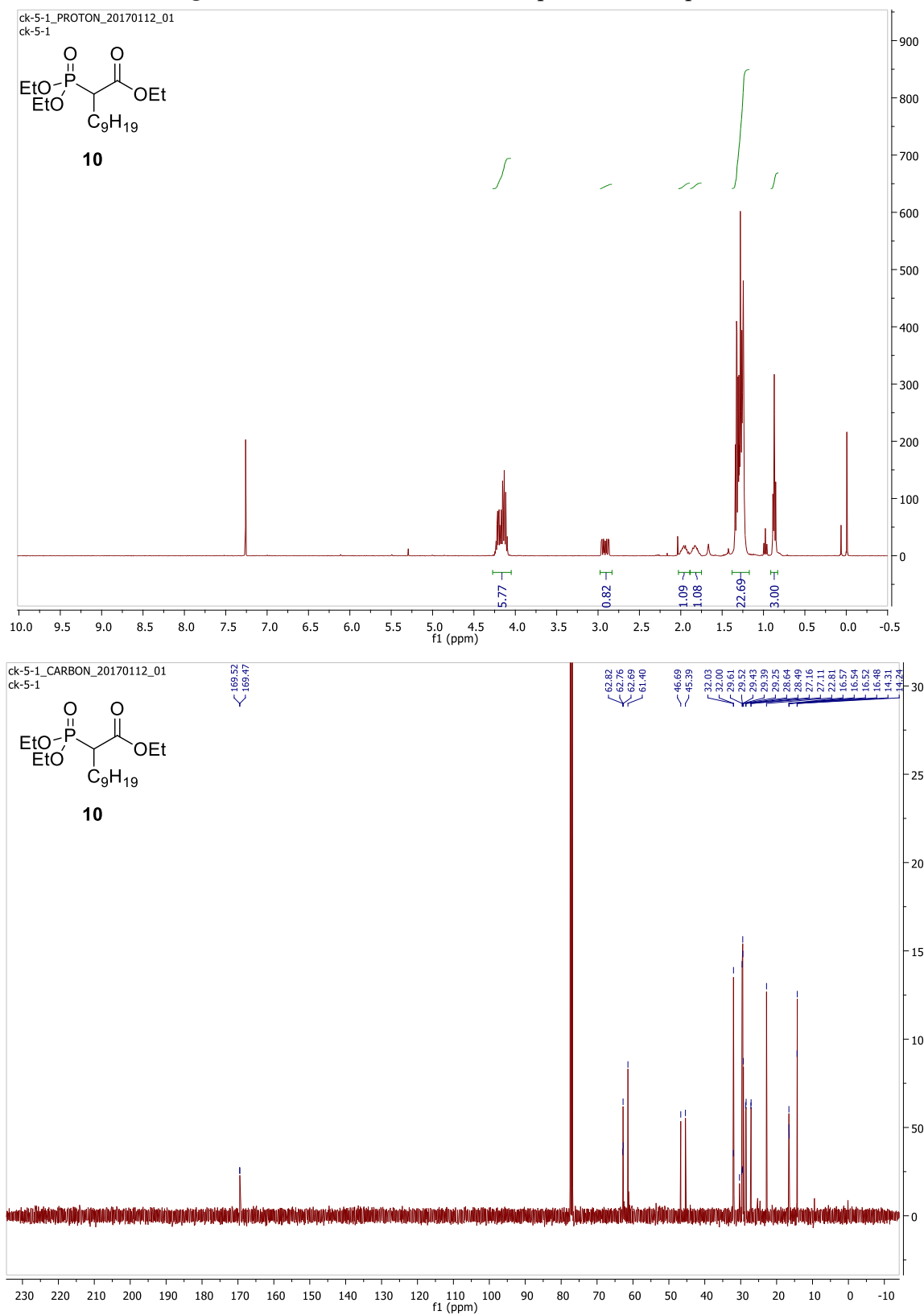


Comparison was also made to a chromatogram obtained from a previous synthesis of intermediate **14** in which a slightly lower *ee* was observed. In this case, the small peak for the (*S*)-enantiomer is visible.

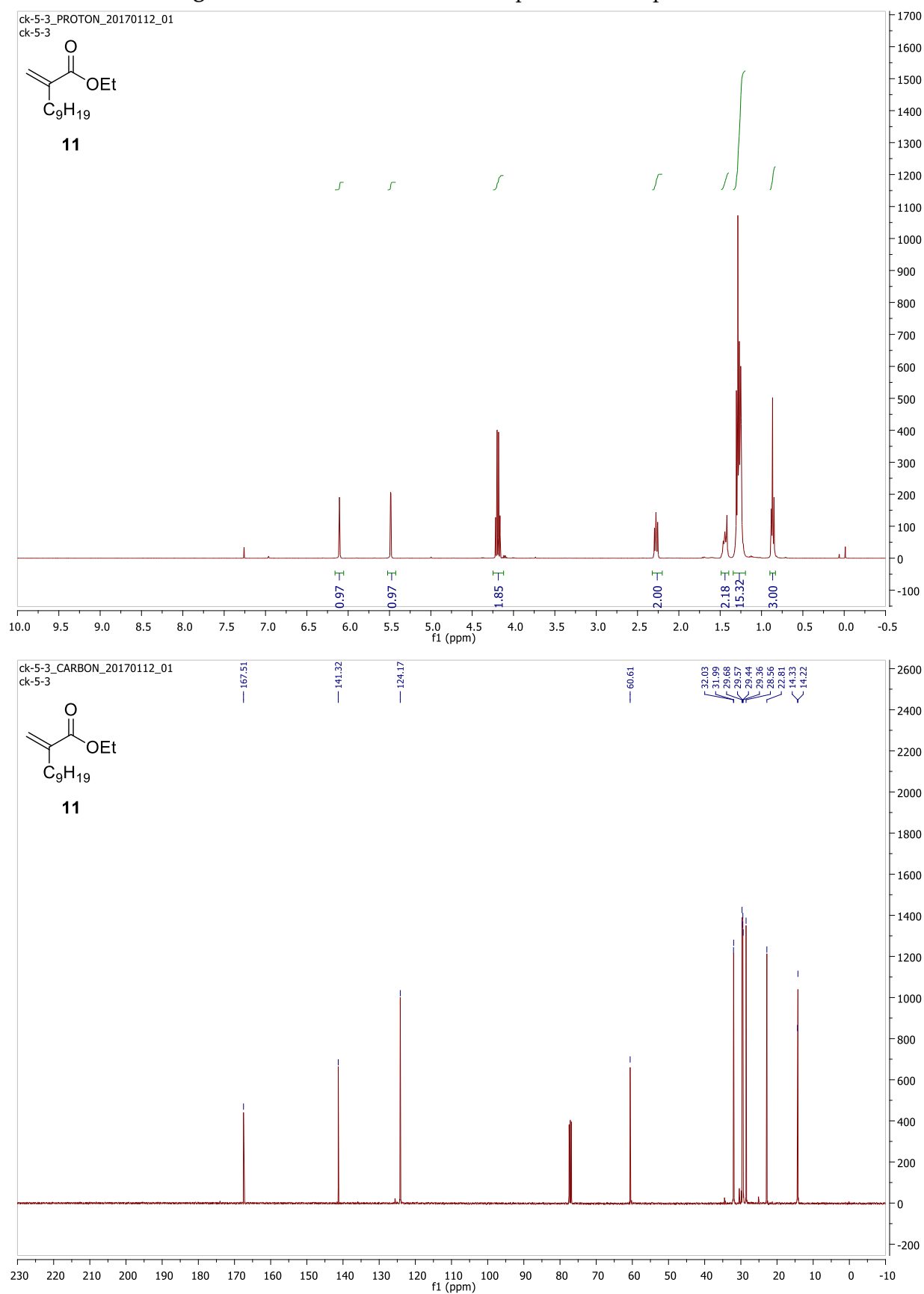
**Figure S1.** SFC Chromatograms of Reference and Intermediate **14**



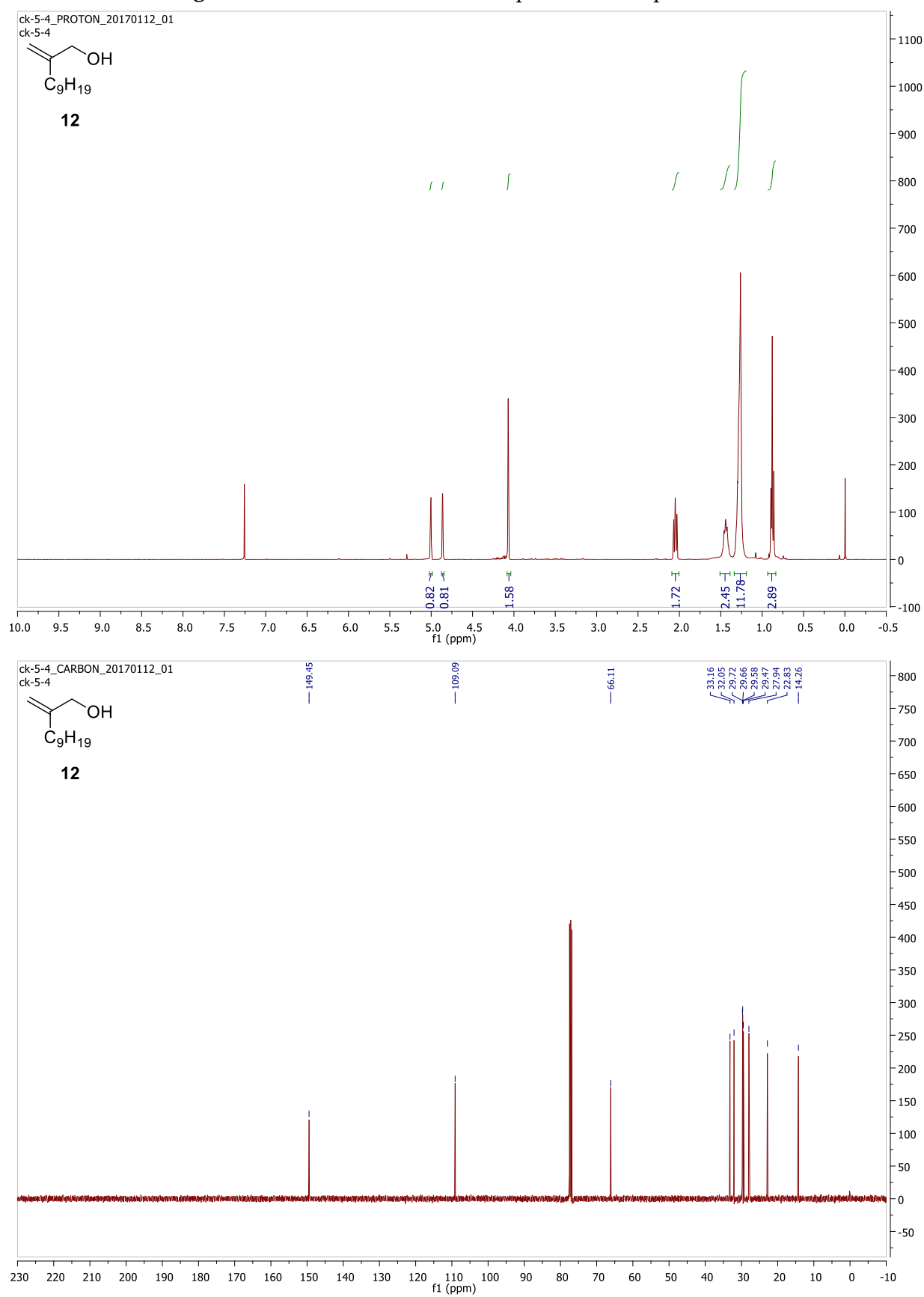
**Figure S2:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 10**



**Figure S3:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 11**



**Figure S4:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 12**



**Figure S5:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 13**

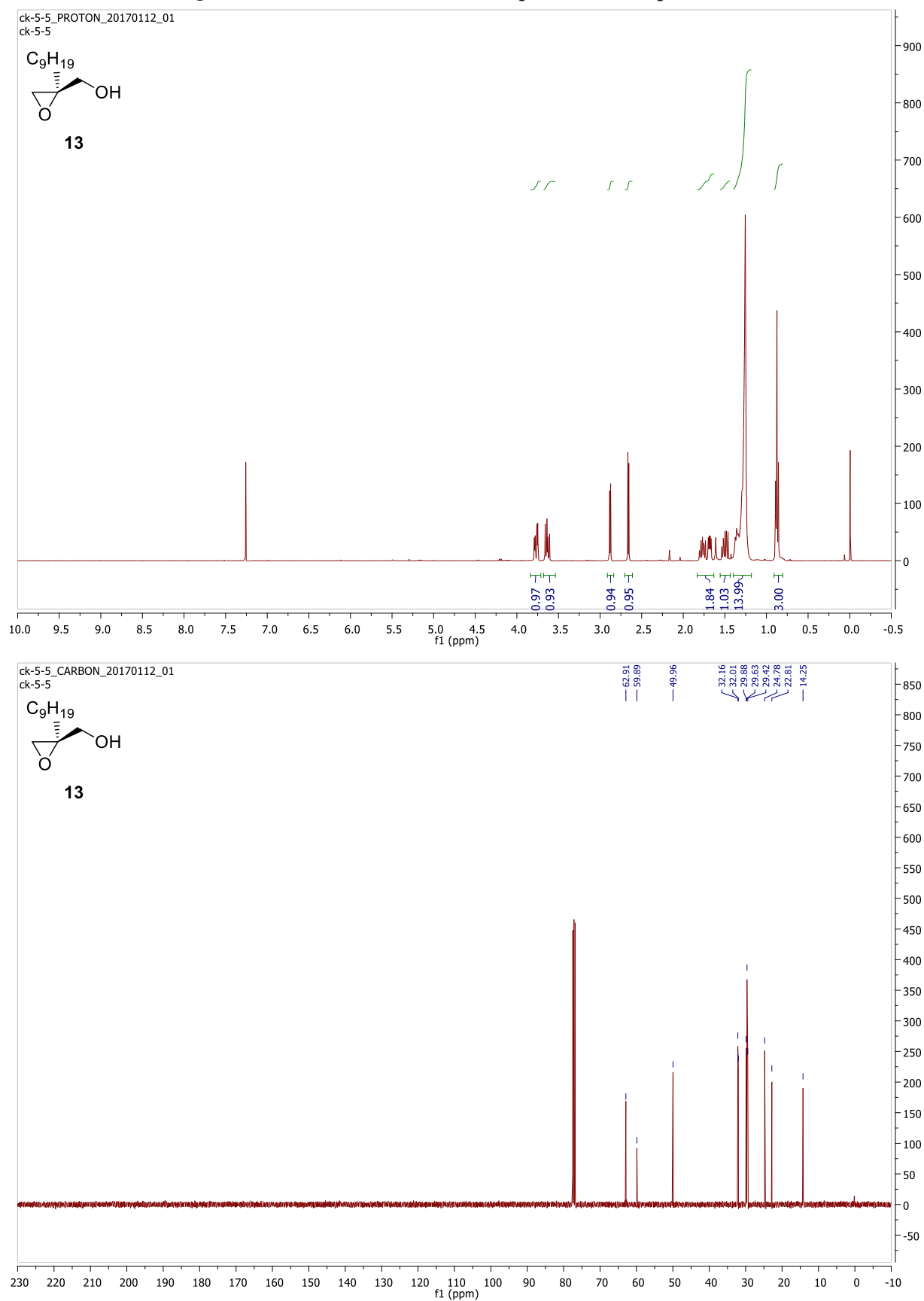
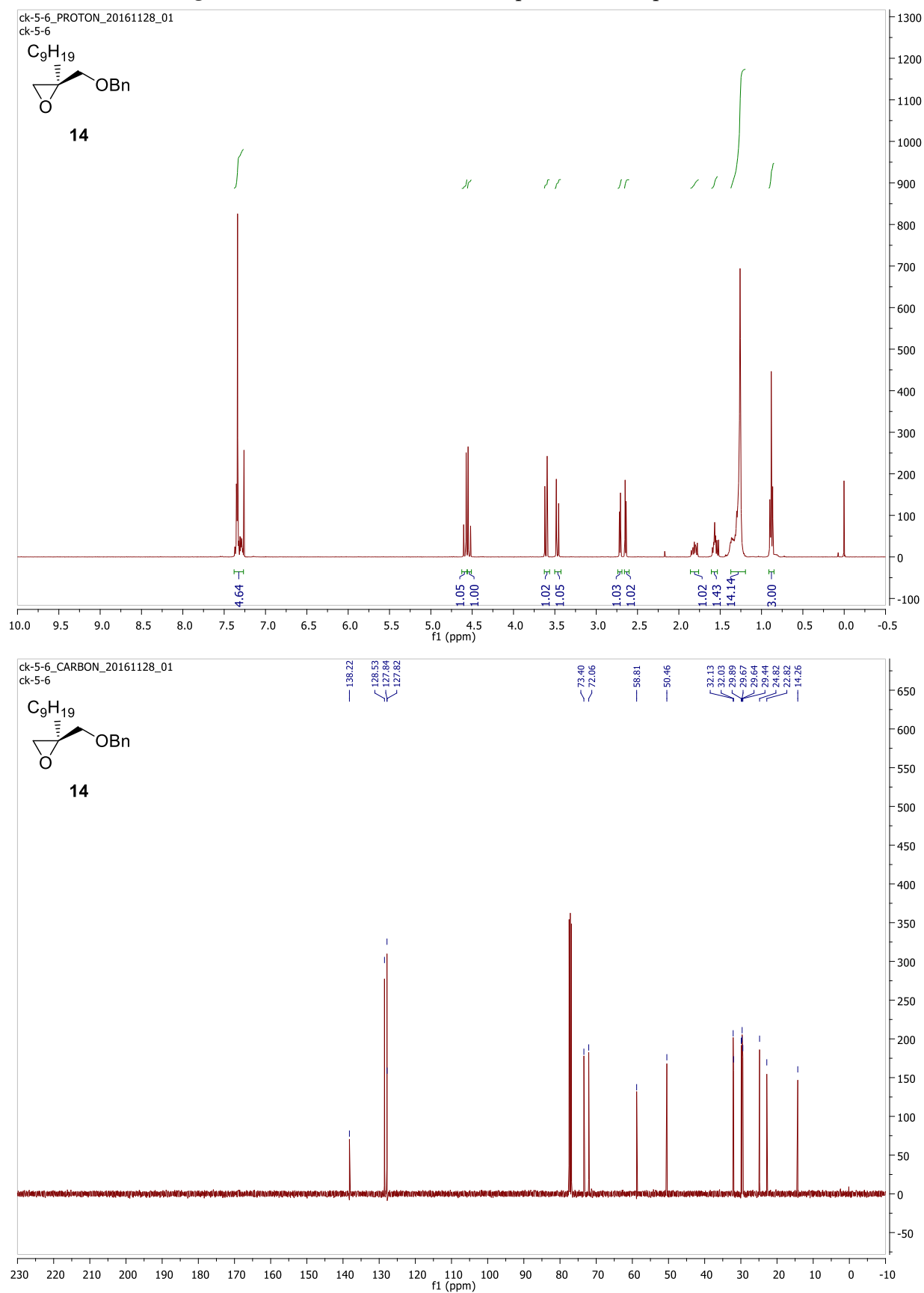


Figure S6:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 14



**Figure S7:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 15**

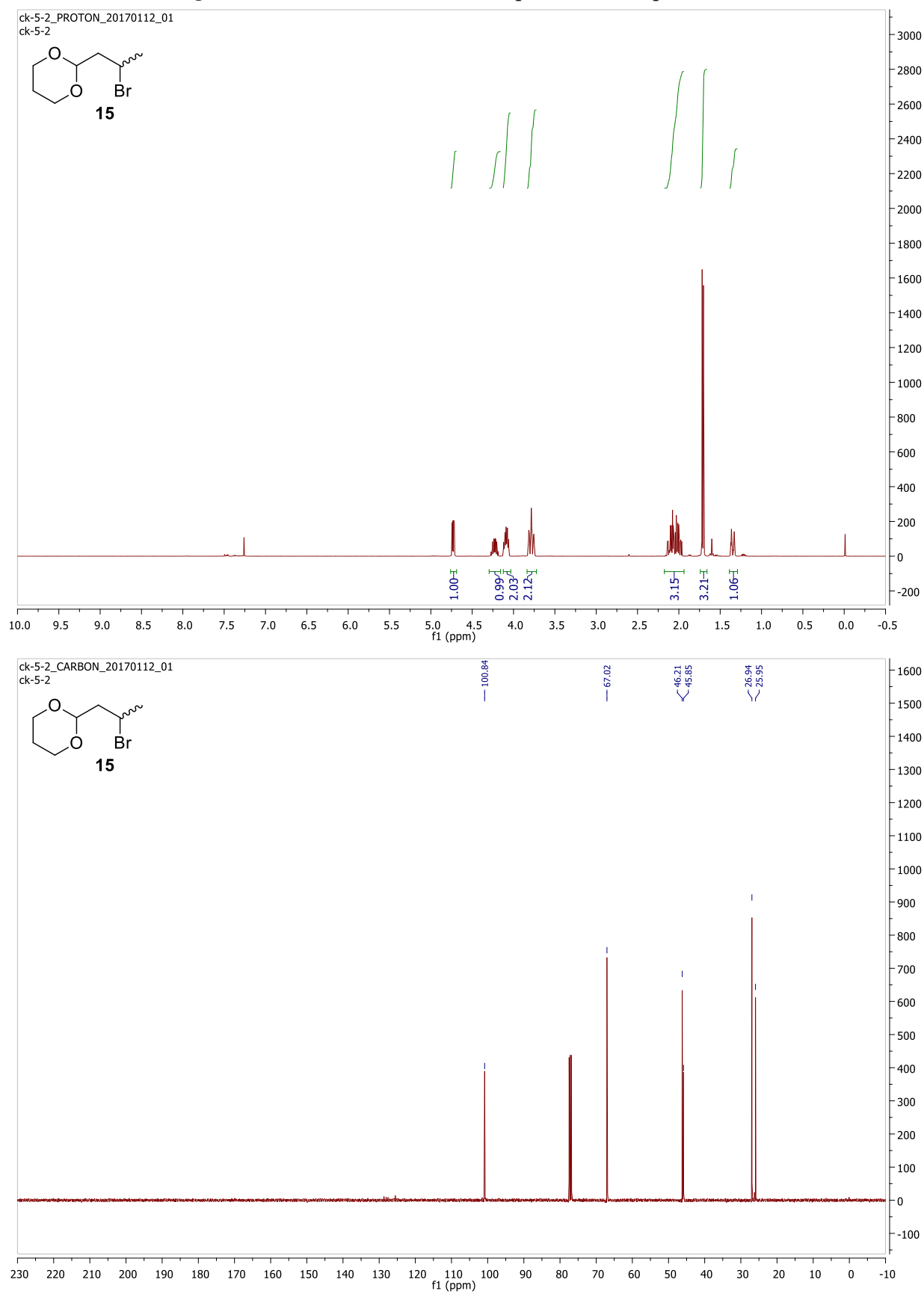
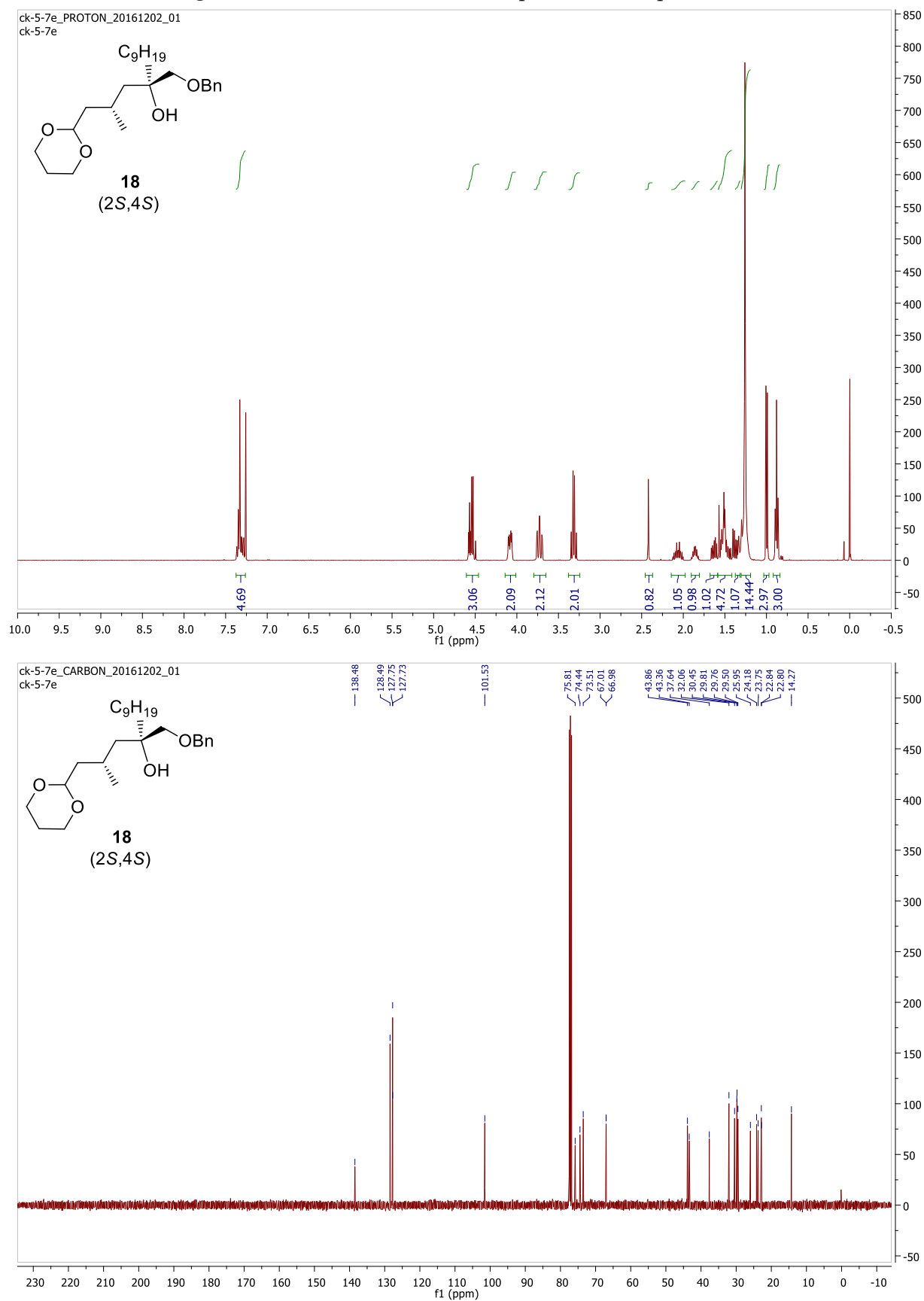


Figure S8:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 18





**Figure S9:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 22**

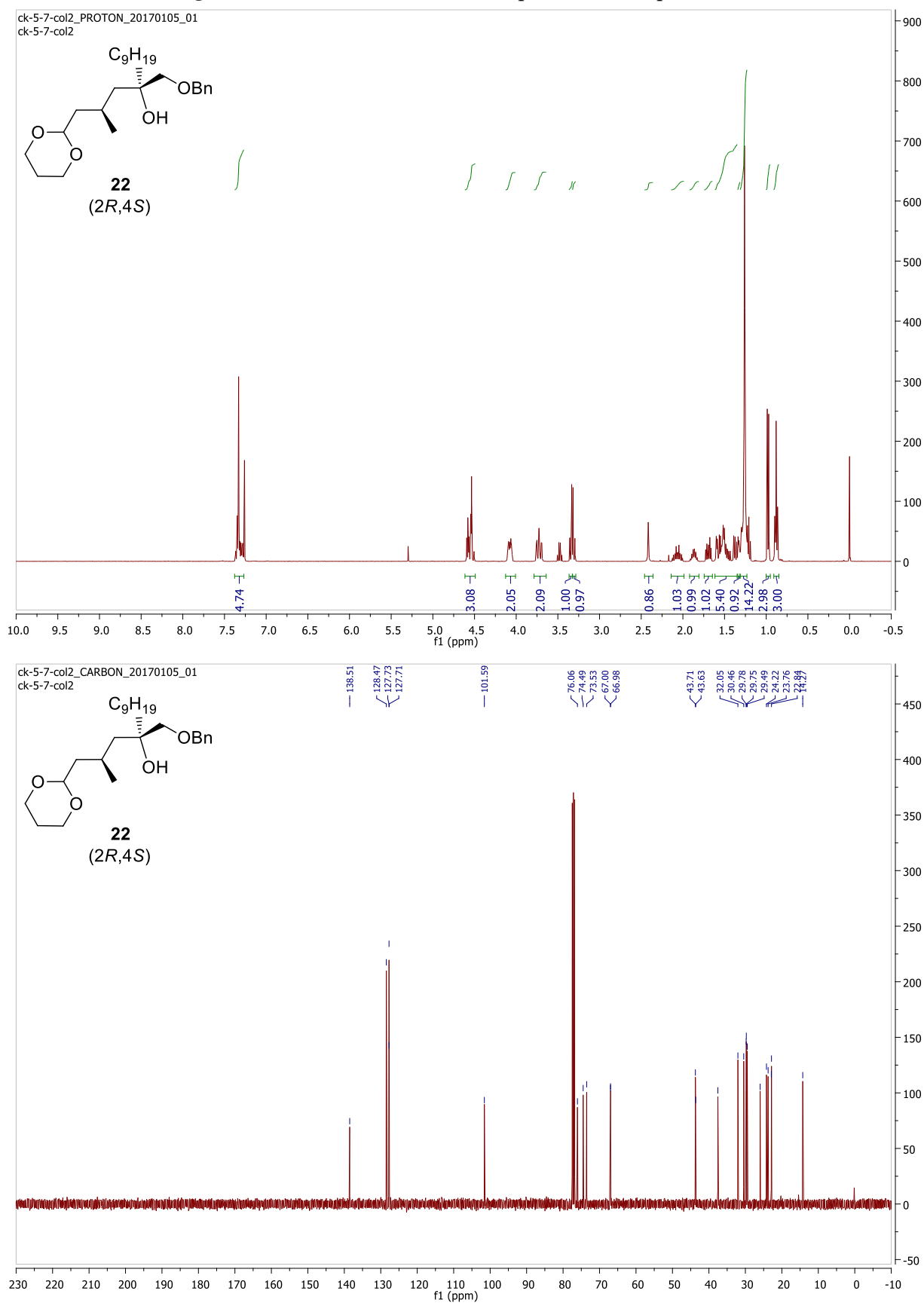
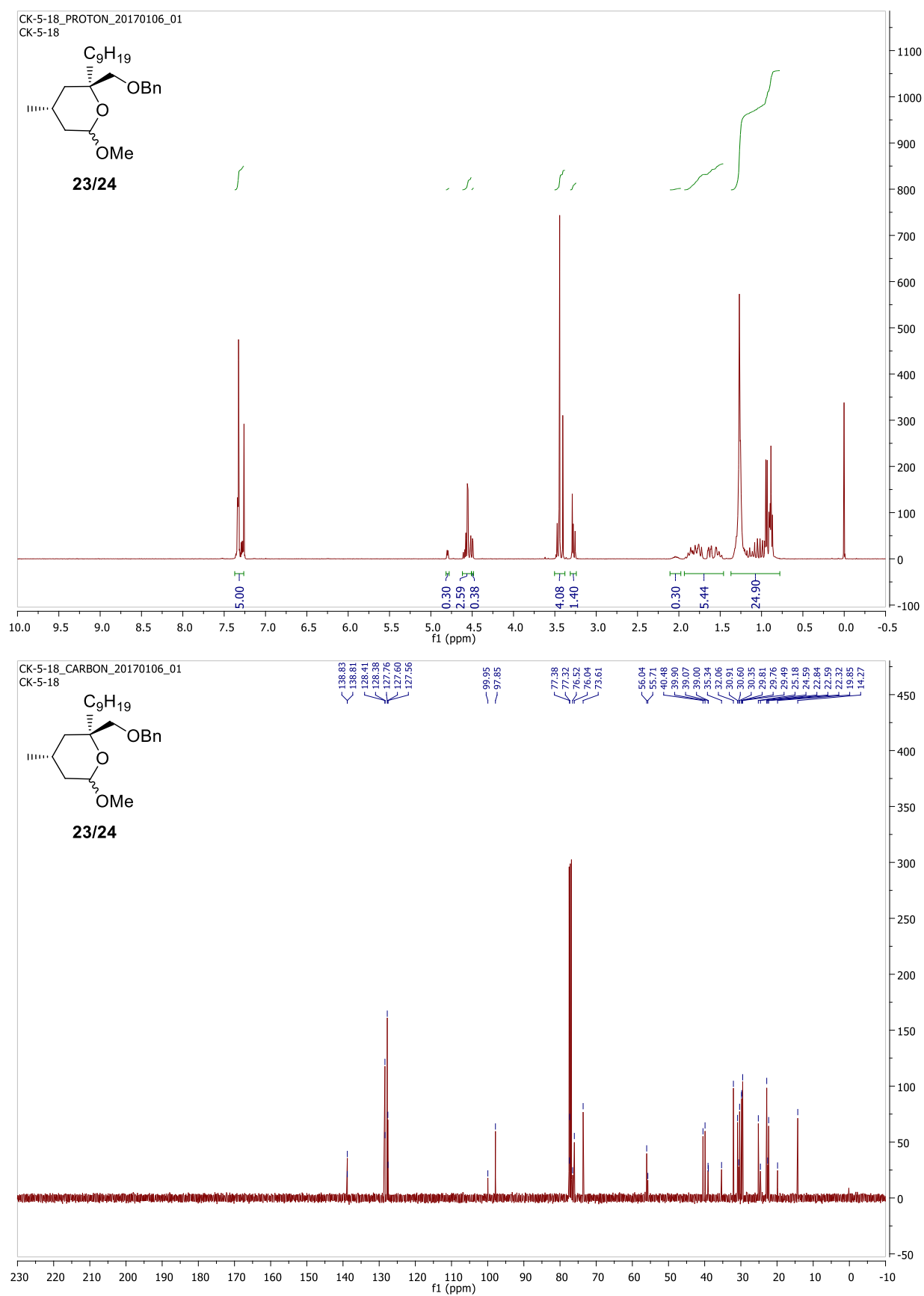


Figure S10:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compounds 23/24



**Figure S11:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compounds 19/20**

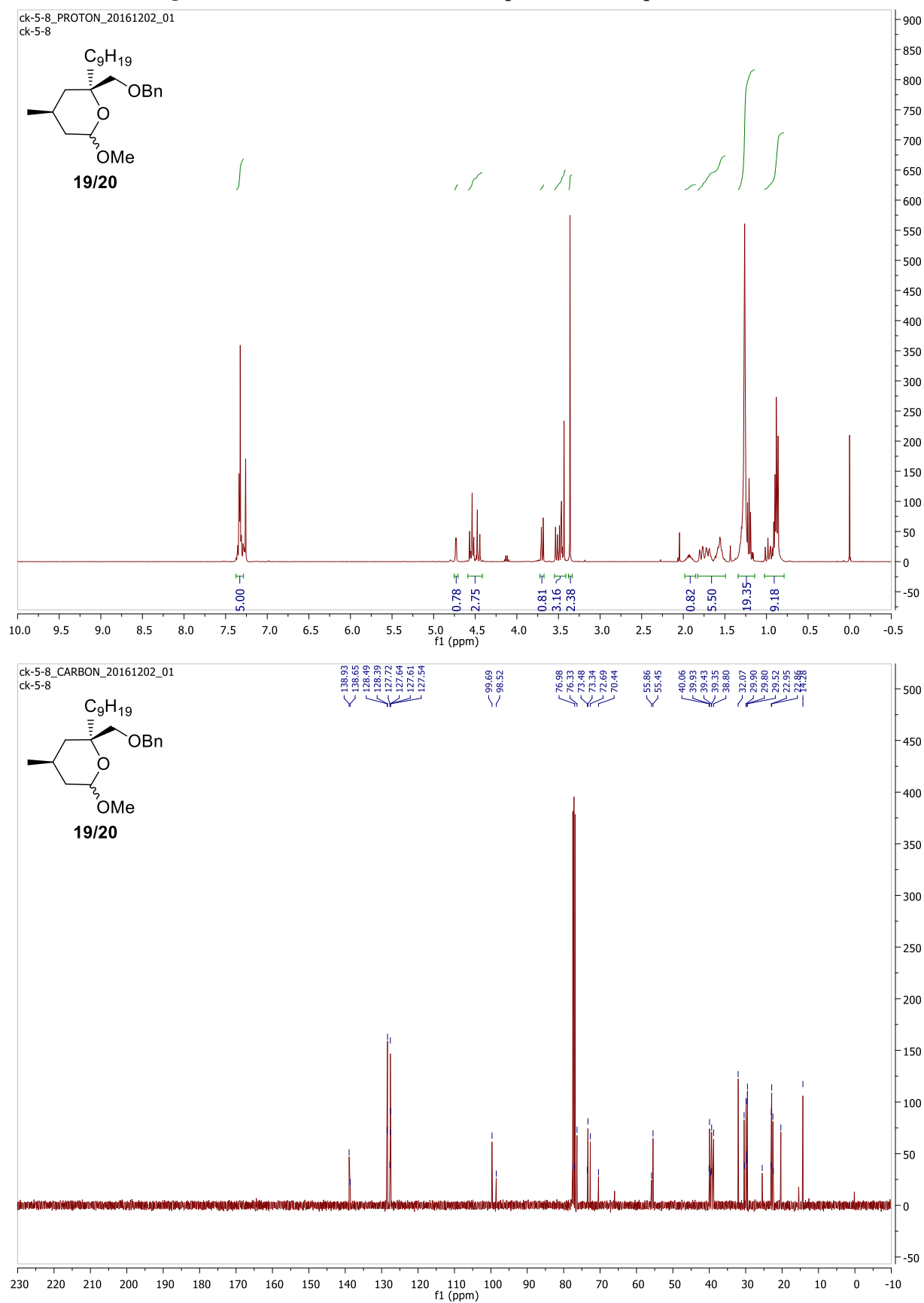


Figure S12:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 25

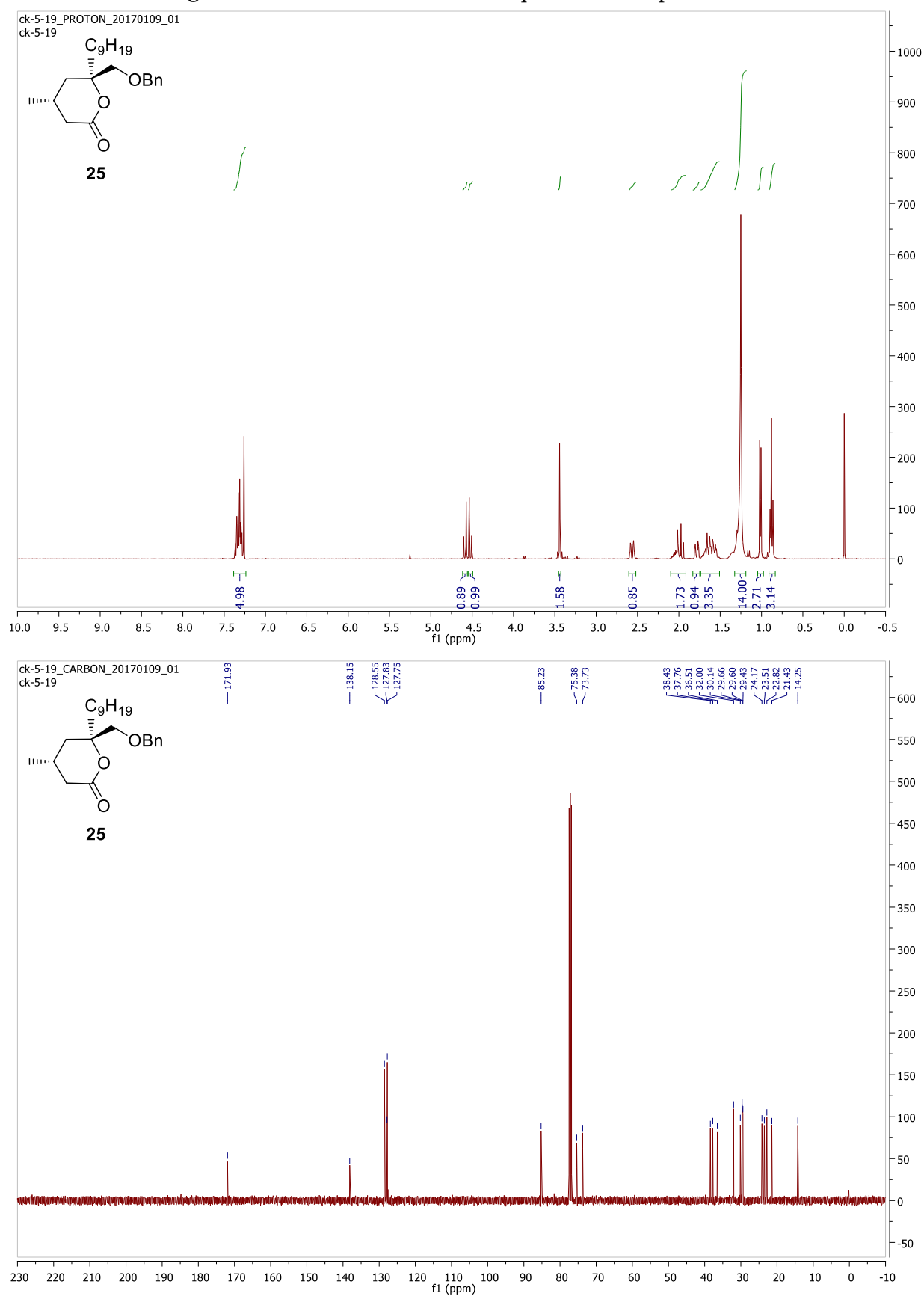
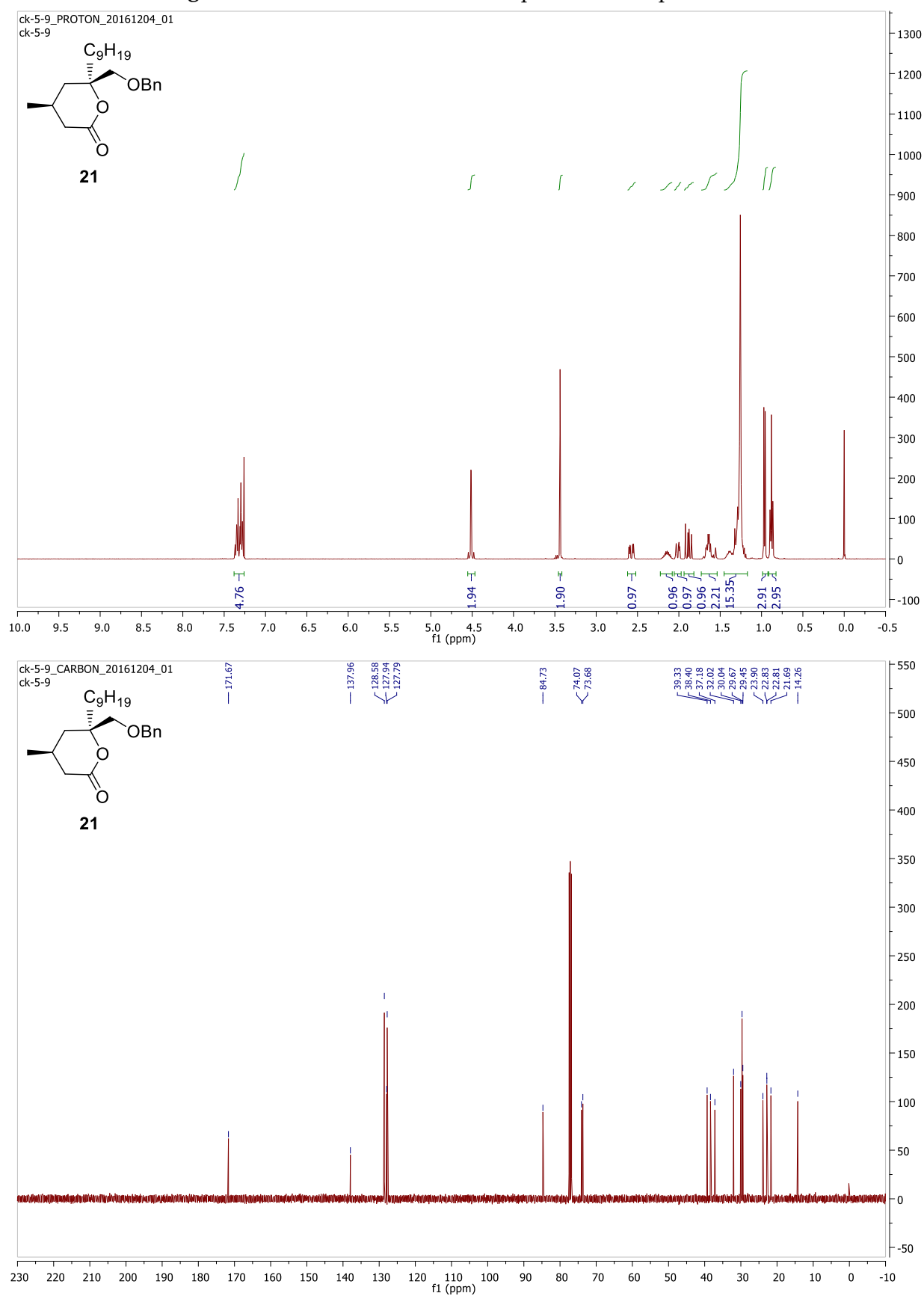
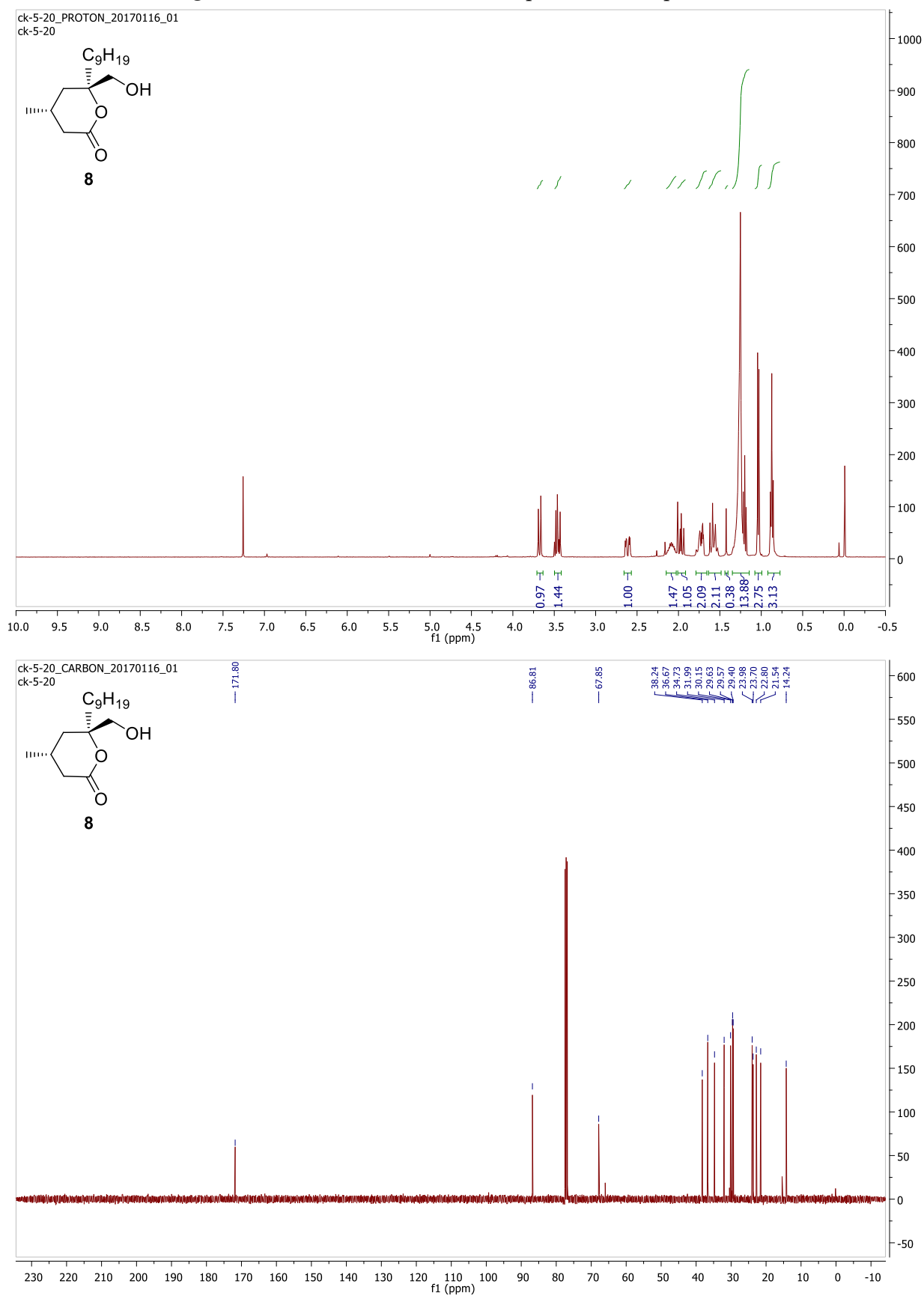


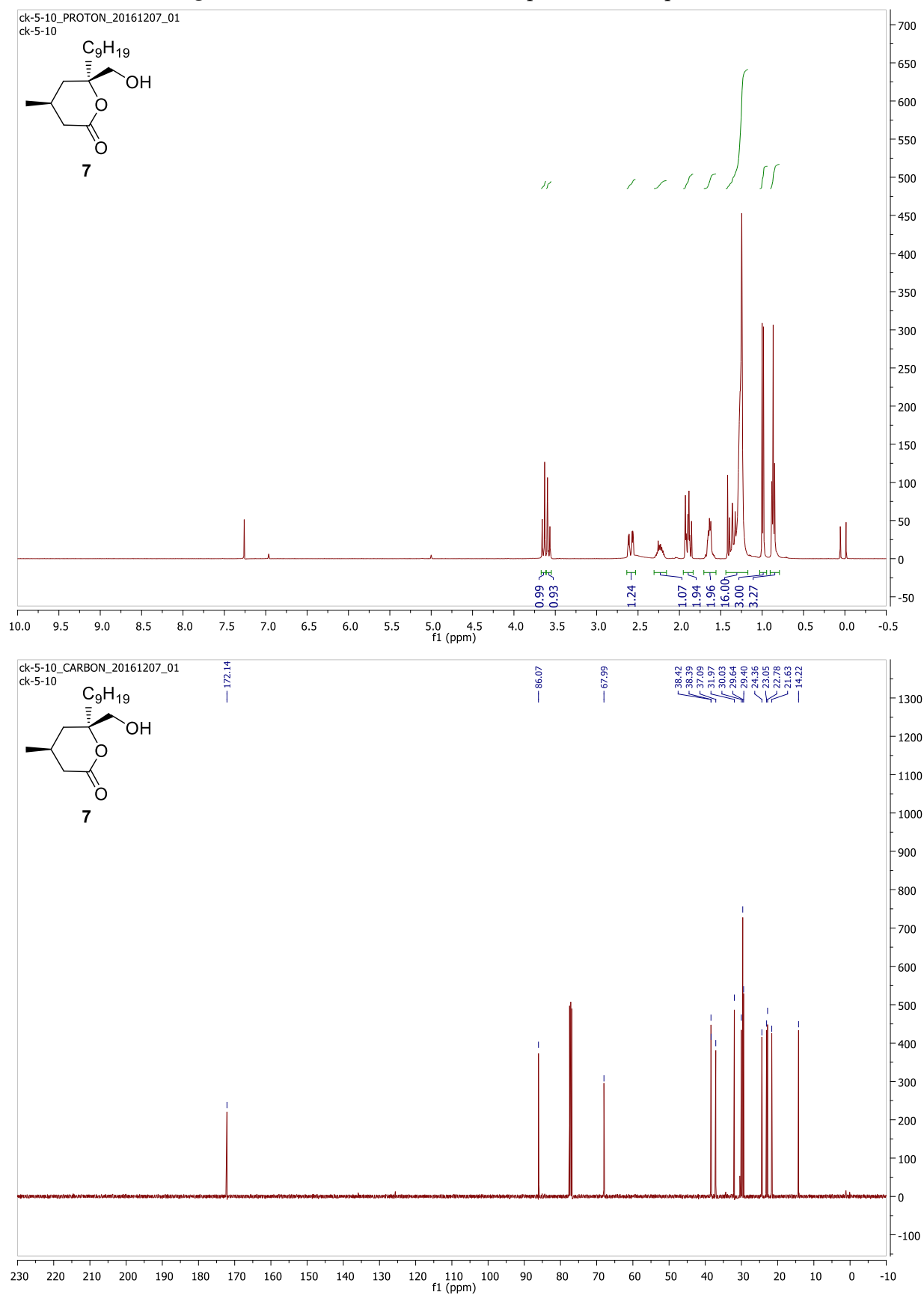
Figure S13:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 21



**Figure S14:**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 8



**Figure S15:  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR Spectra of Compound 7**



## Biological Testing

### Antimicrobial testing of compounds

**Preparation of compounds.** Samples were reconstituted into an appropriate volume of DMSO to achieve a final concentration of 10 mg/mL.

**Antibacterial activity testing – determination of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC).** Samples of each of these chemical compounds were reconstituted into an appropriate volume of DMSO to achieve a final concentration of 10 mg/mL. MIC values for these compounds was determined by two-fold broth microdilution in 96-well microtiter plates. Briefly, overnight cultures of *Escherichia coli* ATCC 25922, *Escherichia coli* 4, MRSA ATCC 43300 and MRSA 06/04 (see table S1 for further information about the isolates) were diluted in sterilised PBS to approximately  $10^5$  CFU/mL. Aliquots of 5  $\mu$ L were then transferred to separate wells in a 96-well plate that contained 100  $\mu$ L of each compound at varying concentrations (ranging from 100-0.195  $\mu$ g/mL) prepared from two-fold serial dilutions in Mueller-Hinton (MH) broth. Plates were incubated at 37 °C for 18 hours using an Omnilog® automated incubator (Biolog Inc.;21124 Cabot Boulevard, Hayward, CA 94545,USA) and MIC values recorded.

Determination of the MBC values for all compounds tested above was performed in MH broth media. Again, 5 $\mu$ L were collected from the MICs 96-well plates (above) and re-inoculated into fresh sterile 96-well plates containing fresh MH. Plates were incubated under the same conditions mentioned above. The assay was performed in triplicate for each compound.

**Table S1.** UCD Centre for Food Safety strains used for determination of antibacterial activity.

	Strains	Type	Hospital/Isolation	Source	ARP (Resistant to)
Gram-negative	<i>E. coli</i> 25922	Reference	FDA strain Seattle 1946 [DSM 1103, NCIB 12210]	-	PEN; VAN; AMP; CLI; CL
	<i>E. coli</i> 4	Clinical isolate	UCD Veterinary Hospital	Bovine	AMC; AMP; C; CIP; F; Fc; Gm; N; NAL; S; Su; TET; TMP
Gram-positive	MRSA ATCC 43300	Reference	Kansas	Human	AMP; PEN; OXA; MET; AXO; CIP; LEVO; GAT; ERY; CLI
	MRSA 06/04	Clinical isolate	-	Human	AMP; PEN; OXA; MET; AXO; CIP; LEVO; GAT; ERY

**Abbreviations:** MRSA – Methicillin-resistant *Staphylococcus aureus*; ARP – Antibiotic resistant profile; AMC – Amoxicillin-Clavulanic acid; C – Chloramphenicol; F – Furazolidone; Fc – Florfenicol; Gm – Gentamycin; N – Neomycin; NAL – Nalidixic acid; S – Streptomycin; Su – Sulfonamides; TET – Tetracycline; TMP – Trimetoprim; AMP – Ampicillin; PEN – Penicillin; OXA – Oxacillin; MET – Methicillin; AXO – Ceftriaxone; CIP – Ciprofloxacin; LEVO – Levofloxacin; GAT – Gatifloxacin; ERY – Erythromycin; CLI – Clindamycin; CL – Cephalixin; VAN - Vancomycin.



**Table S2.** Antibacterial activity of compounds tested – MIC and MBC results (triplicates).

Compound	<i>E. coli</i> 25922		<i>E. coli</i> 4		MRSA ATCC 43300		MRSA 06/04	
	MIC <sup>[b]</sup>	MBC	MIC	MBC	MIC	MBC	MIC	MBC
<b>3</b>	>100	>100	>100	>100	>100	>100	>100*	>100
<b>4</b>	>100	>100	>100	>100	>100	>100	>100*	>100
<b>5</b>	>100	>100	>100	>100	<b>12.5</b>	<b>12.5</b>	<b>12.5</b>	<b>50</b>
<b>6</b>	>100	>100	>100	>100	>100	>100	>100*	>100
<b>S1</b>	>100	>100	>100	>100	<b>25</b>	<b>25</b>	<b>25</b>	<b>50</b>
<b>7</b>	>100	>100	>100	>100	<b>50</b>	<b>50</b>	<b>50</b>	<b>50</b>
<b>8</b>	>100	>100	>100	>100	<b>50</b>	<b>100</b>	<b>50</b>	<b>50</b>

\* denotes a change in strain phenotype [b] MIC – minimum inhibitory concentration, MBC – minimum bactericidal concentration. Values are given in mg/L. Bold denotes compounds that showed activity against the tested strains. The maximum concentration of compounds tested was 100 mg/L.

The intermediates in the synthesis of **3-6** were also tested and it was observed one compound, **S1**, displayed anti-MRSA activity and was stable for the duration of the assay. However, **5** was found to be more potent.

