

Supplementary material

Fish DNA Sensors for Authenticity Assessment—Application to Sardine Species Identification

Myrto Kakarelidou ¹, Panagiotis Christopoulos ¹, Alexis Conides ², Despina P. Kalogianni ^{1,*}
and Theodore K. Christopoulos ^{1,3,*}

¹ Analytical/Bioanalytical Chemistry & Nanotechnology Group, Department of Chemistry, University of Patras, Rio, 26504 Patras, Greece; myrtokakarelidou@gmail.com (M.K.); pkchristop@gmail.com (P.C.)

² Hellenic Centre for Marine Research, Institute for Marine Biological Resources, 46.7 km Athens-Sounion, Anavyssos, 19013 Attika, Greece; conides@hcmr.gr

³ Institute of Chemical Engineering Sciences, Foundation for Research and Technology Hellas (FORTH/ICE-HT), Platani, 26504 Patras, Greece

* Correspondence: kalogian@upatras.gr (D.P.K.); tchrist@upatras.gr (T.K.C.)

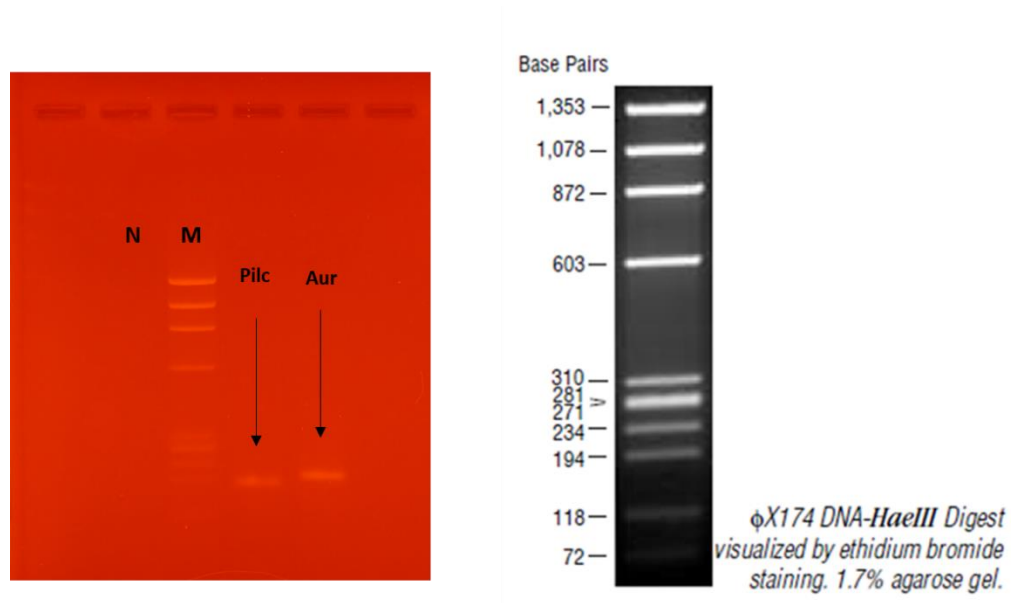


Figure S1. Electropherogram of PCR products for *S. pilchardus* with *S. aurita*. N: negative, M: DNA marker (ϕ X174 DNA HaeIII Digest on the right), Pilc: *S. pilchardus* and Aur: *S. aurita*.

Mixtures of PCR products

% Content of S. aurita in S. pilchardus

DNA sensor with S. pilchardus probe

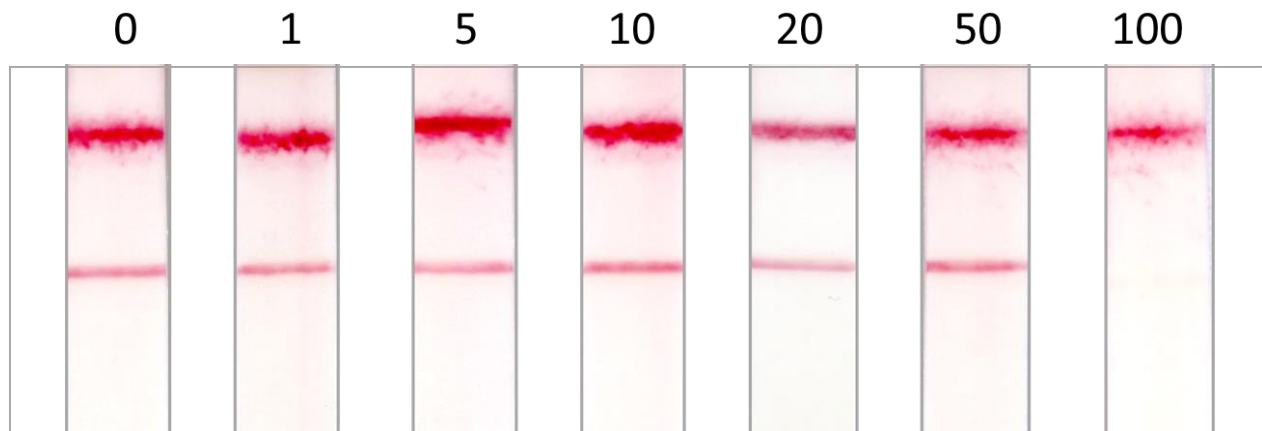


Figure S2. Mixtures of PCR products. The mixtures contained 0 – 100 % PCR product from *S. aurita* in PCR product from *S. pilchardus* with a total amount of 100 fmol on the strip.

Mixtures of processed samples

% Content of S. aurita in S. pilchardus

DNA sensor with S. pilchardus probe

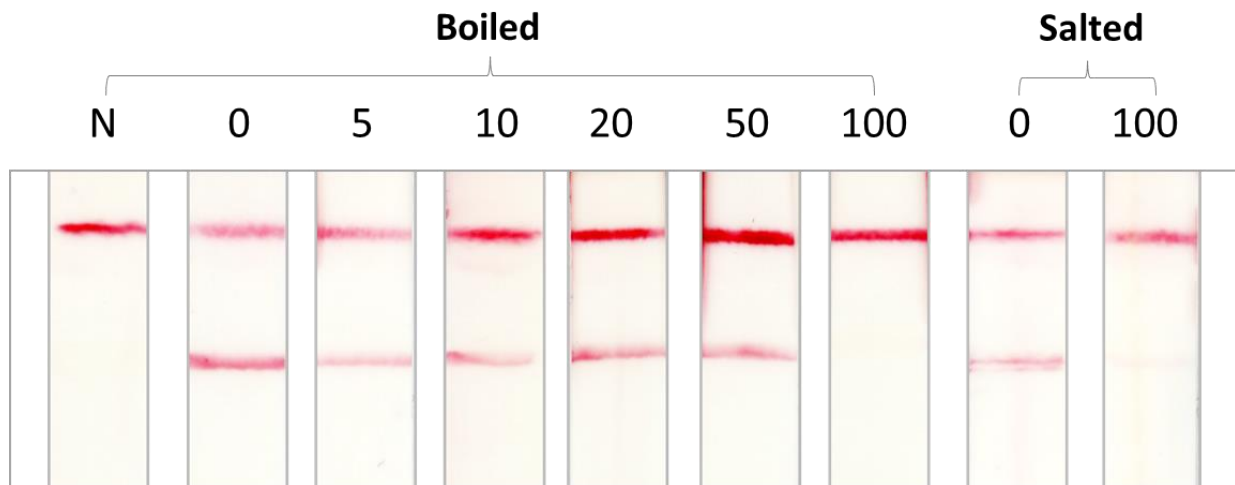


Figure S3. Mixtures of processed samples. The mixtures contained 0 – 100 % of *S. aurita* (tissue) in *S. pilchardus*.

Table S1. Comparison of methods for sardines' adulteration detection

Method	Species examined	LOD	Discrimination capability	Quantitation capability	Ref.
PCR-RFLP and phylogenetic analysis	<i>S. pilchardus</i> <i>S. aurita</i> <i>S. melanostictus</i> <i>S. caeruleus</i> <i>S. maderensis</i>	-	Low	-	12
PCR-RFLP and DNA sequencing	<i>S. pilchardus</i> <i>S. aurita</i> <i>S. brasiliensis</i> <i>S. sagax</i> <i>S. caeruleus</i>	-	Low for PCR-RFLP and high for DNA sequencing	✓ for DNA sequencing	13
PCR-RFLP and phylogenetic analysis	<i>S. pilchardus</i> <i>S. aurita</i>	-	Low	-	14
DNA sequencing and phylogenetic analysis	<i>S. pilchardus</i> <i>S. aurita</i>	5%	High	✓	6
DNA sequencing and phylogenetic analysis	<i>S. pilchardus</i> <i>S. aurita</i> <i>S. sagax</i> <i>S. caeruleus</i> <i>S. melanostictus</i> <i>S. maderensis</i> <i>S. longiceps</i>	-	High	✓	15
DNA sequencing and phylogenetic analysis	<i>S. pilchardus</i> <i>S. aurita</i> <i>S. longiceps</i> <i>S. lemuru</i> <i>S. brasiliensis</i> <i>S. gibbose</i> <i>S. jussieu</i> <i>S. fimbriata</i> <i>S. tawilis</i>	-	High	✓	16
PCR and agarose gel electrophoresis	<i>S. pilchardus</i>	-	Medium	-	17
PCR and agarose gel electrophoresis	<i>S. pilchardus</i> <i>S. aurita</i> <i>S. maderensis</i>	-	Medium	-	18
PCR and agarose gel electrophoresis	<i>S. pilchardus</i>	-	Medium	-	19
Exon-primed intron-crossing (EPIC) PCR with acrylamide gel electrophoresis	<i>S. pilchardus</i>	-	Medium	-	20
Real-time PCR / SYBR Green	<i>S. pilchardus</i>	-	Medium	✓	21
Real-time PCR / Taqman probes	<i>S. pilchardus</i> <i>S. aurita</i>	-	High	✓	5
Real-time PCR / Taqman probes	<i>S. pilchardus</i>	-	High	✓	22
Real-time PCR / Melting curve analysis	<i>S. pilchardus</i>	-	High	✓	23
Real-time PCR / High-resolution Melting curve analysis (HRM)	<i>S. pilchardus</i>	-	High	✓	24
DNA sensor	<i>S. pilchardus</i> <i>S. aurita</i>	5%	High	-	This work