

Genome Sequencing and Analysis of *Trichoderma* (Hypocreaceae) Isolates Exhibiting Antagonistic Activity against the Papaya Dieback Pathogen, *Erwinia mallotivora*

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Supplementary Information

Table S1. Concentration of the extracted gDNA measured through Qubit.

Fungal isolate	DNA Concentration (ng/ μ l)
RA3a	127
RA5	168
RA6	103

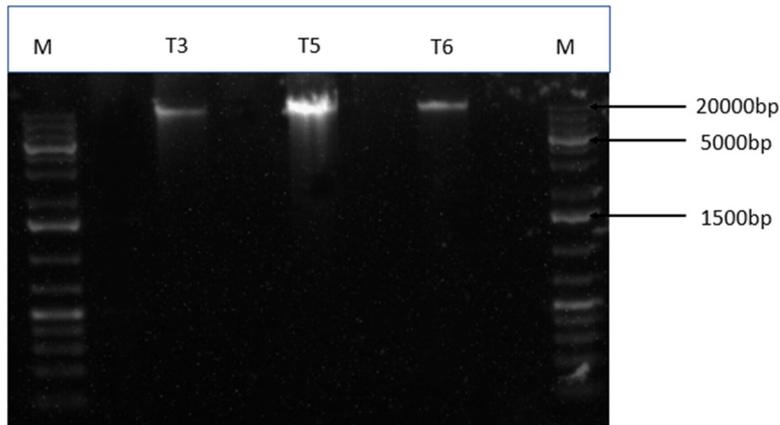


Figure S1. HMW genomic DNA of the three *Trichoderma* isolates. Lane M: GeneRuler 1 Kb Plus DNA ladder (Thermo Scientific). Lane T3: 1 μ l of genomic DNA purified from isolate RA3a. Lane T5: 1 μ l of genomic DNA purified from isolate RA5. Lane T6: 1 μ l of genomic DNA purified from isolate RA6.

Table S2. Primers and PCR amplicon sizes for molecular identification of isolates.

Marker	Forward primer	Primer sequence 5'->3'	Reverse primer	Primer sequence 5'->3'	Amplicon size (bp)	Reference
Internal Transcribed Spacer (ITS)	ITS1F	CTTGGTCATTTAGAGGAAGTAA	ITS4	TCCTCCGCTTATTGATATGC	650	(Schoch et al., 2012)
Translation elongation factor 1 alpha (<i>tef1</i>) gene	EF1-728F	CATCGAGAAGTTCGAGAAGG	TEF1LLErev	AACTTGCAGGCAATGTGG	1,300	(Druzhinina and Kubicek, 2005)
RNA polymerase B subunit II (<i>rpb2</i>) gene	RPB2-5f	GAYGAYMGWGATCAYTTYGG	RPB2-7cR	CCCATRGCTTGYTTRCCCAT	1,200	(Liu et al., 1999)

Table S3. Details on the number of predicted non-coding RNAs identified in each genome.

Non-coding RNA's	RA3a	RA5	RA6
Riboswitches	1 (0.30%)	1 (0.29%)	1 (0.29%)
Ribozymes	2 (0.60%)	2 (0.59%)	2 (0.57%)
Ribosomal RNAs (rRNA)	89 (26.49%)	97 (28.53%)	101 (29.02%)
Small nuclear RNAs (snRNA)	9 (2.68%)	9 (2.65%)	9 (2.59%)
Small nucleolar RNAs (snoRNA)	31 (9.23%)	28 (8.24%)	30 (8.62%)
Small RNAs (sRNA)	4 (1.19%)	4 (1.18%)	4 (1.15%)
Transfer RNAs (tRNA)	195 (58.04%)	193 (56.76%)	196 (56.32%)
Other	5 (1.49%)	5 (1.47%)	5 (1.44%)
intron	0	1 (0.29%)	0

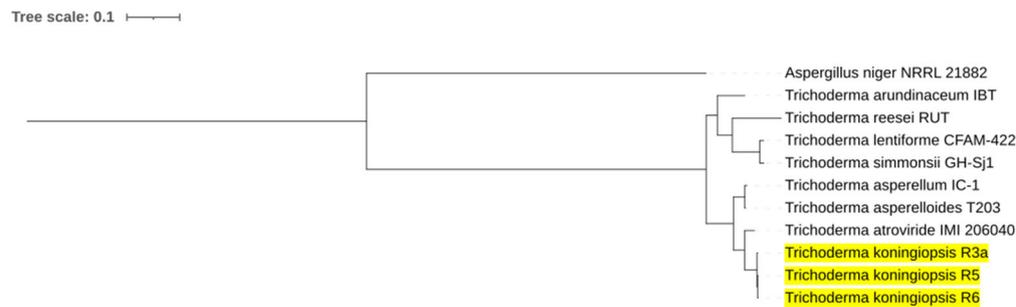


Figure S2. Phylogenomic tree from alignment of 3,392 single-copy orthologous proteins from selected *Trichoderma* spp. The three isolates sequenced in the present work are highlighted in yellow. Bootstrap values were omitted from the tree as all the branches showed 100% bootstrap value.

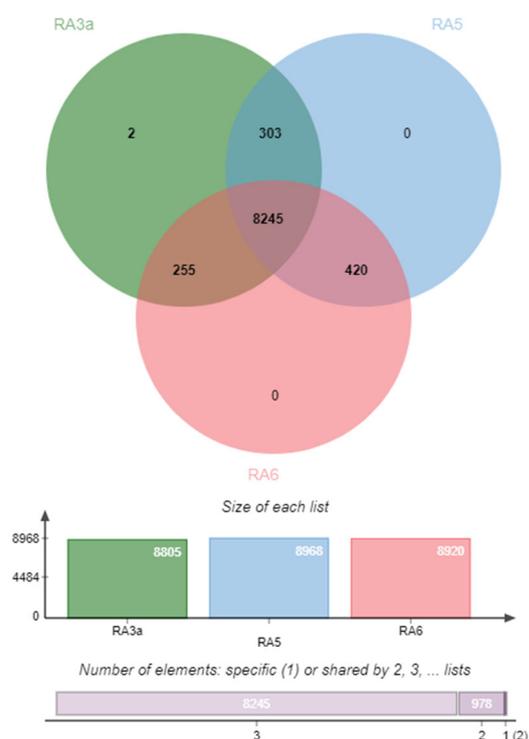


Figure S3. Shared and unique orthologous protein group among isolates sequenced in this study as inferred from OrthoVenn2 web server analysis.

Table S4. Distribution of predicted BGCs found across the fungal genomes.

Gene Type	Fungal Genome	Region	Span		Cluster size (kb)	Most similar BGC according to AntiSMASH (% indicates the proportion of genes with similarities)
			From	To		
Hybrid PKS/NRPS	RA3a	2.1	83,384	178,034	94.6	fusaric acid, 41% (5/12)
	RA5	7.3	3,470,945	3,555,885	84.9	fusaric acid, 50% (6/12)
	RA6	4.4	3,796,740	3,881,680	84.9	fusaric acid, 50% (6/12)
PKS	RA3a	6.1	78,041	120,092	42	naphthopyrone, 100% (1/1)
	RA5	5.9	3,799,152	3,841,850	42.6	naphthopyrone, 100% (1/1)
	RA6	5.9	3,806,331	3,849,029	42.6	naphthopyrone, 100% (1/1)
PKS	RA3a	6.3	725,578	749,075	23.4	neurosporin A, 100% (1/1)
	RA5	5.8	3,170,168	3,193,667	23.5	neurosporin A, 100% (1/1)
	RA6	5.8	3,180,784	3,200,851	20.0	neurosporin A, 100% (1/1)
Hybrid PKS/Terpene	RA3a	9.1	464,928	537,653	72.7	ascochlorin, 38% (3/8)
	RA5	1.6	6,455,881	6,532,090	76.2	ascochlorin, 38% (3/8)
	RA6	8.4	1,744,822	1,821,031	76.2	ascochlorin, 38% (3/8)
Terpene	RA3a	9.2	636,393	658,750	22.3	clavaric acid, 100% (1/1)
	RA5	1.4	6,335,790	6,356,639	20.8	clavaric acid, 100% (1/1)
	RA6	8.3	1,623,721	1,646,078	22.3	clavaric acid, 100% (1/1)

Table S5. Putative salicylaldehyde cluster found across the three sequenced fungal genomes.

Gene Locus	Homologue (% Identity/% similarity), Organism, Accession number	Proposed function
RA3a_007616, RA5_007274, RA6_006441	Highly reducing polyketide synthase <i>virA</i> , 84%/92%, <i>Trichoderma virens</i> Gv29-8, G9N4B2.1	Biosynthesis of the polyketide unit
RA3a_007617, RA5_007273, RA6_006440	Short-chain dehydrogenase <i>virB</i> , 75%/81%, <i>Trichoderma virens</i> Gv29-8, G9N4B1.1	Tailoring of the scaffold
RA3a_007618, RA5_007272, RA6_006439	Cupin-domain-containing oxidoreductase <i>virC</i> , 88%/95%, <i>Trichoderma virens</i> Gv29-8, G9N4B0.1	
RA5_007271	Short-chain dehydrogenase <i>virD</i> , 62%/67%, <i>Trichoderma virens</i> Gv29-8, G9N4A9.1	
RA3a_007619, RA5_007270, RA6_006438	Cytochrome P450 monooxygenase <i>virE</i> , 74%/83%, <i>Trichoderma virens</i> Gv29-8, G9N4A8.1	
RA3a_007620, RA5_007269, RA6_006437	Adenylosuccinate lyase, 49%/63%, <i>Saccharomyces cerevisiae</i> S288C, Q05911.1	
RA3a_007621, RA5_007268, RA6_006436	Probable oxidoreductase <i>virH</i> , 81%/89%, <i>Trichoderma virens</i> Gv29-8] Sequence ID: G9N4A5.1	
RA3a_007622, RA5_007267, RA6_006435	Glutamine--tRNA ligase protein <i>virJ</i> , 90%/93%, <i>Trichoderma virens</i> Gv29-8 G9N4A3.1	
RA3a_007623, RA5_007266	FAD-linked oxidoreductase <i>virF</i> , 59%/73%, <i>Trichoderma virens</i> Gv29-8, G9N4A7.1	

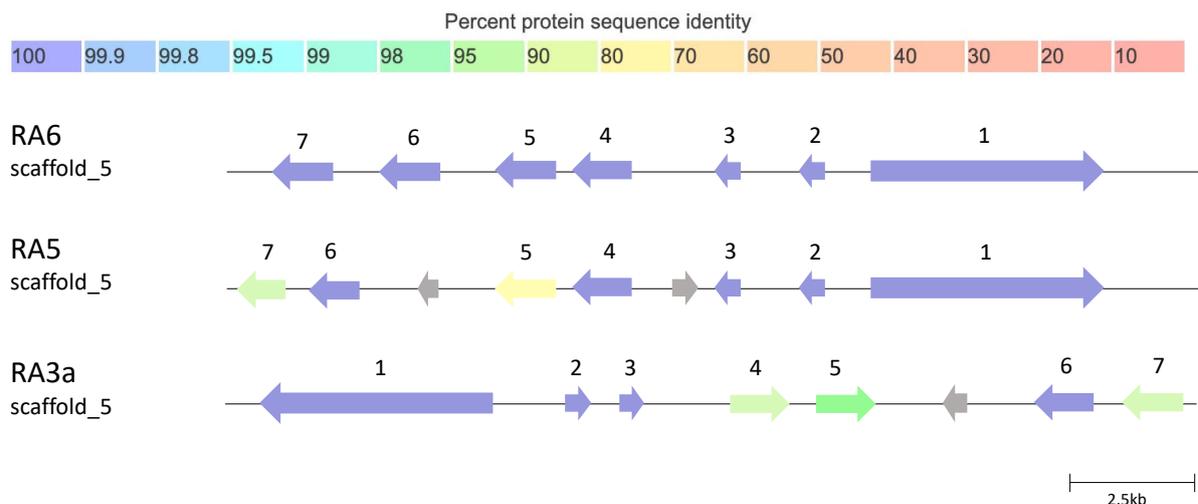


Figure S4. Protein sequence comparison of the putative salicylaldehyde cluster found in the genome of strains RA6, RA5, and RA3a. The proteome comparison service from PATRIC

3.6.12 (Davis et al., 2020) was used to carry out protein sequence comparison using bi-directional BLASTP. Default parameters were used; i.e Minimum % coverage was set to 30%, BLAST E value was set to 1e-5 and Minimum % Identity was set to 10%. The protein sequence of the RA6 cluster was used as a reference sequence to be compared to the RA5 cluster and the RA3a cluster. The sequences shown in grey are sequences that did not show any similarity between clusters. Key: 1, HR-PKS; 2, Short chain dehydrogenase; 3: Cupin-domain-containing oxidoreductase; 4, Cytochrome P450; 5, Lyase; 6, Oxidoreductase; 7, Oxidoreductase.

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