

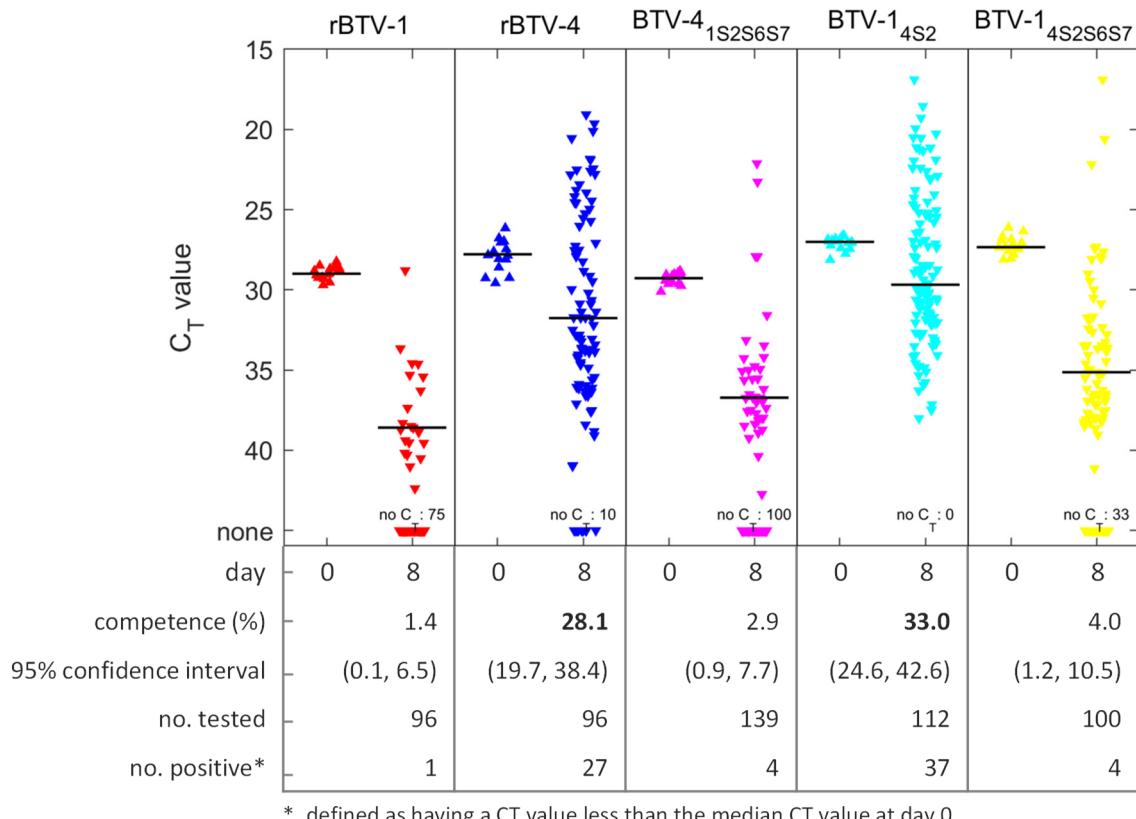
# Identification of a BTV-Strain-Specific Single Gene That Increases *Culicoides* Vector Infection Rate

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**Figure S1.** Reverse-engineered and reassortant BTV strain C<sub>T</sub> values in *C. sonorensis* after feeding. Individuals were tested either immediately after feeding (day 0) or after incubation for 8 days at 25 °C (day 8). Symbols depict the C<sub>T</sub> values for individual insects and horizontal bars depict the median C<sub>T</sub> value (excluding those individuals with no C<sub>T</sub> value). C<sub>T</sub> values were obtained from RT-qPCR assays of one independent experiment for each strain. Comparison amongst strains was carried out using a binomial-family generalized linear model with a logit link function. Note: Figure placed here for comparison to literature data, which is usually expressed in observed C<sub>T</sub> values.

**Table S1.** Sequences of cloning primers. The upstream (forward) primers contained a T7 RNA polymerase promoter sequence and a restriction site for unidirectional cloning, while the downstream (reverse) primers contained restriction sites for cloning and for linearization at the 3' end of the segment [45].

BTM clone, segment	Sequence (5'-3')
<b>BTM-1 MOR2006/06</b>	
Seg-1 forward	TCTAGCGGATCCTAATACGACTCACTATAAGTAAAATGCAATGGTCGCAATC
Seg-1 reverse	TACAGTAAGCGGCCGCGTCTCAGTAAGTGTAAATGCGGCCGCGTGC
Seg-2 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAATAGTAGCGCGATGGATG
Seg-2 reverse	TACAGTAAGCGGCCGCGTCTCAGTAAGTCTAAATAGTGC CGGGATC
Seg-3 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAATTCCGTAGCCATGGCTG
Seg-3 reverse	TACAGTAAGCGGCCGCGTCTCAGTAAGTGTCCCGCTGCCGC
Seg-4 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAACATGCCTGAGCCACACG
Seg-4 reverse	CGTAAGCGGCCGCGTCTCAGTAAGTTGTACATGCCCTCTC
Seg-5 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAAAGTTCTCTAGTTGGC
Seg-5 reverse	TACAGTAAGCGGCCGCGTCTCAGTAAGTTGAAAAGTTCTAGTAGAGTG
Seg-6 forward	CTAGCGAATTCTAATACGACTCACTATAAGTAAAAAGTCGACCCCTAGCGAA
Seg-6 reverse	TACAGTAAGCGGCCGCGGTCTCAGTAAGTGTAAAGTGCCTCCGT CGC
Seg-7 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAATCTATAGAGATGGACA
Seg-7 reverse	TACAGTAAGCGGCCGCGGTCTCAGTAAGTGTAACTAAGAGACGTTG
Seg-8 forward	TCTAGCGGATCCTAATACGACTCACTATAAGTAAAAATCCTTGAGTCATGGAG
Seg-8 reverse	CAGTAAGCGGCCGCGTCTCAGTAAGTGTAAATTCCCCCTAACCC
Seg-9 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAAATCGCATATGTCAGCTG
Seg-9 reverse	TACAGTAAGCGGCCGCGTCTCAGTAAGTGTAAAATGCCCTACGTCA
Seg-10 forward	TCTAGCGAATTCTAATACGACTCACTATAAGTAAAAGTGTGCGCTGCCATGCT
Seg-10 reverse	TACAGTAAGCGGCCGCGTCTCAGTAAGTGTAGCGCCATACCCTC
<b>BTM-4 MOR2004/02</b>	
Seg-1 forward	GTGATGGATCCTAATACGACTCACTATAAGTAAAATGCAATGGTCGCAAT
Seg-1 reverse	GGTATGCGGCCGCGGTCTCAGTAAGTGTAAATGCGGCCGCGTGC
Seg-2 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAAGAGTGTCCCAACATGG
Seg-2 reverse	GGTATGCGGCCGCGGTCTCAGTAAGTGTAAAGAGGCCACAGG
Seg-3 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAATTCCGTAGCCATGG
Seg-3 reverse	GGTATGCGGCCGCGTCTCAGTAAGTGTGTTCCCGCTGCCG

Seg-4 forward	GTGATGGATCTAATACGACTCACTATAAGTAAAACATGCCTGAGCCACA
Seg-4 reverse	GGTATCGGGCCGCCGTCTCAGTAAGTTACATGCCCTCT
Seg-5 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAAAGTTCTCTAGTTG
Seg-5 reverse	GGTATCGGGCCGCCGTCTCAGTAAGTTGAAAAGTTCTAGTAG
Seg-6 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAAAGTGTCTCCTACT
Seg-6 reverse	GGTATCGGGCCGCCGTCTCAGTAAGTGTAAAGCTTCTCCCTCG
Seg-7 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAAATCTATAGAGATGG
Seg-7 reverse	GGTATCGGGCCGCGAAGACCAGTAAGTGTAACTAAGAGACG
Seg-8 forward	GTGATGGATCTAATACGACTCACTATAAGTAAAAATCCTCGAGTCATG
Seg-8 reverse	GGTATCGGGCCGCCGTCTCA GTAAAGTGTAAAATCCCCCTCT
Seg-9 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAAATCGCATATGTCAG
Seg-9 reverse	GGTATCGGGCCGCCGTCTCAGTAAGTGTAAAATGCCCTACG
Seg-10 forward	GTGATGAATTCTAATACGACTCACTATAAGTAAAAAGTGTGCTGCCATG
Seg-10 reverse	GGTATCGGGCCGCCGTCTCAGTAAGTGTAGGCCCGATA

**Table S2.** Restriction enzymes used for reverse genetics.

BTV clone, segment	Upstream site	Downstream site
<b>BTV-1 MOR2006/06</b>		
Seg-1	BamHI	BsmBI
Seg-2	EcoRI	BsmBI
Seg-3	EcoRI	BsmBI
Seg-4	BamHI	BsaI
Seg-5	EcoRI	BsmBI
Seg-6	EcoRI	BsaI
Seg-7	EcoRI	BsaI
Seg-8	BamHI	BsmBI
Seg-9	EcoRI	BsmBI
Seg-10	EcoRI	BsmBI
<b>BTV-4 MOR2004/02</b>		
Seg-1	BamHI	BsaI
Seg-2	EcoRI	BsaI
Seg-3	EcoRI	BsmBI
Seg-4	BamHI	BsaI
Seg-5	EcoRI	BsmBI
Seg-6	EcoRI	BsmBI
Seg-7	EcoRI	BbsI (BpiI)
Seg-8	BamHI	BsmBI
Seg-9	EcoRI	BsmBI
Seg-10	EcoRI	BsmBI

**Table S3.** Rescued and reassortant BTV strains [BSR3 passage] deposited in the Orbivirus Reference Collection (ORC) at the Pirbright Institute, UK ([https://www.reoviridae.org/dsRNA\\_virus\\_proteins/ReoID/rescued%20BTV.html](https://www.reoviridae.org/dsRNA_virus_proteins/ReoID/rescued%20BTV.html)); where reverse engineered rBTV-1 is derivative of wtBTV-1 MOR2006/06 and rBTV-4 is derivative of wtBTV-4 MOR2004/02.

ORC ref. number <sup>a</sup>	Virus name <sup>b</sup>	Genome segment(s) <sup>c</sup>									
		Seg-1	Seg-2	Seg-3	Seg-4	Seg-5	Seg-6	Seg-7	Seg-8	Seg-9	Seg-10
BTV-RV0010*	rBTV-4										
BTV-RV0002	BTV-4 <sub>1S3</sub>			1							
BTV-RV0007	BTV-4 <sub>1S9</sub>									1	
BTV-RV0022*	BTV-4 <sub>1S2S6S7</sub>		1				1	1			
BTV-RV0042*	rBTV-1										
BTV-RV0021*	BTV-1 <sub>4S2</sub>		4								
BTV-RV0035	BTV-1 <sub>4S3</sub>			4							
BTV-RV0040	BTV-1 <sub>4S9</sub>									4	
BTV-RV0038*	BTV-1 <sub>4S2S6S7</sub>		4				4	4			

<sup>a</sup>, all strains used in this study were after additional propagation to cell passage: BSR4.

<sup>b</sup>, the first section of the name refers to the BTV backbone, with the second subscripted section referring to the heterologous segment(s). In this study, all strains were used after additional propagation to cell passage: BSR4

<sup>c</sup>, white-boxes indicate those genome segments derived from wtBTV-1 MOR2006/06; grey-boxes indicate genome segments derived from wtBTV-4 MOR2004/02.

\*, used in this study for in vivo oral infection of *C. sonorensis* experiments at cell passage: BSR4.

**Table S4.** Genome copy numbers obtained from BTV replication kinetics study for wild type (wtBTV-1 and wt-BTV-4) and reverse engineered (rBTV-1 and rBTV-4) strains.

dpi	wtBTV-1			wtBTV-4			rBTV-1			rBTV-4		
	mean gcn	sd	n									
0	3.41×10 <sup>2</sup>	1.37×10 <sup>2</sup>	6	5.98×10 <sup>3</sup>	2.75×10 <sup>3</sup>	6	5.89×10 <sup>2</sup>	8.97×10 <sup>2</sup>	6	4.24×10 <sup>2</sup>	2.65×10 <sup>2</sup>	12
1	7.64×10 <sup>2</sup>	2.97×10 <sup>2</sup>	6	7.95×10 <sup>3</sup>	2.14×10 <sup>3</sup>	6	5.41×10 <sup>2</sup>	2.36×10 <sup>2</sup>	6	9.37×10 <sup>2</sup>	5.16×10 <sup>2</sup>	12
2	5.07×10 <sup>3</sup>	5.36×10 <sup>3</sup>	6	7.76×10 <sup>3</sup>	1.05×10 <sup>3</sup>	6	8.49×10 <sup>2</sup>	3.99×10 <sup>2</sup>	6	1.35×10 <sup>3</sup>	7.42×10 <sup>2</sup>	12
3	7.49×10 <sup>4</sup>	1.03×10 <sup>5</sup>	6	1.20×10 <sup>4</sup>	3.24×10 <sup>3</sup>	6	1.86×10 <sup>3</sup>	6.88×10 <sup>2</sup>	6	2.68×10 <sup>3</sup>	1.91×10 <sup>3</sup>	12
6	2.54×10 <sup>6</sup>	2.57×10 <sup>6</sup>	6	1.42×10 <sup>6</sup>	1.97×10 <sup>6</sup>	6	7.41×10 <sup>5</sup>	4.53×10 <sup>5</sup>	6	3.25×10 <sup>5</sup>	4.51×10 <sup>5</sup>	12
7	5.23×10 <sup>6</sup>	4.20×10 <sup>6</sup>	6	4.41×10 <sup>6</sup>	3.10×10 <sup>6</sup>	6	2.85×10 <sup>6</sup>	7.33×10 <sup>5</sup>	6	1.60×10 <sup>6</sup>	1.86×10 <sup>6</sup>	12
10	9.22×10 <sup>6</sup>	2.88×10 <sup>6</sup>	6	1.26×10 <sup>7</sup>	6.42×10 <sup>6</sup>	6	1.68×10 <sup>7</sup>	1.53×10 <sup>7</sup>	6	6.42×10 <sup>6</sup>	4.05×10 <sup>6</sup>	12

dpi, days post infection; gcn, quantitated genome copy number per RT-qPCR reaction; sd, standard deviation; n, number of replicates (i.e. one independent experiment is composed of three replicates).

**Table S5.** Genome copy numbers obtained from BTV replication kinetics study for reverse engineered (rBTV-1 and rBTV-4) and reassortant (BTV-1<sub>4S2</sub>, BTV-1<sub>4S2S6S7</sub> and BTV-4<sub>1S2S6S7</sub>) strains.

dpi	BTV-1 <sub>4S2</sub>			BTV-1 <sub>4S2S6S7</sub>			BTV-4 <sub>1S2S6S7</sub>		
	mean gcn	sd	n	mean gcn	sd	n	mean gcn	sd	n
0	1.50×10 <sup>3</sup>	8.20×10 <sup>1</sup>	3	5.00×10 <sup>2</sup>	2.75×10 <sup>2</sup>	3	3.09×10 <sup>2</sup>	1.31×10 <sup>2</sup>	6
1	2.02×10 <sup>3</sup>	4.27×10 <sup>2</sup>	3	1.09×10 <sup>3</sup>	2.05×10 <sup>2</sup>	3	6.82×10 <sup>2</sup>	1.96×10 <sup>2</sup>	6
2	2.89×10 <sup>3</sup>	1.99×10 <sup>3</sup>	3	1.19×10 <sup>3</sup>	9.70×10 <sup>1</sup>	3	2.23×10 <sup>3</sup>	5.65×10 <sup>2</sup>	6
3	1.38×10 <sup>4</sup>	6.00×10 <sup>3</sup>	3	5.77×10 <sup>3</sup>	3.40×10 <sup>3</sup>	3	8.99×10 <sup>3</sup>	5.63×10 <sup>3</sup>	6
6	1.04×10 <sup>6</sup>	4.03×10 <sup>5</sup>	3	5.64×10 <sup>5</sup>	1.87×10 <sup>5</sup>	3	6.17×10 <sup>5</sup>	6.79×10 <sup>5</sup>	6
7	2.72×10 <sup>6</sup>	6.14×10 <sup>5</sup>	3	1.39×10 <sup>6</sup>	1.92×10 <sup>5</sup>	3	1.80×10 <sup>6</sup>	9.68×10 <sup>5</sup>	6
10	7.03×10 <sup>6</sup>	1.73×10 <sup>6</sup>	3	4.10×10 <sup>6</sup>	2.34×10 <sup>5</sup>	3	3.68×10 <sup>6</sup>	1.93×10 <sup>6</sup>	6

dpi, days post infection; gcn, quantitated genome copy number per RT-qPCR reaction; sd, standard deviation; n, number of replicates (i.e. one independent experiment is composed of three replicates).

**Table S6.** Deviance information criterion (DIC) for models assessing variation amongst BTV strains in replication curve parameters.

Replication rate	Model*		Initial no. copies	DIC†		
	Asymptote					
	Lower	Upper				
common	common	common	common	560.7		
varies	common	common	common	474.6		
common	varies	common	common	451.5		
common	common	varies	common	531.2		
common	common	common	varies	423.6		
varies	varies	common	common	411.4		
varies	common	varies	common	468.9		
varies	common	common	varies	323.0		
common	varies	varies	common	443.0		
common	varies	common	varies	290.6		
common	common	varies	varies	402.3		
varies	varies	varies	common	411.9		
<b>varies</b>	<b>varies</b>	<b>common</b>	<b>varies</b>	<b>251.2</b>		
varies	common	varies	varies	317.5		
common	varies	varies	varies	272.9		
varies	varies	varies	varies	249.6‡		

\*, "common": parameter common to all strains; "varies": parameter varies amongst strains;

†, a model with a lower DIC is preferred to one with higher DIC;

‡, although this model has a smaller DIC than the one shown in bold, the difference is less than two and the simpler model was preferred as it has the smaller number of parameters.

**Table S7.** Comparison of posterior distributions of replication rates and time of maximum replication between BTV strains in KC cells.

Replication rate	Time of maximum replication
<b>wtBTV-4 &gt; wtBTV-1 (P=0.03)</b>	<b>wtBTV-4 &gt; wtBTV-1 (P&lt;0.001)</b>
<b>rBTV-1 &gt; wtBTV-1 (P=0.003)</b>	<b>rBTV-1 &gt; wtBTV-1 (P&lt;0.001)</b>
<b>rBTV-4 = wtBTV-4 (P=0.06)</b>	<b>rBTV-4 = wtBTV-4 (P=0.78)</b>
<b>rBTV-1 = wtBTV-4 (P=0.55)</b>	<b>rBTV-1 = wtBTV-4 (P=0.20)</b>
<b>rBTV-4 &lt; rBTV-1 (P=0.01)</b>	<b>rBTV-4 &gt; rBTV-1 (P=0.02)</b>
<b>BTV-1<sub>4S2</sub> = rBTV-1 (P=0.05)</b>	<b>BTV-1<sub>4S2</sub> &lt; rBTV-1 (P=0.05)</b>
<b>BTV-1<sub>4S3</sub> = rBTV-1 (P=0.60)</b>	<b>BTV-1<sub>4S3</sub> &lt; rBTV-1 (P&lt;0.001)</b>
<b>BTV-1<sub>4S9</sub> = rBTV-1 (P=0.39)</b>	<b>BTV-1<sub>4S9</sub> &lt; rBTV-1 (P&lt;0.001)</b>
<b>BTV-1<sub>4S2S6S7</sub> &lt; rBTV-1 (P=0.01)</b>	<b>BTV-1<sub>4S2S6S7</sub> = rBTV-1 (P=0.16)</b>
<b>BTV-4<sub>1S2S6S7</sub> &lt; rBTV-1 (P&lt;0.001)</b>	<b>BTV-4<sub>1S2S6S7</sub> &lt; rBTV-1 (P=0.004)</b>
<b>BTV-1<sub>4S2</sub> = rBTV-4 (P=0.53)</b>	<b>BTV-1<sub>4S2</sub> &lt; rBTV-4 (P=0.001)</b>
<b>BTV-4<sub>1S3</sub> = rBTV-4 (P=0.50)</b>	<b>BTV-4<sub>1S3</sub> &lt; rBTV-4 (P&lt;0.001)</b>
<b>BTV-4<sub>1S9</sub> = rBTV-4 (P=0.13)</b>	<b>BTV-4<sub>1S9</sub> &lt; rBTV-4 (P=0.03)</b>
<b>BTV-4<sub>1S2S6S7</sub> &lt; rBTV-4 (P=0.02)</b>	<b>BTV-4<sub>1S2S6S7</sub> &lt; rBTV-4 (P&lt;0.001)</b>