

## Supplementary

# ***Fodinisporobacter ferrooxydans* gen. nov., sp. nov., a Spore-forming Ferrous-oxidizing Bacterium Isolated from a Polymetallic Mine**

Zhen Jiang<sup>1,2†</sup>, Xiutong Li<sup>1,2†</sup>, Zonglin Liang<sup>1,2</sup>, Zebao Tan<sup>3</sup>, Nan Zhou<sup>1</sup>, Ying Liu<sup>1</sup>, Zhenghua Liu<sup>4</sup>, Huaqun Yin<sup>4</sup>, Kun Luo<sup>3</sup>, Supawadee Ingsriswang<sup>5</sup>, Shuangjiang Liu<sup>1,2,6\*</sup>, Chengying Jiang<sup>1,2\*</sup>

<sup>1</sup> State Key Laboratory of Microbial Resources, Institute of Microbiology, Chinese Academy of Sciences, Beijing 100101, China

<sup>2</sup> University of Chinese Academy of Sciences, Beijing 100049, China

<sup>3</sup> College of Plant Protection, Hunan Agricultural University, Changsha 410128, Hunan, China

<sup>4</sup> Key Laboratory of Biomaterials of Ministry of Education, School of Minerals Processing and Bioengineering, Central South University, Changsha 410083, China

<sup>5</sup> Thailand Bioresource Research Center (TBRC), National Center for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency (NSTDA), Pathum Thani 12120, Thailand

<sup>6</sup> State Key Laboratory of Microbial Biotechnology, Shandong University, Qingdao 266237, China

\*Corresponding author: Chengying Jiang, [jiangcy@im.ac.cn](mailto:jiangcy@im.ac.cn); Shuangjiang Liu, [liusj@im.ac.cn](mailto:liusj@im.ac.cn)

<sup>†</sup>These authors contributed equally to this study.

**Table S1.** Results of acid production (API 50CH), assimilation of carbon sources (API 20NE), enzymatic activities (API ZYM) and other biochemical characteristics (API 20NE) of strain MYW30-H2<sup>T</sup>. +, positive; –, negative; W, weak positive.

<b>Acid Production from (API 50CH):</b>		<b>Assimilation of (API 20NE):</b>	
Glycerol	–	D-Glucose	+
Erythritol	–	L-Arabinose	+
D-Arabinose	–	D-Mannose	+
L-Arabinose	–	D-Mannitol	+
D-Ribose	–	N-acetyl-glucosamine	–
D-Xylose	–	D-Maltose	–
L-Xylose	–	Gluconate	–
D-Adonitol	–	Capric acid	–
Methyl- $\beta$ -D-xylopyranoside	–	Adipic acid	–
D-Galactose	–	Malic acid	–
D-Glucose	–	Trisodium citrate	–
D-Fructose	–	Phenylacetic acid	–
D-Mannose	–	<b>Enzymatic activities of (API ZYM):</b>	
L-Sorbose	+	Alkaline phosphatase	–
L-Rhamnose	–	Esterase (C 4)	–
Dulcitol	+	Esterase lipase (C 8)	–
Inositol	+	Lipase (C 14)	–
D-Mannitol	–	Leucine arylamidase	–
D-Sorbitol	–	Valine arylamidase	–
Methyl- $\alpha$ -D-mannopyranoside	+	Cystine arylaminase	–
Methyl- $\alpha$ -D-glucopyranoside	–	Trypsin	–
N-acetyl-glucosamine	+	$\alpha$ -Chymotrypsin	–
Amygdalin	+	Acid phosphatase	+
Arbutin	–	Naphthol-AS-BI-phosphohydrolase	+
Esculin	+	$\alpha$ -galactosidase	–
Salicine	–	$\beta$ -galactosidase	–
D-Cellobiose	–	$\beta$ -glucuronidase	–
D-Maltose	–	$\alpha$ -glucosidase	+
D-Lactose	–	$\beta$ -glucosidase	–
D-Melibiose	–	N-acetyl-glucosaminidase	W
D-Sucrose	–	$\alpha$ -mannosidase	–
D-Trehalose	–	$\beta$ -fucosidase	–

Inulin	+	<b>Other biochemical characteristics (API 20NE):</b>	
D-Melezitose	–	Nitrate reduction	+
D-Raffinose	–	indole production	–
Starch	+	Glucose fermentation	+
Glycogen	+	Arginine dihydrolase	–
Xylitol	–	Urea hydrolysis (Urease)	W
Gentiobiose	–	Esculin hydrolysis	+
D-Turanose	–	Gelatin hydrolysis ( $\beta$ -glucosidase)	–
D-Lyxose	–	$\beta$ -galactosidase	+
D-Tagatose	+		
D-Fucose	+		
L-Fucose	+		
D-Arabitol	–		
L-Arabitol	+		
Gluconate	+		
2-ketogluconate	+		
5-ketogluconate	–		

**Table S2.** General genomic characteristics of strain MYW30-H2<sup>T</sup>.

<b>Characteristics</b>	<b>MYW30-H2<sup>T</sup></b>
<b>INSDC number</b>	CP089291
<b>Sequencing depth</b>	328.8×
<b>Number of replicons</b>	1
<b>Genome size</b>	4856687 bp
<b>G+C content</b>	44.2%
<b>Total genes</b>	4620
<b>tRNA genes</b>	114
<b>5S rRNA genes</b>	12
<b>16S rRNA genes</b>	12
<b>23S rRNA genes</b>	12
<b>Other RNA genes</b>	4